Kabachnik-Fields synthesis of novel 2oxoindolin methyl phosphonate derivatives using CAN.

Presented by

Anna Pratima G. Nikalje^{1*}, Rekha I. Gajare¹, Shailee V. Tiwari¹, Julio A. Seijas²,
M. Pilar Vazquez-Tato²

¹Y.B. Chavan College of Pharmacy, Dr. Rafiq Zakaria Campus, Rauza Baug, Aurangabad, Maharashtra 431001, India;

²Departamento de Química Orgánica, Facultad de Ciencias, Universidad of Santiago De Compostela, Alfonso X el Sabio, Lugo 27002, Spain

* Correspondence: annapratimanikalje@gmail.com

One pot three componant Kabachnik-Fields synthesis of novel 2oxoindolin methyl phosphonates derivatives as anticancer agents.

2

Abstract

The work reports ultrasound promoted facile synthesis of novel ten α -aminophosphonate derivatives coupled with indole-2,3-dione moiety, namely diethyl(substituted phenyl/heteryl)(2-(2-oxoindolin 3ylidene)hydrazinyl)methylphosphonates derivatives **4(a-j)**. The derivatives **4(a-j)** were synthesized through one-pot three component Kabachnik-Fields reaction, by stirring at room temperature in presence of Cerric Ammonium Nitrate (CAN) as a catalyst, to give the final compounds in better yields and in shorter reaction time. Isatin, chemically known as *H*-indole-2,3-dione, and its derivatives possess a broad range of biological and pharmacological properties. Isatin is widely used as starting material for the synthesis of a broad range of heterocyclic compounds and as substrates for drug synthesis. The α -amino phosphonate derivatives constitute an important class of organophosphorus compounds on account of their versatile biological activity. The general low mammalian toxicity of these compounds made them attractive for use in agriculture and medicine. Considering the importance of the two pharmacophores, promoted us to club both the pharmacophores in a single molecule using green synthetic protocol. The structures of the ultrasound synthesized compounds were confirmed by spectral analysis like IR, ¹H NMR, ¹³C NMR, ³¹P NMR and MS.

Keywords: Kabachnik-Fields reaction; Cerric Ammonium Nitrate; Isatin; α-amino phosphonate.

CONTENTS



- Introduction
- Need and objective of Study
- Material and method
- Scheme of synthesis
- Experimental work
- Spectral analysis
- Result and discussion
- Conclusion
- References

INTRODUCTION

4

The basic method for the preparation of α -aminophosphonates, valuable synthetic equivalents and biologically active substrates, involves the condensation of a primary or secondary amine, a carbonyl compound (aldehyde or ketone) and dialkyl phosphit[1]. List of various catalysts used for synthesis of various types α -aminophosphonates and the time (minutes) required for synthesis of α -aminophosphonates are shown in **Table 1**.

Among the synthetic routes towards α -aminophosphonates two main pathways exist[2].

- 1) Three-component reactions (**Kabachnik-Fields reaction**): In this an aldehyde, an amine and di- or trialkyl phosphite are reacted in a one-pot set-up.
- 2) Pudovik reaction:

In this dialkyl phosphites are added to compound containing an imino-bond.

α-aminophosphonates are among the most studied bioactive organo phosphorus derivatives and have been used as enzyme inhibitors [3], inhibitors of serine hydrolase [4], peptide mimics [5], antiviral [6], antibacterial [7], antifungal [8], anticancer [9], anti-HIV [10], antibiotics [11], herbicidal [12] etc.

Indole possesses various medicinal properties like antibacterial, antifungal, anti-malarial, anticonvulsant and anti-inflammatory etc. [13].

INTRODUCTION

5

Isatin, chemically known as 1-*H*-indole-2,3-dione, and its derivatives possess a broad range of biological and pharmacological properties and are widely used as starting materials for the synthesis of a broad range of heterocyclic compounds and as substrates for drug synthesis. It is also used for the inhibition of pro-apoptotic jurkat T cells. In terms of its mode of action, isatin itself is proposed to inhibit cancer cell proliferation via interaction with extracellular signal-related protein kinases (ERKs), thereby promoting apoptosis. These compounds inhibit cancer cell proliferation and tumor growth via interaction with a variety of intracellular targets such as DNA, telomerase, tubulin, P glycoprotein, protein kinases and phosphatases [14]. Isatin-based hydrazones have been identified as inhibitors of the protein tyrosine phosphatase Shp2, which plays an important role in cell signaling, cell proliferation, differentiation and migration [15]. The marketed anticancer drug Sunitinib [16] and Oratinib contains 2-oxoindolin-3-ylidene moiety where as Ilmofosin and Edelfosin contains phosphonate moiety and a recently marketed anticancer drug, Toceranib phosphate [17] contains 2-oxoindol-3-ylidene as well as phosphonates moiety. Considering the biological importance of 2-oxoindolin-3-ylidene and α -aminophosphonates prompted us to synthesize coupled derivatives containing isatin based hydrazone and α -aminophosphonates with the hope to get novel hybrid derivatives with a better anticancer activity and minimized toxicity. The designing protocol for the target molecules is as shown in **Fig.1**.

INTRODUCTION

6

Most remarkable pathway to the synthesis of α -aminophosphonates is the Kabachnik-Fields reaction, the one pot three-component reaction of aromatic/heterocyclic aldehyde, amine and triethylphosphite, also known as Kabachnik–Fields reaction [18]. The novel trends in carrying out this reaction are connected with the application of (i) microwave irradiation itself or in combination with catalyst [19], (ii) ionic liquids as solvents [20], (iii) use of appropriate dehydrating agents [21] and, probably most important, (iv) the use of catalysts, was achieved by using various catalyst like ZrOCl₂.8H₂O [22], YbCl₃ [23], lanthanide triflates [24], Mg(ClO₄)₂ [25], LiClO₄ [26] etc.

