

Proceeding Paper



Smartphone-Adapted Multianalyte Biosensor Platform for Fluorescent Analysis of Human Biomarkers and Immunosuppressive Drugs Using PQQ- and NAD⁺-Dependent Enzymes ⁺

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Abstract: Here we describe a multianalyte biosensor platform for fluorescent analysis of different human state biomarkers (α -amylase, phenylalanine, glucose, lactate/pyruvate, alcohol) and some immunosuppressive drugs (cyclosporine A, tacrolimus, methotrexate, rapamycin) using chimeric PQQ- and natural NAD⁺-dependent enzymes. The principle of the approach is based on the analysis of the brightness of photography of a sensor plate taken with a smartphone camera and processed via ImageJ software. The brightness of the image correlates with the fluorescence intensity of the sensor's spots which is produced by enzymatic reduction of phenazine methosulfate or its derivative used as a fluorescence probe at UV 356 nm irradiation, where the amount of the reduced dye depends on the concentration of the target analyte (the enzymatic substrate) in the tested sample. The sensor plate is composed of simple and cheap components, while the procedure of its preparation and usage is easy and does not require any specific skills or expensive instrumentation. The proposed sensor platform is characterized by a high selectivity and storage stability depending on the selectivity and stability characteristics of the used enzyme in an immobilized state. The proposed sensor platform could be used for precision quantitative analysis of a single (or several) analytes or used for simultaneous qualitative multianalyte assay of numerous of them via Boolean logic gates.

Keywords: smartphone-based biosensor; fluorescent analysis; PQQ- and NAD⁺-dependent enzymes; human biomarkers; immunosuppressive drugs

1. Introduction

The accurate analysis of trace amounts of some immunosuppressive drugs in patients and the main biomarkers indicating the human physiological state is a very actual need of modern clinical diagnostics and medicine. Most of the available techniques for such analysis require valuable equipment, complicated sample pretreatment, and time-consuming technical procedures processed by highly qualified personnel in clinical laboratories. From this point of view, the developments of portable cheap, and easy-to-use biosensors look a very promising direction in modern biotechnologies with a high prospective for their successful commercialization [1]. Nowadays, due to permanent improvements in gadgets and user-friendly software, it starts to be possible to adapt them for point-ofcare biosensor analysis of such analytes substituting the expensive equipment and needs of qualified personnel [2,3]. Usually, biosensors are characterized by a limited number of target analytes to be tested as one biosensor is able to selectively test one specific analyte. So, to estimate the overall information about the human state it is needed to use different biosensors selective to different biomarkers. To solve this limitation we propose a new

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Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/). smartphone-adapted multianalyte biosensor platform for fluorescent analysis of human biomarkers and immunosuppressive drugs using PQQ- and NAD⁺-dependent enzymes. The principle of the approach is based on the analysis of the brightness of photography of the sensor plate taken with a smartphone camera and processed via the free software ImageJ. The brightness of the image correlates with the fluorescence intensity of the sensor's spots which is produced by the enzymatic reduction of the fluorescent probe at a long wave UV irradiation, which correlates with the concentration of the target analyte (the specific enzymatic substrate) in the tested sample. The main goal of the investigation is a combination of different enzymes selective to different analytes on the same sensor's plate to perform the direct simultaneous analysis of several drugs and/or biomarkers in the same tested sample. To simplify the analysis we propose to analyze the specific optical output via Boolean logic gates, which gives information in a binary system as "output 1" or "output 0" that is equivalent to the "Yes" or "No" binary response. The proposed system was already successfully approbated for analysis of different biomarkers and some immunosuppressive drugs in different human liquids (e.g., blood serum, urine, sweat, and saliva) [4–6]. It should be mentioned that the possibility to test the target analytes in different types of human liquids make it more convenient for practical application because it avoids painful procedures associated with blood sampling by qualified medical personnel. This also makes it possible to often sample collection to have continuous information about the efficacy of medication in its dynamics. We expect that the proposed multianalyte biosensor platform will give fast and accurate information to end-users (patients) directly at point-of-care which is more convenient compared to attending clinical diagnostics centers.

2. Background and Goal

The principle of the approach is based on the analysis of the intensity (brightness) of photography of the sensor plate taken with a smartphone camera in a grayscale. The brightness of the image correlates with the fluorescence intensity of the sensor's spots which is produced by enzymatic oxidation of phenazine methosulfate or its derivative used as a fluorescence probe at UV 356 nm irradiation, where the amount of the reduced dye depends on the concentration of the target analyte (the enzymatic substrate) in the tested sample (Figure 1 A,B).





Figure 1. (**A**) Schematic presentation of the biocatalytic process generating fluorescent output proportional to the target analyte (glucose) concentration. Abbreviations: GDH-glucose dehydrogenase, Glc-glucose, GlcA-gluconic acid, PQQ and PQQH₂-oxidized and reduced forms of **pyrrol-oquinoline quinone**, **respectively**, **PMS**_{ox} **and PMS**_{red}-phenazine methosulfate oxidized and reduced forms, λ_{ex} and λ_{em} -wavelength maximum of **PMS**_{red} **fluorescence+-.** Figure adapted from [7] with permission. (**B**) Processing fluorescent image (sensor output) and its analysis with ImageJ software installed in a smartphone. Figure adapted from [8] with permission.

