A Novel Non-Stochastic Quadratic Fingerprints-based Approach for the “in silico” Discovery of New Antitrypanosomal Compounds.

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Abstract

A Non-Stochastic Quadratic Fingerprints-based approach is introduced to classify and design, in a rational way, new antitrypanosomal compounds. A data set of 153 organic-chemicals; 62 with antitrypanosomal activity and 91 having other clinical uses, was processed by a k-means cluster analysis in order to design training and predicting data sets. Afterwards, a linear classification function was derived allowing the discrimination between active and inactive compounds. The model classifies correctly more than 93% of chemicals in both training and external prediction groups. The predictability of this discriminant function was also assessed by a leave-group-out experiment, in which 10% of the compounds were removed at random at each time and their activity a posteriori predicted. Also a comparison with models generated using four well-known families of 2D molecular descriptors was carried out. As an experiment of virtual lead generation, the present TOMOCOMD approach was finally satisfactorily applied on the virtual evaluation of ten already synthesized compounds. The in vitro antitrypanosomal activity of this series against epimastigotes forms of T. cruzi was assayed. The model was able to predict correctly the behaviour of these compounds in 90% of the cases.

Keywords: Antitrypanosomal Compounds, Chagas’Disease, LDA-based-QSAR-Model, Non-Stochastic Quadratic Indices, QSAR, TOMOCOMD Software.
1. Introduction

Once an almost exclusively rural disease in Latin America, Chagas’ disease, is now undergoing a change in its epidemiological profile due to rising levels of urbanization and migration. Latest data from the WHO indicates that over 24 million peoples are infected or at least serologically positive for Trypanosoma cruzi, which is the causative agent of such infection. This quantity roughly represents 8% of the total Latin America population. Another factor, blood transfusion is considered the second most frequent route of transmission in endemic countries, given parasites may survive in whole blood stored for more than 21 days at 4º C and detection techniques are not always strictly applied.¹

Medication for Chagas’ disease is usually effective when it is given during the acute stage of infection. No medication has been proven to be effective once the disease has progressed to later stages. Moreover, synthetic drugs such as nifurtimox and benznidazole have severe side effects, including cardiac and/or renal toxicity. This explains the need for discovering new effective chemotherapeutic and chemoprophylactic agents against T. cruzi.²,³ In this sense, medicinal chemists are called to find new effective drugs in a fast and non-expensive way. In the last decades computer-aided drug design approaches have emerged as promising tools to be used to solve this problematic.⁴⁻¹⁰ With the use of such design strategies it is possible the handling and screening of large databases in order to find reduced sets of potential new drug candidates.¹¹,¹² Thus, the development of computational approaches based on discrimination functions plays an important role, allowing the identification from large chemical libraries of structural subsystems responsible for a property or biological activity, and in this way, the classification of active compounds from inactive ones.

In this context, our research group has recently developed a novel scheme to generate molecular fingerprints based on the application of discrete mathematics and linear algebra theory, which permits to perform rational in silico molecular design (selection/identification) and QSAR/QSPR studies. Known as TOMOCOMD (acronym of TOpological MOlecular COMputer Design),¹³⁻¹⁶ this approach has been successfully applied to the prediction of several physical, physicochemical, chemical, pharmacokinetical as well as biological properties.¹⁷⁻¹⁹ It was, for instance, successfully used on the virtual screening of novel antihelminthic compounds, which were then synthesized and in vivo evaluated on Fasciola hepatica.²⁰,²¹ Other studies for the rational discovery of novel paramphistomicides,²² antimalarial²³ and antibacterial²⁴ compounds were also conducted with the TOMOCOMD approach. This method has been extended to consider three-dimensional features of
small/medium-sized molecules upon the base of the application of a trigonometric 3D-chirality correction factor.\textsuperscript{25}

In the present study, \textit{TOMOCOMD} strategy is used to find a classification model which allows discriminating antitrypanosomal compounds from inactive ones. It is also an objective of the present work to assess the model robustness and predictive power by using external and internal cross-validation techniques. The \textit{in silico} evaluation of ten new heterocyclic compounds is finally performed, and their \textit{in vitro} antitrypanosomal activity against epimastigotes forms of \textit{T. cruzi} investigated. The results of the current study are presented as starting point for the development of new non-expensive antitrypanosomals. The general procedure is depicted in the following figure.

![Graphical abstract](image)

\textbf{Figure 1. Graphical abstract}

\section*{2. Results and Discussion}

\textbf{Computing Non-Stochastic Quadratic Molecular Fingerprints.} In order to obtain quantitative structure-property or structure-activity relationships (abbreviated QSPR and QSAR, respectively), it is necessary to “convert” the molecular structures into numbers that could be later statistically processed; it means, a structural parameterization is required. By means of the computation of molecular descriptors, such problematic is overcome.\textsuperscript{26} In the last decades a great number of molecular fingerprints have been presented in the literature.\textsuperscript{27,28} Atomic, atom-type and total non-stochastic quadratic indices have shown a great ability to encode chemical information, which can be used for the development of QSARs. The theoretical scaffold of this \textit{TOMOCOMD}'s molecular fingerprints has been presented in details in previous papers.\textsuperscript{14-25} Here just a short overview will be given.