Kabachnik–Fields reaction was promoted by using Cerric (IV) ammonium nitrate (CAN) as a catalyst because of its advantages like high solubility in organic solvent, ease of handling, and low toxicity [27]. List of various catalysts used for synthesis of various types α -aminophosphonates and the time (minutes) required for synthesis of α -aminophosphonates are shown in **Table 5.**

NEED OF THE STUDY

7

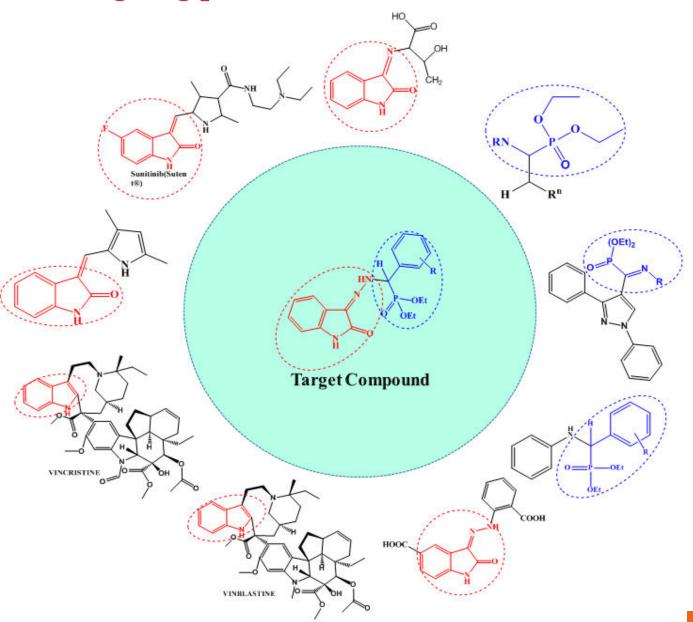
Green Chemistry Tools such as Ultrasound Synthesizer, multi component reactions, use of CAN catalyst, Molecular sieves, have become a promising alternative tools for various chemical processes due to their economty status like less time and electricity consumption, faster reaction and Cerric (IV) ammonium nitrate (CAN) as a catalyst because of its advantages like high solubility in organic solvent, ease of handling, and low toxicity respectively.

OBJECTIVE OF THE STUDY



- To design and synthesis of novel isatin coupled (α-aminophosphonate) derivatives with appropriate pharmacophore/suitable substituents like Imine linkage (azomethine linkage) i.e. Diethyl(substituted phenyl/heteryl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl phosphonates derivatives 4(a-j).
- To synthesize intermediates and final derivatives by Green Chemistry Tools such as Ultrasound Synthesizer, multi component reactions, use of CAN catalyst, Molecular sieves.
- Characterisation and structural confirmation of the synthesized intermediates & final α-aminophosphonate derivatives by analytical tests and spectral analysis such as TLC, IR, ¹HNMR, ¹³CNMR, ³¹PNMR and MASS.

Fig. 1: The Designing protocol



MATERIALS AND METHODS

10

- All the chemicals used for synthesis are of Research lab, Merck, Sigma, Qualigens make and Himedia.
- The reactions were carried out by conventional method and using Ultrasound synthesizer with solid probe (**Ultrasonic Processor VCX-500-220**) at 40°C.
- Melting points were determined in the open capillaries using melting point apparatus and are uncorrected.
- FTIR spectra were recorded by JASCO FTIR (PS-4000) using KBr powder technique, ¹H NMR and ¹³C NMR spectra of synthesized compounds were recorded on Bruker Avance II 400 NMR Spectrometer at 400 MHz Frequency in CDCl₃ and using TMS as internal standard (chemical shift δ in ppm), Mass spectra of some compounds were scanned on Water's Micromass Q-Tof system. ³¹P NMR of compounds were recorded at δ250 to δ250 in CDCl₃ and using Phosphoric acid (H₃PO₄) as an external standard (chemical shift δ in ppm)

Scheme of synthesis

Scheme 1

Structural Activity Relationship (SAR) diethyl(substituted phenyl/heteryl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl phosphonates derivatives. 4(a-j)

- Isatin is essential for biological and pharmacological activity activity.
- C=N linkage Schiff base in isatin is essential for biological and pharmacological activity.
- Phosphonate group is essential for biological and pharmacological but it should not be free.

EXPERIMENTAL WORK

Procedures:

Step I: Synthesis of 3-hydrazonoindolin-2-one (1) (Schiff base)²⁸

A) Conventional method ²⁸

A mixture of indole-2,3-dione (isatin) (1 mmol) and hydrazine hydrate (1 mmol) in 15 ml of methanol was refluxed for 3-4 hr in presence of molecular sieves. Microporous 3\AA molecular sieves are alumino silicate minerals with chemical composition of $^2/_3K_2O\cdot ^1/_3Na_2O\cdot Al_2O_3\cdot 2SiO_2\cdot ^9/_2H_2O.^{29}$ Since the 1990's, these molecular sieves have attracted considerable attention due to their potential use in catalysis, as they absorb water formed in the reaction and drive the reaction to completion. The separated crystals were filtered, washed with a little amount of methanol, dried and recrystallized with chloroform solvent(s), M.P. 284°C, Yield 82%.

B) Ultrasonication Method

Equimolar quantities of indole-2,3-dione (isatin) (1 mmol) and hydrazine hydrate (1mmol) in the presence of catalytic amount of glacial acetic acid in absolute ethanol (5 ml) was sonicated by keeping the reaction mixture in acoustic box containing Ultrasonic solid probe at 25-40°Cand at 25 amplitude for 15 -20 min. The completion of reaction was monitored by TLC. The reaction mixture was concentrated and cooled. The obtained solid was filtered and dried. The product was recrystallized from ethanol. 3-Hydrazonoindolin-2-one, $C_8H_7O_1N_3$, MW: 161.13. Yield: 95%; melting point: 279-284°C. The melting point was uncorrected.