The sensor plate is composed of simple and cheap components, while the procedure of its preparation is easy and does not require any specific skills or expensive instrumentation (Figure 2).



Figure 2. The components used for the biosensor preparation: VWR fiberglass filter paper, doublesided tape, microscopic glass slides with deposited fiberglass disks, an office hole-puncher for preparing the sensing disks, a multi-pipet, and a smartphone with a camera for taking photos of the biosensor platform. Figure adapted from [8] with permission.

Then, the surface of the fiberglass sheets of the sensor plate was functionalized by bioselective elements (enzymes) which were covalently immobilized on silica nanoparticles (SiO₂-NPs). The immobilization of the enzymes onto SiO₂-NPs provides simultaneously two functions: (i) high sensor stability preventing washing out the enzymes during the operation, and (ii) increasing the sensor sensitivity (due to amplification of fluorescence intensity) penetrating enzyme-functionalized SiO₂-NPs deep inside of the fiberglass support and forming a multilayer structure. It should be noted, that the reduced dye in

contrast to its oxidized non-fluorescent form is characterized by high hydrophobicity. This characteristic of the probe additionally supports high output's stability preventing its leak in buffer solutions during the washing of the sensor plate (Figure 3).



Figure 3. (A) The scheme of immobilization of enzymes onto SiO₂-NPs. (**B**,**C**) Scanning electron microscopy images showing the enzyme-functionalized SiO₂-NPs deposited on fiberglass support. (**D**) Microphotography of confocal microscopy visualized the distribution of enzyme-functionalized SiO₂-NPs in the sensing spot. The penetration of enzyme-functionalized SiO₂-NPs into 125 μ m of the glass paper (blue color) support corresponds to app. 625 of single monolayers of SiO₂-NPs (200 nm in diameter each).

The analytical procedure of the biomarker detection by a ready-to-use sensor plate is presented in Figure 4.



Figure 4. The sample photos of the biosensing system for analysis of glucose (Glc) obtained at different stages of sensor analysis: as prepared, prior to application of glucose, after applying glucose at different concentrations on each sensing spot; after rinsing the sensing spots with buffer, after converting the previous photo to the gray format ready to be processed with software "ImageJ" installed in a smartphone. Figure adapted from [8] with permission.

To take photos of the sensor plate during its irradiation by UV 356 nm, it is convenient to use the UV fluorescence analysis cabinet. However, it should be noted, that instead of a photo cabinet with an integrated UV lamp, it could be used a dark room and a cheap small lantern (with lamp 365 nm) used for in-home pet tick inspection. The proposed sensor platform is characterized by a high selectivity which depends on the selectivity characteristics of the used enzymes. The examples of the analysis selectivity for glucose-selective biosensor based on NAD⁺-dependent glucose dehydrogenase is presented in Figure 5.



Figure 5. Selectivity test of the glucose-sensitive biosensor towards possible interferents of common blood serum compounds. Photographs show the sensing plate with the spots reacted with the target analyte glucose and with three interferents—urate, lactate, and ethanol (1 mM each) and a bar chart showing the fluorescence intensity obtained with the target analyte glucose and with three interferents. The intensity was derived from the image as the brightness of the spots was calculated with the ImageJ software. Figure adapted from [9] with permission.

The biosensor plate storage stability depends on the stability characteristics of the used enzyme in an immobilized state. It was shown that the sensors based on different enzymes were sufficiently stable for at least of 2 months of storage at +8 °C (shelf-storage). The authors believe, that the storage stability of the sensor could be improved by pretreatment of the plates with enzyme stabilizing agents, using freeze-dry, vacuum packing, etc.

The proposed sensor system was approbated for analysis of human biomarkers and some immunosuppressive drugs in different human liquids (blood serum, saliva, sweat, urine). The list of tested analytes is presented in Table 1 with indicated corresponding references where the detailed procedures of assay are described.

PQQ-Dependent GDH (Including Chimeric Forms):		NAD*-Dependent Dehydrogenases:	
1. Glucose	[7]	1. Phenylalanine	[6]
2. α-Amylase	[5]	2. Glucose	[9]
3. Cyclosporine A	[4]	3. Lactate	-
4. Tacrolimus (FK 506)	[4]	4. Pyruvate	-
5. Rapamycin	[10]	5. Alcohol	[9]
6. Methotrexate	[11]		

Table 1. The list of analytes tested by means of the developed multianalyte biosensor platform.

The proposed sensor platform could be used for precision quantitative analysis of a single (or several) analytes or used for simultaneous qualitative multianalyte assay of numerous of them.

For the simultaneous multianalyte analysis, the most convenient seems to be a binary system used in Boolean logic gates where a logical operation uses one or more binary inputs to produce a single binary output [12]. In this case, the over amount of the analyte will be represented as fluorescent output (1) when its amount in the normal concentration or below the norm will produce the fluorescent output (0) or the signal will significantly lower than output (1). The sample of using the proposed sensor platform via Boolean logic gates is presented in Figure 6.