Atomic, atom-type and total molecular quadratic molecular indices have been defined in analogy to the quadratic mathematical maps.\textsuperscript{14,16} After constructing the molecular pseudograph’s atom adjacency matrix $M(G)$ and the molecular vector ($X$), whose components $x_1,\ldots,x_n$ are numeric values or weights (atom-labels or atom-properties) for the vertices of the
pseudograph, $k$th total quadratic indices, $q_k(x)$, can be computed for a given molecule composed of $n$ atoms as shown in Eq. 1,

$$q_k(x) = \sum_{i=1}^{n} \sum_{j=1}^{n} k a_{ij} x_i x_j$$

(1)

where, $k a_{ij}$ are the elements of the $k$th power of the symmetrical square matrix $M(G)$ of the molecular pseudograph (G), and are defined as follows:

$$k a_{ij} = P_{ij} \text{ if } i \neq j \text{ and } \exists e_k \in E(G)$$

$$= L_{ii} \text{ if } i = j$$

$$= 0 \text{ otherwise}$$

(2)

$E(G)$ represents the set of edges (bonds) of $G$. $P_{ij}$ is the number of edges between vertices (atoms) $v_i$ and $v_j$, and $L_{ii}$ is the number of loops in $v_i$.

Equation (1) for $q_k(x)$ can also be written as a single matrix equation:

$$q_k(x) = X^t M^k X$$

(3)

where $X$ is a column vector (a $n \times 1$ matrix), $X^t$ the transpose of $X$ (a $1 \times n$ matrix) and $M^k$ the $k$th power of the matrix $M$ of the molecular pseudograph $G$.

In a similar way, local-fragment (atomic and atom-type) formalisms can be developed. The local quadratic indices, $q_{kL}(x)^{14,16}$ for a fragment containing $m$ atoms can be computed as follows:

$$q_{kL}(x) = \sum_{i=1}^{m} \sum_{j=1}^{m} k a_{ijL} x_i x_j$$

(4)

where $k a_{ijL}$ is the element of the row “$i$” and column “$j$” of the matrix $M^k_{L}$ and is defined as follows:

$$k a_{ijL} = k a_{ij} \text{ if } \text{both } v_i \text{ and } v_j \text{ are atoms contained within the molecular fragment}$$

$$= \frac{1}{2} k a_{ij} \text{ if } v_i \text{ or } v_j \text{ is an atom contained within the molecular fragment but not both}$$

$$= 0 \text{ otherwise}$$

(5)

These local analogues can also be expressed in matrix form by the expression:

$$q_{kL}(x) = X^t M^k_{L} X$$

(6)

For every partition of a molecule into $Z$ molecular fragment there will be $Z$ local molecular fragment matrices $M^k_{L}$. The $k$th power of matrix $M$ is exactly the sum of the $k$th power of the local $Z$ matrices and in this way, the total quadratic indices are the sum of the quadratic indices of the $Z$ molecular fragments:
$$q_k(x) = \sum_{L=1}^{Z} q_{kL}(x)$$ (7)

Atom and atom-type quadratic fingerprints are specific cases of local quadratic indices. In the atom-type quadratic indices formalism, each atom in the molecule is classified into an atom-type (fragment), such as heteroatoms, hydrogen bonding (H-bonding) to heteroatoms (O, N and S), halogen atoms, etc. For all data sets, considering those with a common molecular scaffold as well as those with diverse ones, the $k^{th}$ atom-type quadratic indices have demonstrated to encode important structural information.

In the current work, the $k^{th}$ total quadratic indices [$q_k(x)$ and $q_k^H(x)$] and the $k^{th}$ local ones (atom-type = heteroatoms: S, N, O) [$q_{kL}(x_E)$ and $q_{kL}^H(x_E)$] not considering and considering H-atoms respectively were computed.

**Training and test sets design.** In order to obtain mathematical expressions capable of discriminating between active and inactive compounds, the chemical information contained in a great number of compounds with and without the desired biological activity must be statistically processed. Taking into account that the most critical aspect on the construction of a training data set is the molecular diversity of the included compounds, we selected a group of 153 organic chemicals having as much structural variability as possible. The antitrypanosomals considered on this study are representatives of families with diverse structural patterns.\textsuperscript{29-38} Figure 2 shows the whole active set collected from the literature for this work.

**Figure 2.** Structures of active compounds in training and test groups.

\begin{tabular}{|c|c|c|c|c|c|c|c|c|c|c|}
\hline
& 1 & 2 & 3 & 4 & 7 & 41 & 42 & 43 & 44 & 45 \\
\hline
X & H & H & Cl & Cl & Cl & Cl & Cl & Cl & Cl & Cl \\
\hline
Y & H & HN$_3$ & HN$_3$ & HN$_4$ & S & N & H & HN$_2$ & HN$_4$ & S$_4$N$_4$ \\
\hline
Z & Cl & OMe & OMe & H & OMe & H & OMe & H & OMe & OMe \\
\hline
\end{tabular}
The selected inactive group included antivirals, sedative/hypnotics, diuretics, anticonvulsivants, hemostatics, oral hypoglycemics, antihypertensives, antihelminthics, anticancer compounds as well as some other kinds of drugs, guaranteeing at the same time a great structural variability.\(^{39}\)