Table 1: Difference in parameters for Conventional method and Ultrasound Method

Parameters	Conventional method	Ultrasound Method
Catalyst	glacial acetic acid, molecular sieves	glacial acetic acid
Solvent	absolute methanol	absolute ethanol
Temperature	40-50°C	25-40°C
Time	3-4 h	15-20 min.
yield	82%	95%

Characterization

I) Conventional Method (3-4 h)

Table 2: Physical constants data for 3-hydrazonoindolin-2-one (1)

Molecular	Molecular	% Yield(%)	M. P.(⁰ C)	R _f value
formula	weight (gm)			
C ₈ H ₇ O ₁ N ₃	161.13	82	279-284	0.70

II) Ultrasound Method (15-20 min.)

Table 3: Physical constants data for 3-hydrazonoindolin-2-one (1)

Molecular	Molecular	% Yield (%)	M. P.(⁰ C)	R _f value
formula	weight (gm)			
$C_8 H_7 O_1 N_3$	161.13	95	280-285	0.70

Solvent system chosen for R_f value determination was benzene: methanol (8:2).

Step II: diethyl(substituted phenyl/heteryl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methylphosphonates derivatives 4(a-j) [One pot Kabachnik–Fields reaction]

Equimolar quantity of 3-hydrazonoindolin-2-one (1) (1mmol), substituted aromatic aldehyde/heteryl aldehydes 2(a-j) (1mmol) and tri-ethyl phosphite (3) (1mmol) was stirred at room temperature in absolute ethanol, in presence of Cerric Ammonium Nitrate (CAN) as a catalyst. The progress of reaction was monitored by TLC. After completion of the reaction, the reaction mixture was cooled and poured in water, filtered and the solid obtained was dried and recrystallized with ethanol. The time required for completion of reaction varies from 70 min. to 90 min.

Table 4: Physical constant data diethyl(substituted phenyl/heteryl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methylphosphonates derivatives 4(a-j)

Code	-Ar	Molecular formula	Molecular weight (gm)	Time required (min)	% Yield	Melting point (⁰ C)
4a	Phenyl	$C_{19}H_{22}N_3O_4P$	387.37	75	90	195-196
4 b	p-chloro Phenyl	$C_{19}H_{21}CIN_3O_4P$	421.81	70	92	150-152
4c	p-hydroxy Phenyl	$C_{19}H_{21}FN_3O_4P$	405.36	75	95	176-180
4d	p-methoxy Phenyl	$C_{20}H_{24}N_3O_5P$	417.40	85	89	179-180
4e	3,4-dimethoxy Phenyl	$C_{21}H_{26}N_3O_6P$	447.42	90	90	189-190
4f	p-fluoro Phenyl	$C_{19}H_{22}N_3O_5P$	403.37	80	88	140-142
4 g	4-hyroxy-3-methoxy Phenyl	$C_{20}H_{24}N_3O_6P$	433.39	75	94	112-114
4h	4-hyroxy-3-ethoxy Phenyl	$C_{21}H_{26}N_3O_6P$	447.44	80	92	160-162
4i	Thiophen-2-yl	$C_{17}H_{20}N_3O_4PS$	393.40	80	87	178-180
4j	Furan-2-yl	$C_{17}H_{20}N_3O_5P$	377.33	80	84	176-178

Solvent system chosen for R_f value determination was benzene: methanol (8:2).

Table 5: Catalyst used for the synthesis of α -aminophosphonate (4a-j) as compared with other reported time and catalyst used.

Entry	Catalyst used	Time(minutes)	References
1	Bismuth salt (10 mol%)	120-360	30
2	Yb (OTf) ₃ (10mol%)	270-2160	31
3	2 mol % HfCl ₄	300-2880	32
4	CBr ₄ (5 mol%)82	180	33
5	Oxalyl chloride (1.5 mmol)	360	34
6	AlCl ₃ (10 mol%)80	570	35
7	CAN(Reflux)	30	36
8	CAN(RT)	180-210	37
9	CAN	75-90	Present work

I R SPECTROSCOPY Spectrum no.1:

$Die thyl\ ((4-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl) methyl) phosphonate$

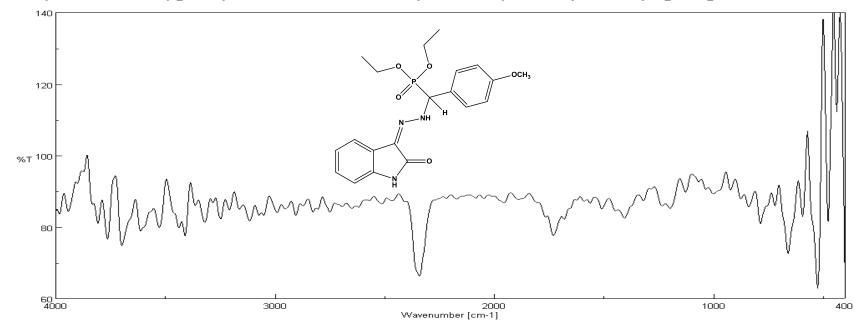


Table 6: IR interpretation of diethyl ((4-methoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Functional group	Wavelength (cm ⁻¹)
1.	N-H stretching	3350.41
2.	C-H stretching of aromatics	2970.76
3.	CH stretching of alkyl	2800
4.	C=N Streching	2200-2300
5.	C=O stretching of Amide	1610.26
6.	C-H bending of –CH ₂ -	1456.58
7.	C-N Streching	1230-1030

I R SPECTROSCOPY Spectrum no.2:

$\underline{Diethyl\ ((\textbf{3,4-dimethoxyphenyl})(\textbf{2-(2-oxoindolin-3ylidene}) hydrazinyl) methyl) phosphonate}$

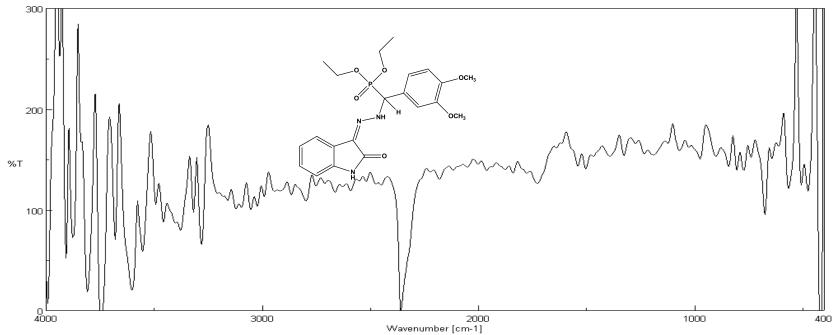


Table 7: IR interpretation of diethyl ((3,4-dimethoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Functional group	Wavelength (cm ⁻¹)
1.	N-H stretching	3352
3.	C-H stretching of aromatics	3000.76
4.	CH stretching of alkyl	2800.53
5.	C=N Streching	2350
6.	C=O stretching of Amide	1650.23
7.	C-N Streching	1232-1040
8.	C-H bending of –CH ₂ -	1456.58

IR SPECTROSCOPY

Spectrum no.3:

<u>Diethyl ((furfuryl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate.</u>

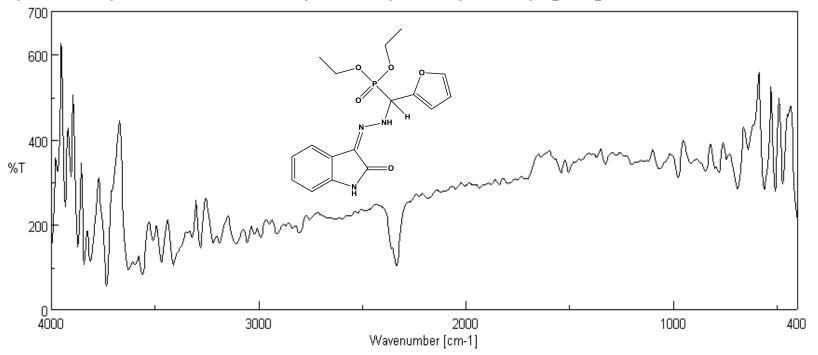


Table 8 IR interpretation of diethyl ((furfuryl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Functional group	Wavelength (cm ⁻¹)
1.	N-H stretching	3350.11
2.	C-H stretching of aromatics	2970
3.	CH stretching of alkyl	2890.02
4.	C=N Streching	2330
5.	C=O stretching of Amide	1710.66
6.	C-H bending of –CH ₂ -	1416.88
7.	C-N Streching	1230-1130
8.	-O- Streching	1050.52

IR SPECTROSCOPY

Spectrum no.4:

$\underline{Diethyl\ ((phenyl)(2\text{-}(2\text{-}oxoindolin-3ylidene)hydrazinyl)methyl)phosphonate}$

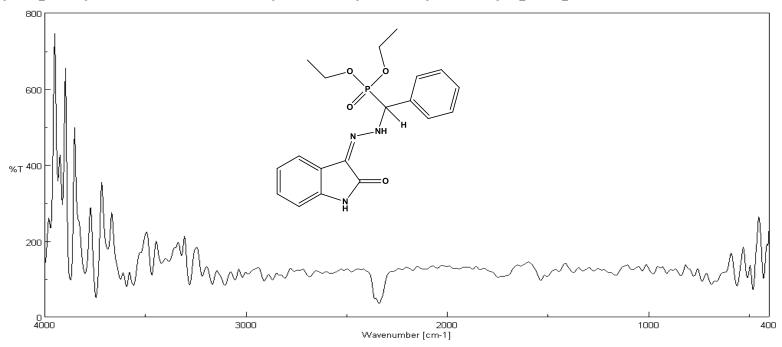


Table 9: IR interpretation of diethyl ((phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Functional group	Wavelength (cm ⁻¹)
1.	N-H stretching	3340.21
2.	C-H stretching of aromatics	2960.76
3.	CH stretching of alkyl	2870
4.	C=N Streching	2200-2350
5.	C=O stretching of Amide	1610.66
6.	C-H bending of –CH ₂ -	1466.55
7.	C-N Streching	1250-1020

MASS SPECTROSCOPY Spectrum no.5:

Diethyl ((4-chlorophenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate (4b)

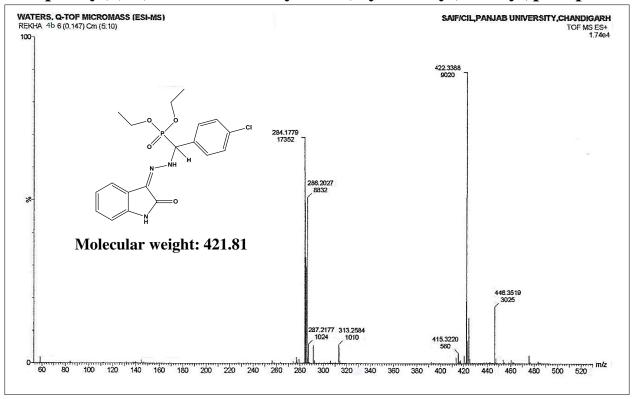


Table 10: Mass interpretation of diethyl ((4-chloro phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Fragmentation	m/e
1.	M+1	422.33(base peak)
2.	M+2	423
3.	$M-C_4H_{11}O_3P$	284.17