Figure 6. The sample of using the proposed fluorescent biosensor system for multianalyte analysis via Boolean logic gates. Each pair of sensing spots is modified with different enzymes selective to a unique analyte. When all the sensor spots are treated with the same tested sample (e.g., human serum from one patient) the increased amount of some biomarkers will provide the intensive spot's fluorescence (output 1) (see intensive blue spots in (**A**)) when their normal level will be taken as "no output" (output **0**) (see dark blue/weakly visible spots in (**A**)). (**B**) The bar chart shows the fluorescence intensity of the spots corresponding to the logic outputs **0** and **1** derived from the images by calculating the image brightness using the ImageJ software.

The simultaneous combination of outputs **1** from spots sensitive to glucose, lactate, cholesterol, and/or **glycerol** will indicate a high probability of diabetes in the tested body or some adiposity-related metabolic dysfunctions. The combination of other biomarkers from the list in Table 1, will predict other health issues or the current human physiological states and this is the aim of the ongoing work.

3. Summary and Outlook

The proposed multianalyte optical biosensor platform is characterized by high selectivity, sensitivity, and sufficient storage stability. Moreover, it is cheap and easy in preparing and use. It was tested for highly sensitive accurate analysis of such human biomarkers as α -amylase, phenylalanine, glucose, lactate/pyruvate, alcohol, and some immunosuppressive drugs like cyclosporine A, tacrolimus, methotrexate, and rapamycin. The possibility to analyze the target analytes in different human bioliquids (e.g., blood serum, urine, sweat, and saliva) was demonstrated, thus allowing sample collections from patients, avoiding painful puncturing procedures. Due to the possibility of fast simultaneous analysis of several biomarkers/drugs, the system could be very helpful for pointof-care use substituting expensive and time-consuming routine techniques of clinical diagnostics labs.

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References

- 1. Zhang, J.; Chen, M.; Peng, Y.; Li, S.; Han, D.; Ren, S.; Qin, K.; Li, S.; Han, T.; Wang, Y.; Gao, Z. Wearable biosensors for human fatigue diagnosis: A review. *Bioeng. Transl. Med.* **2023**, *8*, e10318.
- 2. Qian, S.; Cui, Y.; Cai, Z.; Li, L. Applications of smartphone-based colorimetric biosensors. Biosens. Bioelectron. X 2022, 11, 100173.
- 3. Zhang, M.; Cui, X.; Li, N. Smartphone-based mobile biosensors for the point-of-care testing of human metabolites. *Mater. Today Bio.* **2022**, *14*, 100254.
- 4. Wells, P.K.; Smutok, O.; Guo, Z.; Alexandrov, K.; Katz, E. Nanostructured interface loaded with chimeric enzymes for fluorimetric quantification of Cyclosporine A and FK506. *ACS Anal. Chem.* **2022**, *94*, 7303–7310.
- 5. Wells, P.K.; Smutok, O.; Guo, Z.; Alexandrov, K.; Katz, E. Fluorometric biosensing of *α*-amylase using an artificial allosteric biosensor immobilized on nanostructured interface. *Talanta* **2023**, *255*, 124215.
- 6. Melman, Y.; Katz, E.; Smutok, O. Phenylalanine biosensor based on a nanostructured fiberglass paper support and fluorescent output signal readable with a smartphone. *Microchem. J.* **2022**, *179*, 107497.
- Wells, P.K.; Melman, A.; Katz, E.; Smutok, O. Fluorescent sensor based on pyrroloquinoline quinone (PQQ)-glucose dehydrogenase for glucose detection with smartphone-adapted signal analysis. *Microchim Acta* 2022, 189, 371.
- 8. Melman, Y.; Katz, E.; Smutok, O. A universal multichannel platform for assembling enzyme-based boolean logic gates. *ChemPhysChem* **2022**, *23*, e202200352.
- 9. Melman, Y.; Wells, P.K.; Katz, E.; Smutok, O. A universal nanostructured bioanalytical platform for NAD⁺-dependent enzymes based on the fluorescent output reading with a smartphone. *Talanta* **2022**, *243*, 123325.
- Guo, Z.; Smutok, O.; Johnston, W.; Ergun Ayva, C.; Walden, P.; McWhinney, B.; Ungerer, J.; Melman, A.; Katz, E.; Alexandrov, K. Circular permutated PQQ-glucose dehydrogenase as an ultrasensitive electrochemical biosensor. *Angew. Chem. Int. Ed.* 2022, 61, e202109005.
- 11. Guo, Z.; Smutok, O.; Johnston, W.; Walden, P.; Ungerer, J.; Peat, T.; Newman, J.; Parker, J.; Nebl, T.; Hepburn, C.; Melman, A.; Suderman, R.; Katz, E.; Alexandrov, K. Design of methotrexate-controlled chemical dimerization system and its use in bio-electronic devices. *Nat. Commun.* **2021**, *12*, 7137.
- 12. Katz, E. Enzyme-Based Computing Systems, 1st ed.; Wiley-VCH Verlag GmbH & Co. KGaA: Hoboken, NJ, USA, 2019; pp. 113– 149.

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