In order to split the whole group into two data sets (training and predicting ones), two \(k\)-MCA\(^ {40,41}\) were performed for antitrypanosomal and inactive compounds respectively. In this sense, a partition of either active or inactive series of chemicals in several statistically

\[
\begin{array}{cccc}
34 & 35 & 58 \\
X & Br & CF_3 & Cl \\
Y & H & H & Cl \\
Z & H & CF_2 & H \\
\end{array}
\]

\[
\begin{array}{cccc}
36 & 37 & 59 & 60 \\
X & Me & Me & H \\
Y & Cl & CF_3 & Br \\
Z & Cl & H & H \\
\end{array}
\]

\[
\begin{array}{cccc}
38 & 39 & 40 & 62 \\
X & H & H & H \\
Y & OMe & H & NO_2 \\
\end{array}
\]

\[
\begin{array}{cccc}
34 & 35 & 58 \\
X & Br & CF_3 & Cl \\
Y & H & H & Cl \\
Z & H & CF_2 & H \\
\end{array}
\]

\[
\begin{array}{cccc}
36 & 37 & 59 & 60 \\
X & Me & Me & H \\
Y & Cl & CF_3 & Br \\
Z & Cl & H & H \\
\end{array}
\]

\[
\begin{array}{cccc}
38 & 39 & 40 & 62 \\
X & H & H & H \\
Y & OMe & H & NO_2 \\
\end{array}
\]
representative classes of compounds is performed. This process ensures that any chemical class identified by the \( k \)-MCA will be represented in both, training and test sets. A first \( k \)-MCA (I) split antitrypanosomals in 6 clusters with 12, 3, 2, 13, 13 and 19 members. The inactive compound series was also partitioned by a second \( k \)-MCA (II) into 6 clusters with 17, 12, 14, 19, 18 and 11 compounds in each one. Afterwards, the selection of the training and prediction sets was performed taking compounds belonging to each cluster at random. From these 153 chemicals, 101 were chosen to form the training set, being 40 of them actives and 61 inactive ones. The remaining group composed of 20 antitrypanosomals and 32 compounds with other different biological properties were prepared as test sets for the external model validation process. These 52 compounds were not used in the development of the classification model. Figure 3 graphically illustrates the above-described procedure.

**Figure 3.** Training and test data sets design throughout \( k \)-means cluster analysis.

An inspection of the standard deviation between and within each cluster, the Fisher ratios and the \( p \)-levels of significance for each variable, permits us to ensure that the data partition into the respective clusters can be considered as a statistically acceptable process. The \( k \)th total and atom-type non-stochastic quadratic indices were used in this analysis, with all variables showing \( p \)-levels <0.05 for the Fisher test. The main results are depicted in Table 1.
Table 1. Main results of the \(k\)-means cluster analysis, for antitrypanosomal and inactive compounds.

<table>
<thead>
<tr>
<th>Analysis of Variance</th>
<th>Total and atom-type quadratic indices</th>
<th>Between SS(^a)</th>
<th>Within SS(^b)</th>
<th>Fisher ratio (F)</th>
<th>(p)-level(^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Antitrypanosomal agents clusters (k-MCA I)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(q_6(x))</td>
<td>29.27</td>
<td>3.03</td>
<td>108.10</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_7(x))</td>
<td>32.78</td>
<td>3.25</td>
<td>112.91</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_8(x))</td>
<td>35.06</td>
<td>3.34</td>
<td>117.24</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_9(x))</td>
<td>38.80</td>
<td>4.01</td>
<td>108.24</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{10}(x))</td>
<td>41.12</td>
<td>4.37</td>
<td>105.37</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{13L}(x_E))</td>
<td>21.07</td>
<td>4.90</td>
<td>48.10</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{14L}(x_E))</td>
<td>19.53</td>
<td>4.81</td>
<td>45.38</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{15L}(x_E))</td>
<td>21.38</td>
<td>4.60</td>
<td>51.96</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_4^H(x))</td>
<td>27.35</td>
<td>6.92</td>
<td>44.23</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_7^H(x))</td>
<td>34.81</td>
<td>3.82</td>
<td>102.06</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_9^H(x))</td>
<td>37.28</td>
<td>3.60</td>
<td>115.76</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{10}^H(x))</td>
<td>39.06</td>
<td>3.47</td>
<td>126.07</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{11}^H(x))</td>
<td>41.68</td>
<td>3.52</td>
<td>132.52</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{12}^H(x))</td>
<td>43.61</td>
<td>3.58</td>
<td>136.16</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td><strong>Non-antitrypanosomal agents clusters (k-MCA II)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(q_6(x))</td>
<td>60.14</td>
<td>9.33</td>
<td>109.57</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_7(x))</td>
<td>53.57</td>
<td>9.60</td>
<td>94.82</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_8(x))</td>
<td>56.86</td>
<td>9.70</td>
<td>99.64</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_9(x))</td>
<td>52.16</td>
<td>10.34</td>
<td>85.73</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{10}(x))</td>
<td>55.78</td>
<td>10.44</td>
<td>90.76</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{13L}(x_E))</td>
<td>121.32</td>
<td>11.01</td>
<td>187.23</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{14L}(x_E))</td>
<td>126.71</td>
<td>10.94</td>
<td>196.72</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{15L}(x_E))</td>
<td>123.22</td>
<td>14.02</td>
<td>149.32</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_4^H(x))</td>
<td>77.20</td>
<td>16.76</td>
<td>78.27</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_7^H(x))</td>
<td>58.04</td>
<td>11.25</td>
<td>87.66</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_9^H(x))</td>
<td>53.15</td>
<td>11.47</td>
<td>78.74</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{10}^H(x))</td>
<td>52.98</td>
<td>11.46</td>
<td>78.57</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{11}^H(x))</td>
<td>49.55</td>
<td>11.96</td>
<td>70.39</td>
<td>0.00</td>
<td></td>
</tr>
<tr>
<td>(q_{12}^H(x))</td>
<td>50.01</td>
<td>12.18</td>
<td>69.75</td>
<td>0.00</td>
<td></td>
</tr>
</tbody>
</table>