MASS SPECTROSCOPY

Spectrum no.6:

Diethyl ((4-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate (4d)

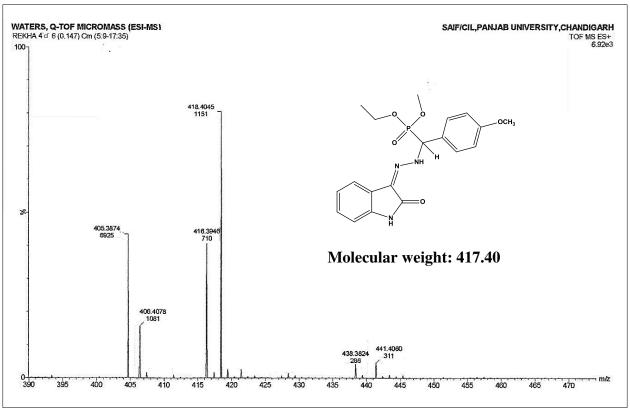


Table 11: Mass interpretation of diethyl ((4-methoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Fragmentation	m/e
1.	M+1	418.40(base peak)
2.	$M-CH_3$	405.38

MASS SPECTROSCOPY Spectrum no.7:

Diethyl ((4-hydroxy-3-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate (4g)

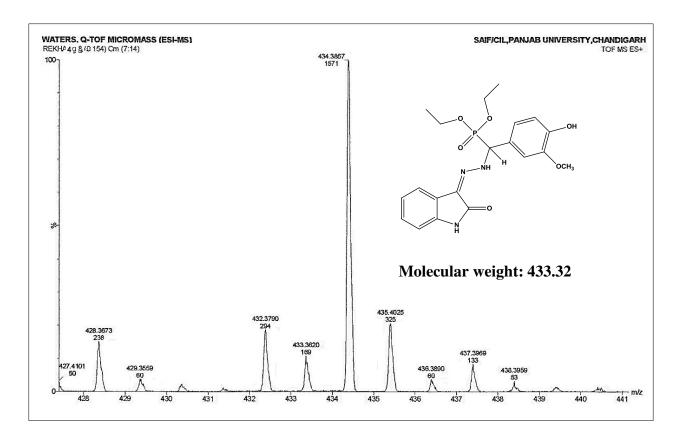


Table 12: Mass interpretation of diethyl ((4-hydroxy-3-methoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	Fragmentation	m/e
1.	M+1	334.31(base peak)

¹H NMR SPECTROSCOPY Spectrum no.8

Diethyl ((4-chlorophenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate(4b)

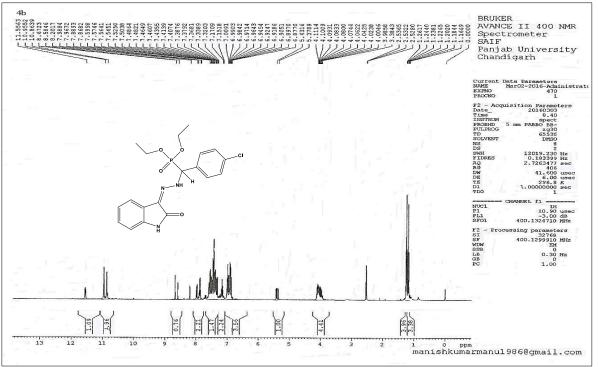


Table 13: ¹H NMR interpretation of diethyl ((4-chloro phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	δ Values (ppm)	Multiplicity	No. of proton	Group
1	1.1-1.2	Т	6H	-CH ₃
2	4.41	Q	4H	-CH ₂
3	3.4-4	D	1H	-СН
4	7-7.19	M	8H	Aromatic protons
5	8.6	S	1H	Aliphatic-NH
6	11.55	S	1H	Indole-NH

¹H NMR SPECTROSCOPY Spectrum no.9:

Diethyl ((4-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate (4d)

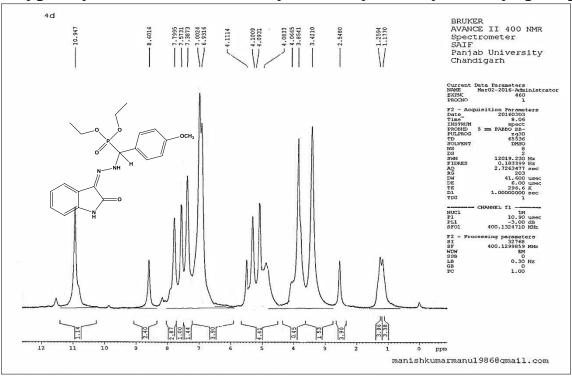


Table 14: ¹H NMR interpretation of diethyl ((4-methoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	δ Values (ppm)	Multiplicity	No. of proton	Group
1	1.1-1.2	Т	6Н	-CH ₃
2	2.5	S	3Н	-OCH ₃
3	3.42-3.85	D	1H	-СН
4	4.08-4.11	Q	4H	CH_2
5	6.9-8.40	M	8H	Aromatic protons
6	8.6	S	1H	Aliphatic-NH
7	10.94	S	1H	Indole-NH

¹H NMR SPECTROSCOPY Spectrum no.10:

Diethyl ((4-hydroxy-3-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate.(4g)