\(^a\)Variability between groups.  
\(^b\)Variability within groups.  
\(^c\)Level of significance.

In this sense, it can be concluded that the data set of antitrypanosomal compounds considered for this study encloses compounds of six general structural patterns codified by TOMOCOMD descriptors and recognized by a \(k\)-means cluster analysis.

**Developing a Discriminant Function.** Linear discriminant analysis (LDA) has become an important tool for the prediction of chemicals properties. On the basis of the simplicity of this method many useful discriminant models have been developed and presented by different
authors in the literature.\textsuperscript{7-10} Being the election technique used on the generation of the \textit{TOMOCOMD} approaches reported to date,\textsuperscript{20-24} LDA was also employed in the current work to generate a discriminant function. The principle of parsimony (Occam's razor) was taken into account as strategy for model selection.\textsuperscript{42} It means, that we select the model with higher statistical signification but having as few parameters ($a_k$) as possible.

Making use of the LDA technique implemented in the STATICTICA software,\textsuperscript{43} the following linear model was obtained:

\[
\text{Class} = -5.18 + 2.36 \times 10^{-4} q_1(x) - 1.30 \times 10^{-4} q_2(x) + 2.08 \times 10^{-5} q_3(x) + 0.97 \times 10^{-7} q_{14L}(x_E) - 2.92 \times 10^{-8} q_{15L}(x_E) - 3.28 \times 10^{-4} q_4^H(x)
\]

\textit{(8)}

N = 101 \quad \lambda = 0.36 \quad D^2 = 7.10 \quad F(6,94) = 27.16 \quad p<0.0001

where, N is the number of compounds, $\lambda$ is Wilk's coefficient, F is the Fisher ratio, $D^2$ is the squared Mahalanobis distance and $p$-value is the significance level. The antiparasitic activity was codified by a dummy variable “\textit{Class}”, which indicates either the presence of an active compound (\textit{Class} = 1) or an inactive one (\textit{Class} = -1). The classification of cases was performed by means of the posterior classification probabilities, which is the probability that the respective case belongs to a particular group (active or inactive). By using the models, each compound can be then classified as active, if $\Delta P\% > 0$, being $\Delta P\% = [P(\text{Active}) - P(\text{Inactive})] \times 100$ or as inactive otherwise. Compounds with $\Delta P\% < 5\%$ were considered as non classified. Table 2 shows these results.

\textbf{Table 2.} Classification of active and inactive compounds included in the training set using Model 8.

<table>
<thead>
<tr>
<th>Compound</th>
<th>$\Delta P%$</th>
<th>Class.</th>
<th>Compound</th>
<th>$\Delta P%$</th>
<th>Class.</th>
</tr>
</thead>
</table>
| \textit{Training Active Group} | \begin{tabular}{lrr}
1 & 0.99 & + \\
2 & 0.98 & + \\
3 (Mecaprine) & 0.99 & + \\
4 & 0.98 & + \\
5 & 0.87 & + \\
6 (Chlorotacrine) & 0.94 & + \\
7 & 1.00 & + \\
8 (Formycin B) & 0.89 & + \\
9 (Tubercidin) & 0.68 & + \\
10 (Megazol) & 0.95 & + \\
11 (Allopurinol) & 0.93 & + \\
12 & 0.98 & + \\
13 & 1.00 & + \\
14 & 0.76 & + \\
15 & 0.77 & + \\
16 & 0.78 & + \\
\end{tabular} | | \begin{tabular}{lrr}
21 & 0.80 & + \\
22 & 1.00 & + \\
23 & 1.00 & + \\
24 & 0.43 & + \\
25 & 0.84 & + \\
26 & 0.42 & + \\
27 & 0.50 & + \\
28 & 0.03 & NC \\
29 & 0.62 & + \\
30 & -0.73 & - \\
31 & -0.27 & - \\
32 & 0.53 & + \\
33 & 0.99 & + \\
34 & -0.31 & - \\
35 & 0.63 & + \\
36 & 0.68 & + \\
\end{tabular} |
As can be computed from the results showed in Table 2, model 8 classified correctly 93.02% of the whole training data set (accuracy). This model showed a high Matthews’ correlation coefficient (MCC) of 0.87. MCC is a measure that may provide a much more balanced evaluation of the prediction than the percentages of good classification, because it uses all four numbers (true positive, true negative, false positive and false negatives). Also the probability of correctly predicting a positive example (sensitivity or hit rate) and the probability that a positive prediction will be correct (specificity) were computed for the model. In both cases 92.31% was the obtained value. While these two later measures bring some information of the predictivity for positive observations, the negative predictive value (sensitivity of the negative
category) gives a criterion of good classification for the inactive group. In this case a 95.08% was observed. These results, as well as the “false positive rate” (false alarm rate) are depicted in Table 3.

Every statistical model which is generated based on a previously selected data set of observations, includes information of just a portion of the universe and has an error range, which the researcher tries to minimize during the modeling process. In this sense, the false positive rate, as well as the false negative rate, are used as measures of the error range and the confiability of the model. A correct selection of a training data sets can reduce the magnitude of both measures. We took this aspect into consideration and built a training data set choosing chemicals with so much structural variability as possible. Despite of the previous precaution, it can happen that the combination of some structural paterns of a positive case, for instance, results in mathematical values wich are closer to those obtained from the combination of structural fragments in a negative observation. In such a case the model will not recognize the true class of the observation. In the present study, three active and three inactive compounds were missclasified. Here it is also important to note, that the declaration of each non-antitrypanosomal compound as “inactive” does not mean that there not exist antitrypanosomal side-effects, given it can include organic drugs for which antitrypanosomal activity has been left undetected so far. In this sense, any discriminant model can be continuously transformed and improved, taking into consideration unavailable information at the time of the model’s development. This problem can affect in some degree the results of further classification. Just testing the biological activity of them it is possible to ensure the absence of antitrypanosomal effects. In this sense, we can recommend carrying out the biological assays for previously declared “inactive” compounds, for which the model give a positive classification.

Considering that a discriminant model could be accepted or rejected depending on its predictive power, it is clear to see that validation processes constitute obligated steps for the assessment of any structure-activity relationship. As Golbraikh and Tropsha emphasized, the predictive ability of a QSAR model can only be estimated using an external test set of compounds which were never used for the development of the model. In this sense, it is important to secure, that the prediction algorithm is able to perform well on novel data from the same data domain. In our case, as first validation experiment, an external prediction data set was evaluated. The computation of some performance measures such as Matthews correlation coefficient, percentage of global good classification (accuracy), sensitivity, specificity, false alarm rate and negative predictive value (sensitivity of the negative category)
permitted us to carry out the assessment of the model. In Table 3 are also depicted the results for this validation process.

**Table 3.** Overall measures of accuracy obtained in the training and prediction sets for the model 7.

<table>
<thead>
<tr>
<th>Matthews Corr. Coefficient</th>
<th>Accuracy (%)</th>
<th>Sensitivity (hit rate%)</th>
<th>Specificity (%)</th>
<th>False alarm rate (%)</th>
<th>Predictive value (-) (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Training set</td>
<td>0.87</td>
<td>93.06</td>
<td>92.31</td>
<td>92.31</td>
<td>4.92</td>
</tr>
<tr>
<td>Test set</td>
<td>0.88</td>
<td>94.23</td>
<td>90.91</td>
<td>95.24</td>
<td>3.33</td>
</tr>
</tbody>
</table>

The classification’s results using model 8 for active and inactive compounds in the selected test set are shown in Table 4.