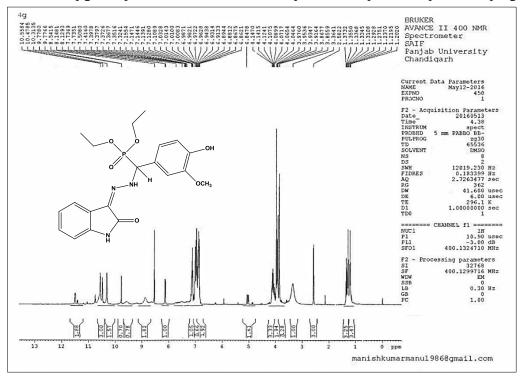


Table 15: ¹H NMR interpretation of diethyl ((4-hydroxy-3-methoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	δ Values (ppm)	Multiplicity	No. of proton	Group
1	1.2-1.3	Т	6Н	-CH ₃
2	2.54	S	3Н	-OCH ₃
3	3.36-3.86	D	1H	-CH
4	3.86-4.24	Q	4H	-CH ₂
5	5.14	S	1H	-ОН
6	6.84-7.73	M	7H	Aromatic protons
7	8.14	S	1H	Aliphatic-NH
8	10.31	S	1H	Indole-NH

¹³C NMR SPECTROSCOPY Spectrum no.11:

Diethyl ((4-methoxyphenyl)(2-(2-oxoindolin-3ylidene)hydrazinyl)methyl)phosphonate (4d)

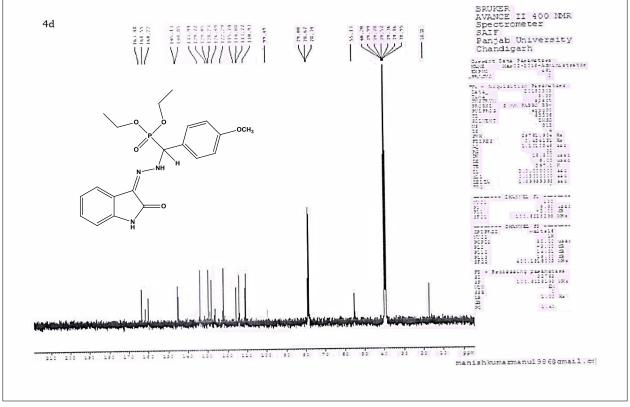


Table 16: ¹³C NMR interpretation of diethyl ((4-methoxy phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	δ Values (ppm)	Carbon group
1	16	-CH ₃
2	40	-OCH ₃
3	57	-CH ₂
5	100	-CH
6	134	Aromatic
7	145	-C-OCH ₃
9	164	C=O

¹³C NMR SPECTROSCOPY Spectrum no.12:

Diethyl ((4-chlorophenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate (4b)

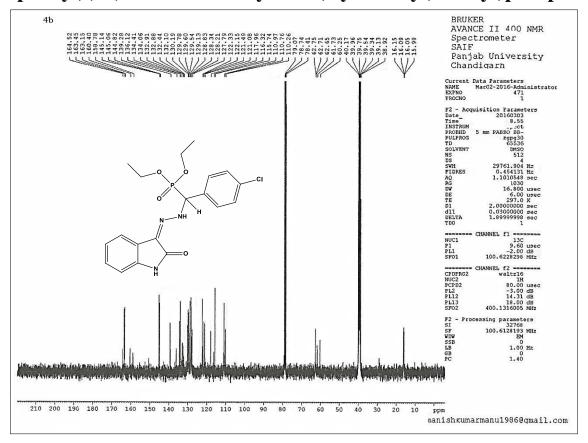


Table 17: ¹³C NMR interpretation of diethyl ((4-chloro phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	δ Values (ppm)	Carbon group
1	16	-CH ₃
2	62	-CH ₂
3	80	-СН
4	110-134	Aromatic
5	164	C=O

¹³C NMR SPECTROSCOPY

Spectrum no.13:

Diethyl ((4-hydroxy-3-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

(4g)

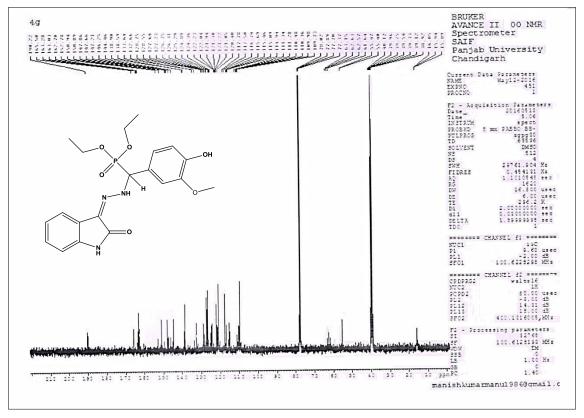


Table 18: ¹³C NMR interpretation of diethyl ((4-hydroxy-3-methoxyphenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate

Sr. No.	δ Values (ppm)	Carbon group
1	16	-CH ₃
2	40	-OCH ₃
3	62	-CH ₂
4	78	-CH
5	109-165	Aromatic CH
6	190	C=O

³¹P NMR SPECTROSCOPY

Spectrum no.14:

diethyl ((4-chlorophenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate.(4b)

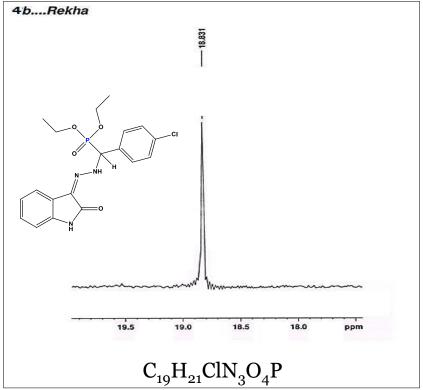


Table 19: ³¹P NMR interpretation of diethyl ((4-choro phenyl)(2-(2-oxoindolin-3-ylidene)hydrazinyl)methyl)phosphonate.