**Table 4.** Classification of active and inactive compounds included in test series using the model 8.

<table>
<thead>
<tr>
<th>Compound</th>
<th>ΔP%</th>
<th>Class.</th>
<th>Compound</th>
<th>ΔP%</th>
<th>Class.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Test Active Set</td>
<td></td>
<td></td>
<td>Test Inactive Set</td>
<td></td>
<td></td>
</tr>
<tr>
<td>41</td>
<td>1.00</td>
<td>+</td>
<td>52</td>
<td>0.79</td>
<td>+</td>
</tr>
<tr>
<td>42</td>
<td>1.00</td>
<td>+</td>
<td>53</td>
<td>0.97</td>
<td>+</td>
</tr>
<tr>
<td>43</td>
<td>0.98</td>
<td>+</td>
<td>54</td>
<td>-0.07</td>
<td>-</td>
</tr>
<tr>
<td>44</td>
<td>0.99</td>
<td>+</td>
<td>55</td>
<td>0.99</td>
<td>+</td>
</tr>
<tr>
<td>45</td>
<td>0.99</td>
<td>+</td>
<td>56 (Brazilizone A)</td>
<td>1.00</td>
<td>+</td>
</tr>
<tr>
<td>46 (Nifurtimox)</td>
<td>-0.95</td>
<td>-</td>
<td>57</td>
<td>0.58</td>
<td>+</td>
</tr>
<tr>
<td>47 (Benznidazol)</td>
<td>0.42</td>
<td>+</td>
<td>58</td>
<td>0.22</td>
<td>+</td>
</tr>
<tr>
<td>48</td>
<td>0.91</td>
<td>+</td>
<td>59</td>
<td>0.29</td>
<td>+</td>
</tr>
<tr>
<td>49</td>
<td>0.76</td>
<td>+</td>
<td>60</td>
<td>0.59</td>
<td>+</td>
</tr>
<tr>
<td>50</td>
<td>0.77</td>
<td>+</td>
<td>61</td>
<td>0.41</td>
<td>+</td>
</tr>
<tr>
<td>51</td>
<td>0.43</td>
<td>+</td>
<td>62</td>
<td>1.00</td>
<td>+</td>
</tr>
<tr>
<td>Amantadine</td>
<td>-1.00</td>
<td>-</td>
<td>Cyclopropane</td>
<td>-0.99</td>
<td>-</td>
</tr>
<tr>
<td>Mizoribine</td>
<td>-0.16</td>
<td>-</td>
<td>Basedol</td>
<td>-0.95</td>
<td>-</td>
</tr>
<tr>
<td>Triclofos</td>
<td>-1.00</td>
<td>-</td>
<td>Mipimazole</td>
<td>-0.99</td>
<td>-</td>
</tr>
<tr>
<td>Nitroinosite</td>
<td>-0.92</td>
<td>-</td>
<td>Didym levulinate</td>
<td>-1.00</td>
<td>-</td>
</tr>
<tr>
<td>Methenamine</td>
<td>-0.99</td>
<td>-</td>
<td>Metriponate</td>
<td>-1.00</td>
<td>-</td>
</tr>
<tr>
<td>Cobalti glutamas</td>
<td>-1.00</td>
<td>-</td>
<td>Prasterone</td>
<td>-0.98</td>
<td>-</td>
</tr>
<tr>
<td>Cobalti besilas</td>
<td>-1.00</td>
<td>-</td>
<td>Febensamin</td>
<td>-1.00</td>
<td>-</td>
</tr>
<tr>
<td>Camrenone</td>
<td>-0.92</td>
<td>-</td>
<td>Guanazole</td>
<td>-0.99</td>
<td>-</td>
</tr>
<tr>
<td>Urea</td>
<td>-1.00</td>
<td>-</td>
<td>Fluorembichin</td>
<td>-1.00</td>
<td>-</td>
</tr>
<tr>
<td>Pallirad</td>
<td>-0.99</td>
<td>-</td>
<td>Mitoguazone</td>
<td>-0.99</td>
<td>-</td>
</tr>
<tr>
<td>Quimbosan</td>
<td>-0.99</td>
<td>-</td>
<td>Acetylcholine</td>
<td>-1.00</td>
<td>-</td>
</tr>
<tr>
<td>Glicondamide</td>
<td>0.65</td>
<td>+</td>
<td>Methacholine chloride</td>
<td>-1.00</td>
<td>-</td>
</tr>
<tr>
<td>RMI 11894</td>
<td>-1.00</td>
<td>-</td>
<td>Dopamine</td>
<td>-0.78</td>
<td>-</td>
</tr>
<tr>
<td>Barbismetylis iodidum</td>
<td>-0.59</td>
<td>-</td>
<td>Ampicillin</td>
<td>-0.42</td>
<td>-</td>
</tr>
<tr>
<td>Frigen 113</td>
<td>-0.98</td>
<td>-</td>
<td>Kanamycin A</td>
<td>-1.00</td>
<td>-</td>
</tr>
</tbody>
</table>

*Results of the classification of compounds obtained from Eq. 8 (using non-stochastic quadratic indices): ΔP% = \[\frac{P(Active) - P(Inactive)}{NC} \times 100\], NC= not classified.
A second validation experiment was also developed upon the base of a Leave-Group-Out internal cross-validation strategy. In this case, groups of compounds including 10% of the training data set are left out and afterwards predicted for the model obtained with the remaining 90%. This process was repeated 10 times for each one of the 10 unique subsets selected at random and each observation predicted once (in its group of left-out observations). The overall mean for this process (10% full leave-out cross-validation) was used as a good indication of robustness and stability of the obtained models. In Table 5 the results of classification for each 10%-group, as well as the classification for the remaining training set leaving out each one of those groups are depicted. From these results we can conclude, that also by this experiment, our model had a robust and stable behavior.

Table 5. Predictability based on the use of ten randomly selected subsets (LGO cross-validation) of the LDA Model.

<table>
<thead>
<tr>
<th>Group</th>
<th>% Global Good Classification</th>
<th>Test set (10%)</th>
<th>Remaining training set</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>100.00</td>
<td>93.40</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>100.00</td>
<td>93.40</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>100.00</td>
<td>93.40</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>100.00</td>
<td>93.40</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>80.00</td>
<td>92.30</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>90.00</td>
<td>93.40</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>90.00</td>
<td>94.50</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>80.00</td>
<td>95.60</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>90.00</td>
<td>92.30</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>90.90</td>
<td>93.33</td>
<td></td>
</tr>
<tr>
<td>Overall mean</td>
<td>91.38</td>
<td>93.50</td>
<td></td>
</tr>
<tr>
<td>Standard deviation</td>
<td>7.86%</td>
<td>0.96%</td>
<td></td>
</tr>
</tbody>
</table>

No previous reports related to the application of pattern recognition techniques to the selection of antitrypanosomal compounds from a heterogeneous series of compounds were found in the literature. In this sense, the present algorithm constitutes a step forward in the search of efficient ways to discover new antitrypanosomal drugs.

With the aim to evaluate the applicability of the present TOMOCOMD methodology, four families of 2D molecular descriptors were computed with the DRAGON Software and the respective models were generated. The statistical parameters are shown in Table 6. As can be seen, in not one case the classification results are better than those obtained using model 8. These results are a proof of the usefulness of the TOMOCOMD strategy in the study of this biological property.
Table 6. Comparison between model 8 and four models obtained using different kinds of 2D descriptors.

<table>
<thead>
<tr>
<th>Models</th>
<th>Matthews Corr. Coefficient</th>
<th>Accuracy (%)</th>
<th>Number of Variables</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Training Set</td>
<td>Test Set</td>
<td>Training Set</td>
</tr>
<tr>
<td>Model 8</td>
<td>0.87</td>
<td>0.88</td>
<td>93.06</td>
</tr>
<tr>
<td>Topological descriptors</td>
<td>0.80</td>
<td>0.68</td>
<td>85.14</td>
</tr>
<tr>
<td>Molecular walk counts</td>
<td>0.45</td>
<td>0.42</td>
<td>69.31</td>
</tr>
<tr>
<td>BCUT descriptors</td>
<td>0.64</td>
<td>0.60</td>
<td>81.19</td>
</tr>
<tr>
<td>2D autocorrelations</td>
<td>0.86</td>
<td>0.85</td>
<td>91.09</td>
</tr>
</tbody>
</table>

An experiment of rational search of novel antitrypanosomal compounds. The importance and usefulness of QSAR model can only be assessed, by predicting the activity of new compounds not used in the process of constructing the classification algorithm and afterwards, carrying out the biological corroborations of such predictions. With the aim of testing the ability of our model to detecting new lead compounds, we design a simulated virtual screening experiment using model 8. To avoid the manipulation of large databases of chemicals, and just as an example of applicability of our approach, we selected a series of ten new synthesized heterocyclic compounds obtained in one of our research groups.\(^{48-51}\)

As first step of this virtual screening, all structures were drawn using de drawing mode implemented in the TOMOCOMD software. After that, the the \(^k\)th total quadratic indices \([q_k(x)]\) and \([q_k^H(x)]\) and the \(^k\)th local ones (atom-type = heteroatoms: S, N, O) \([q_k^L(x_E)]\) and \([q_k^L^H(x_E)]\) not considering and considering H-atoms respectively were computed. Each compound was evaluated using model 8 and finally in vitro assayed against epimastigotes forms of *Trypanosoma cruzi*. Epimastigotes are the extracellular multiplying forms of the mentioned parasite which are relatively sensitive to a drug action. They can be easily cultured and in this sense, are an excellent platform for preliminary in vitro screening of antitrypanosomal activity.\(^{52}\) However, once this first assays have been performed, more selective methods are required to determine the activity of novel compounds. In the current work, the preliminary screening on epimastigotes forms was the election way to evaluate the activity of the predicted compounds, which structures are shown in Figure 4.
Figure 4. Structures of ten synthesized compounds evaluated using model 8.

![Chemical structures](image)

The results of the prediction process using model 8, as well as the anti-epimastigotes percentage for each assayed compound are summarised in Table 7. In this case, nifurtimox was employed as reference drug.

Table 7. Compounds which were evaluated in the present study, their classification (ΔP%) according to the TOMOCOMD approach, their antitrypanosomal activity at three different concentrations (100, 10 and 1 µg/ml) and antitrypanosomal activity of Nifurtimox (Reference).

<table>
<thead>
<tr>
<th>Comp.</th>
<th>ΔP%</th>
<th>Class.</th>
<th>Obs.</th>
<th>%AE (SS)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>100(µg/ml)</td>
<td>10(µg/ml)</td>
</tr>
<tr>
<td>1s</td>
<td>1.00</td>
<td>+</td>
<td>79.90 (1.60)</td>
<td>73.50 (0.70)</td>
</tr>
<tr>
<td>2s</td>
<td>1.00</td>
<td>+</td>
<td>99.68 (0.15)</td>
<td>36.78 (0.28)</td>
</tr>
<tr>
<td>3s</td>
<td>0.97</td>
<td>+</td>
<td>83.87 (0.40)</td>
<td>23.13 (1.56)</td>
</tr>
<tr>
<td>4s</td>
<td>0.97</td>
<td>+</td>
<td>89.60 (0.61)</td>
<td>84.60 (1.50)</td>
</tr>
<tr>
<td>5s</td>
<td>0.99</td>
<td>+</td>
<td>100.00 (0.60)</td>
<td>56.82 (0.25)</td>
</tr>
<tr>
<td>6s</td>
<td>-0.04</td>
<td>NC</td>
<td>-</td>
<td>49.78 (0.40)</td>
</tr>
<tr>
<td>7s</td>
<td>-0.34</td>
<td>-</td>
<td>-</td>
<td>0.00 (1.16)</td>
</tr>
<tr>
<td>8s</td>
<td>-0.26</td>
<td>-</td>
<td>-</td>
<td>6.90 (4.19)</td>
</tr>
<tr>
<td>9s</td>
<td>-0.28</td>
<td>-</td>
<td>-</td>
<td>37.56 (1.11)</td>
</tr>
<tr>
<td>10s</td>
<td>-0.10</td>
<td>-</td>
<td>-</td>
<td>58.65 (3.70)</td>
</tr>
<tr>
<td>Nifurtimox</td>
<td>98.73 (0.56)</td>
<td>90.05 (1.80)</td>
<td>75.50 (3.89)</td>
<td></td>
</tr>
</tbody>
</table>

*Results of the classification of compounds obtained from Model 8, ΔP% = [P(Active) - P(Inactive)]x100, Class. Classification, Obs. Observed activity, %AE Anti-epimastigotes percentage and standard deviation (SS), NC= not classified.