Sr. No.	Obtained Values	Theoretical value
	(ppm)	(ppm)
1	18.831	15-25

RESULT AND DISCUSSION

Diethyl (substituted phenyl/heteryl)(2-(2-oxoindolin-3ylidene)hydrazinyl)methyl phosphonates derivatives **4(a-j)** were synthesized by Green protocol as outlined in **Scheme 1**. 3-hydrazonoindolin-2-one (1) was synthesized by reacting indole-2,3-dione (isatin) (1mmol) with hydrazine hydrate (1mmol) in the presence of glacial acetic acid as a catalyst by conventional method in methanol using molecular sieves and by ultrasonication method by replacing methanol with ethanol. Ultrasound method is better than the conventional method because, methanol being toxic solvent is replaced by benign solvent ethanol. The amount of solvent required is also less than that required for conventional method. Ultrasound assisted method gives better yield in 15-20 minutes against 3-4 hrs required for conventional method. α -Aminophosphonate derivatives 4(a-j) were synthesized by reacting 3hydrazonoindolin-2-one (1), substituted/heteryl aldehydes 2(a-j) and triethylphosphite (3) via one pot synthetic step in presence of CAN as a catalyst. CAN activates the imine formation due to which addition of phosphite is facilitated to give a phosphonium intermediate. This phosphonium intermediate undergoes reaction with water to give the title compounds. CAN catalyst being water soluble can be easily removed after completion of reaction. The synthesized compounds were characterized and confirmed by FTIR, ¹H NMR, ¹³C NMR, ³¹P NMR, MS and elemental analyses. The purity of the synthesized compounds was checked by TLC and melting points were determined in open capillary tubes and are uncorrected. Physical constant data for diethyl (substituted phenyl/hetery)(2-(2-oxoindolin-3-ylidene)hydrazinyl) methyl phosphonates **4(a-j)** is shown in Table 4.

CONCLUSION

- ➤ **Ultrasound synthesizer** have become a promising alternative green tool for various chemical reactions due to their economical status like less time and electricity consumption by faster reaction.
- ➤ Intermediates (1) were synthesized by Green protocol such as by using ultrasound synthesizer, gives better yield in 15-20 minutes while conventional method requires 3-4 hrs. Final compounds 4(a-j) were synthesized by one pot synthetic step in presence of CAN as a catalyst.
- Novel diethyl (substituted phenyl/heteryl)(2-(2-oxoindolin-3ylidene)hydrazinyl)methylphosphonate derivatives **4(a-j)** were synthesized at room temperature in facile one pot reaction using CAN as a catalyst, gives faster reaction at room temperature, CAN catalyst being water soluble can be easily removed after completion of reaction
- > The compounds were characterized by TLC, IR, NMR, and Mass spectrometry.

REFERENCES

- 34)
- 1) M.I. Kabachnik, T.Y. Medved, New synthesis of aminophosphonic acids. *Dokl. Akad. Nauk SSSR* **1952**, *83*, 689–692.
- 2) Christian V. Stevens *et.al.*, Straightforward continuous synthesis of α-aminophosphonates under microreactor conditions, General Papers ARKIVOC 2006 (i) 31-45.
- 3) C. Li, B. Song, K. Yan, G. Xu, D. Hu, S. Yang, L. Jin, W. Xue, P. Lu, One Pot Synthesis of α-Aminophosphonates Containing Bromo and 3,4,5-Trimethoxybenzyl Groups under Solvent-free Conditions, Molecules, 12 (2007), 163-172.
- 4) G.S. Prasad, J.R. Krishna, M. Manjunath, O.V.S. Reddy, M. Krishnaiah, C.S. Reddy, V.G. Puranikd, Synthesis, NMR, X-ray crystallography and bioactivity of some α-aminophosphonates, ARKIVOC, 13 (2007), 133-141.
- 5) X. Rao, Z. Song, L. He, Synthesis and antitumor activity of novel α-aminophosphonates from diterpenicdehydroabietylamine, Heteroatom Chem. 19 (2008), 512-516.
- 6) E.D. Naydenova, P.T. Todorov, P.I. Mateeva, R.N. Zamfirova, N.D. Pavlov, S.B. Todorov, Synthesis and biological activity of novel small peptides with aminophosphonates moiety as NOP receptor ligands, Amino Acids, 39 (2010), 1537-1543.
- 7) <u>L.</u> Tusek-bozic, M. Juribasic, P. Traldi, V. Scarcia, A. Furlani, Synthesis, characterization and antitumor activity of palladium(II) complexes of monoethyl 8-quinolylmethylphosphonate, Polyhedron, 27 (2008), 1317–1328.
- 8) B. Wang, Z.W. Miao, J. Wang, R.Y. Chen, X.D. Zhang, Synthesis and biological evaluation of novel naphthoquinone fused cyclic aminoalkylphosphonates and aminoalkylphosphonic monoester, Amino Acids, 35 (2008), 463-468.
- 9) Z. Rezaei, H. Firouzabadi, N. Iranpoor, A. Ghaderi, M.R. Jafari, A.A. Jafari, H.R. Zare, Design and one-pot synthesis of alpha-aminophosphonates and bis(alpha-aminophosphonates) by iron(III) chloride and cytotoxic activity, See comment in PubMed Commons below Eur. J. Med. Chem., 44 (2009), 4266-4275.
- 10) N. Onita, I. Sisu, M. Penescu, V.L. Purcarea, L. Kurunczi, Synthesis, characterization and biological activity of some α-aminophosphonates, Farmacia, 58 (2010), 531-545.
- 11) X. Zhang, Y. Qu, X. Fan, C. Bores, D. Feng, G. Andrei, R. Snoeck, E. De Clercq, P.M. Loiseau, Solvent-free synthesis of pyrimidine nucleoside-aminophosphonate hybrids and their biological activity evaluation, Nucleosides Nucleotides Nucleic Acids, 29 (2010), 616-627.
- 12) J. Liu, S. Yang, X. Li, H. Fan, P. Bhadury, W. Xu, J. Wu, Z. Wang, Synthesis and antiviral bioactivity of chiral thioureas containing leucine and phosphonate moieties, Molecules, 15 (2010), 5112-5123.
- 13) S. Biswal, U. Sahoo, S. Sethy, H.K.S. Kumar, M. Banerjee, Indole: The Molecule of Diverse Biological Activities, Asian J. Pharm. Clin. Res., 5 (2012), 1-6.
- 14) K.L. Vine, L. Matesic, J.M. Locke, D. Skropeta, Recent Highlights in the Development of Isatin-Based Anticancer Agents, University of Wollongong Research Online Faculty of Science, Medicine and Health, 59 (2013) 254-312.

REFERENCES

- H.R. Lawrence, R. Pireddu, L. Chen, Y. Luo, S.S. Sung, A.M. Szymanski, M.L.R. Yip, W. C. Guida, S.M. Sebti, J. Wu, N.J. Lawrence, Inhibitors of Src homology-2 domain containing protein tyrosine phosphatase-2 (Shp2) based on oxindole scaffolds, J. Med. Chem. 51 (2008), 4948-4956.
- K.G. Hui, S. Bostjan, J.J. Knox, Sunitinib in solid tumors, Expert Opin Investig Drugs, 18 (2009), 821–834.
- M.F. Vancy, merritt, J.A. White, S.A. Marsh, C.W. locuson, Distribution, metabolism, and excretion of toceranib phosphate (PalladiaTM, SU11654), a novel tyrosine kinase inhibitor, in dogs, 33 (2010), 154-161
- E.D. Matveeva, N.S. Catalytic Kabachnik-Fields reaction: new horizons for old reaction ARKIVOC, 1 (2008), 1-17.
- 19) B. Kaboudin, R. Nazari, *Tetrahedron Lett.* 42 (2001), 8211.
- 20) J. S. Yadav, B. Reddy, P. Sreedhar, *Green Chem.* (4) 2002, 436.
- 21) C. Qian, T. Huang, J. Org. Chem. (63) 1998, 4125.
- 22) (a)J.S. Yadav, B.V.S. Reddy, K.S. Raj, K.B. Reddy, A.R. Prasad, Synthesis (2001) 2277–2280; (b) S.Bhagat, A.K. Chakraborti, J. Org. Chem., 73 (2008) 6029-6032.
- F. Xu, Y. Luo, J. Wu, Q. Shen, H. Chen, Facile one-pot synthesis of α --aminophosphonates using lanthanide chloride as catalyst, Heteroatom Chem. 17 (2006), 389–392.
- C. Qian, T.J. Huang, One-Pot Synthesis of α-Amino Phosphonates from Aldehydes Using Lanthanide Triflate as a Catalyst, J. Org. Chem., 63 (1998), 4125-4128.
- J. Wu, W. Sun, H.G. Xia, X. Sun, A facile and highly efficient route to α-amino phosphonates via three-component reactions catalyzed by $Mg(ClO_4)_2$ or molecular iodine, Org. Biomol. Chem., 4(2006), 1663-1666.
- 26) N. Azizi, F. Rajabi, M.R. Saidi, Tetrahedron Lett. 45 (2004), 9233-9235.
- J.N. Sangshetti, N.D. Kokare, S.A. Kotharkar, D.B. Shinde, Ceric ammonium nitrate catalyzed three component one-pot efficient synthesis of 2,4,5-triaryl-1Himidazoles, J. Chem. Sci. 120 (2008), 463-467.
- K. swathi *et.al.*, "synthesis and anti-inflammatory activity of a novel series of isatin hydrazone & isatin thiosemicarbazone derivatives", world journal of pharmacy and pharmaceutical sciences, Volume 3, Issue 2, 2070-2078.
- A. Corma, 'From Microporous to mesoporous molecular sieve materials and their use in Catalysis', *Chem. Rev.* **1997**, *97*, 2373-2420.
- Bimal K. Banik *et. al.*, A Highly Efficient Bismuth Salts-Catalyzed Route for the Synthesis of α-Aminophosphonates, Molecules, 2010, 15, 8205-8213.

REFERENCES



- Doo Ok Jang *et. al.*, Efficient one-pot synthesis of a-aminophosphonates from aldehydes and ketones catalyzed by ytterbium(III) triflate, Tetrahedron Letters 53 (2012) 3897–3899.
- Shan-Shan Gong *et. al.*, Highly efficient synthesis of a-aminophosphonates catalyzed by hafnium(IV) chloride, Tetrahedron Letters 57 (2016) 1782–1785.
- Sidhanath V. Bhosale *et. al.*, Synthesis and Biological Evaluation of Novel α-Aminophosphonate Derivatives Possessing Thiazole-Piperidine Skeleton as Cytotoxic Agents, Chemistry & Biology Interface, 2014, 4, 1, 48-57.
- Ye Zhang *et. al.*, Synthesis and antitumor activities of novel thiourea a-aminophosphonates from dehydroabietic acid, European Journal of Medicinal Chemistry 69 (2013) 508-520.
- 35) M.J. Bloemink et. al., European Journal of Inorganic Chemistry ,10 (1999) 1655–1657.
- K. Ravinder, A. Vijender Reddy, P. Krishnaiah, G. Venkataramana, V. L. Niranjan Reddy, Y. Venkateswarlu, CAN Catalyzed One-Pot Synthesis of α-Amino Phosphonates from Carbonyl Compounds, Synthetic Comm., 34(2004)1677–1683.
- M. Kasthuraiah, K. A. Kumar, C. S. Reddy, C. D. Reddy, Syntheses, Spectral Property, and Antimicrobial Activities of 6-Amino Dibenzo[d,f][1,3,2]Dioxaphosphepin 6-Oxides, Heteroatom Chemistry, 18(2007) 223-229.

THANK YOU