As predicted, five compounds (compounds 1s-5s) showed trypanocidal activity (%AE>70). Compounds 2s, 3s and 5s are only active against epimastigotes at 100 µg/ml. Two compounds, 1s and 4s, showed appreciable activity at concentration of 10 µg/ml. Specifically compound
4s resulted very interesting for having inhibition percentages (%AE) higher than 80% at 100, 10 and 1 µg/ml. Further research will be required to investigate the mechanism of action of these compounds and to evaluate their cytotoxicity at the assayed concentrations. The remaining five compounds which were classified as inactive for the model, showed very low inhibition percentages. For compounds 7s and 8s for instance, 0.0% and 6.9% of inhibition were determined at 100µg/ml and 0.0 % at other concentrations. According to the model, these are the compounds with a greater probability to result inactive ones.

This first virtual screening demonstrates the ability of the present TOMOCOMD approach to be used for discriminating compounds with potential antitrypanosomal activity from those without this action. These results open at the same time a door for the study of several families of heterocyclic compound, which seems to be promissory sources of antitrypanosomal drugs. Current investigations are been developed in this direction by our research groups.

3. Concluding remarks

The search of effective and rational methodologies for the discovery of new drugs has turn into a first-line objective for the pharmaceutical research. In spite of some criticism, topological-indices-based approaches have demonstrated their usefulness in drug discovery processes. TOMOCOMD methodology has become an attractive tool to be used in chem- and bioinformatic research. This strategy allowed us to generate a mathematical model able to discriminate antitrypanosomal compounds from inactive ones and to predict, in a rational way, the activity against Trypanosoma cruzi of novel heterocyclic compounds. This family constitutes a starting point for the design and synthesis of each time more effective and less toxic antitrypanosomal compounds.

The current approach can be used in further computational screenings of larger chemical libraries in order to discover new candidates to antitrypanosomal drugs using a minimum of resources. The interactive and flexible character of the TOMOCOMD approach permits the posterior inclusion of other active and inactive compounds in the training set and the generation of each time more refinished models capable of identifying structural patterns not considered in the present study.

Upon the base of the current results we can conclude, that the TOMOCOMD strategy can be successfully used in the rational search of novel antitrypanosomal compounds.
4. Experimental Section

Computational Approach. Calculations were carried out on a PC Pentium-4 2.0 GHz. The CARDD-module implemented in the TOMOCOMD Software\textsuperscript{13} was used to the calculation of total and local non-stochastic quadratic indices. Pauling electronegativities\textsuperscript{53} were used as atomic weights (molecular vector’s components).

Topological descriptors, molecular walk counts, BCUT descriptors and 2D autocorrelations were calculated by using the DRAGON Software.\textsuperscript{47} The molecular structure of each compound was drawn by using the CHEMDRAW software\textsuperscript{54} and saved as a .mol file. After optimization with the MOPAC software\textsuperscript{55} the structures were saved as a .hin file and then processed by the DRAGON Software.

Chemometric Method. Linear discriminant analysis (LDA) was performed as implemented in the STATISTICA 5.5 for Windows package.\textsuperscript{43} Forward stepwise was fixed as strategy for variable selection. The quality of the models was determined by examining Wilk’s $\lambda$ parameter ($U$-statistic), square Mahalanobis distance ($D^2$), Fisher ratio (F) and the corresponding $p$-level (p(F)) as well as the percentage in training and test sets of global good classification, Matthews’ correlation coefficient (MCC), sensitivity, specificity, negative predictive value (sensitivity of the negative category) and false positive rate (false alarm rate).

Models with a proportion between the number of cases and variables in the equation lower than 4 were rejected. The statistical robustness and predictive power of the obtained model was assessed using an external prediction (test) set. A leave-group out (10\%) cross validation procedure was also carried out for this proposes.

Parasites and culture procedure. CL strain parasites (clone CL-B5) stably transfected with the Escherichia coli $\beta$-galactosidase gene (LacZ) were used for the assays. Epimastigotes were grown at 28º C in liver infusion tryptone broth (LIT) with 10% foetal bovine serum (FBS), penicillin and streptomycin.

Antiepimastigote assay.\textsuperscript{52} The screening assay was performed in 96-well microplates with culture that had not reached the stationary phase. Epimastigotes forms, CL strain, were seeded at concentration of $1 \times 10^5$ per ml in 200 µl. The plates were then incubated at 28º C for 72 hours with various concentrations of the drugs (100, 10 and 1 µg/ml), at which time 50 µl of CPRG solution was added to give a final concentration of 200 µM. The plates were incubated at 37º C for an additional 6 hrs and were then read at 595 nm. Each concentration was assayed three times. In order to avoid drawback, medium, negative and drug controls were used in each test. The anti-epimastigotes percentage (%AE) was calculated as follows:
%AE=[(AE-AEB)/(AC-ACB)] x 100, where AE = absorbance of experimental group; AEB = blank of compounds; AC = absorbance of control group; ACB = blank of culture medium. Stock solutions of the compounds to be assayed were prepared in DMSO, with the final concentration in a mixture water/DMSO never exceeding 0.2% of the last solvent.

5. References


43. STADISTICA, version 5.5; Statsoft Inc., 1999.


