

Removal of Phenol from Biomedical Waste by Adsorption Process [†]

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[†] Presented at the 2nd International Electronic Conference on Processes: Process Engineering – Current State and Future Trends (ECP 2023), 17–31 May 2023; Available online: <https://ecp2023.sciforum.net/>.

Abstract: Phenolic chemicals are poisonous and have long-term impacts on humans and animals. Even in low quantities, they destroy red blood cells and the liver as carcinogens. These biological wastes pollute groundwater. Thus, removing these organic chemicals to meet discharge limits is difficult. Electrochemical oxidation, redox reactions, membrane separation, and photocatalytic degradation remove phenolic chemicals from water. Recently, phenolic chemicals are removed via adsorption and photocatalysis employing carbon materials and clays. Due to their unique chemical and physical properties, nanometric materials are crucial to these processes. These substances' structure, classification, sources of an entrance, and reactivity or interaction with other aquatic components have been studied extensively. Phenolic substances are removed from the water before usage. This led to the development of water treatment technologies like activated carbon adsorption, solvent extraction, electro Fenton method, membrane-based separation method, photocatalysis, and others that successfully remove phenolic compounds from water. Activated carbon is the most promising adsorbent for numerous contaminants (dyes, metals, etc.). However, low-cost agricultural materials are used to switch to more environmentally friendly ones. This study uses low-cost, eco-friendly adsorbents to remediate biomedical effluents. Pyrolysis of restaurant waste potato peels produced carbon samples. Adsorption-desorption experiments examined pH, temperature, starting drug concentration, contact time, and regeneration ability.

Keywords: adsorption; phenol removal; environmental-friendly adsorbents; potato peels; activated carbons; toxicity; water pollution; removal methods; kinetics

Citation: Prasun, A.; Singh, A.; Kumar, P.A.; Alam, A.; Kumar, A. Removal of Phenol from Biomedical Waste by Adsorption Process. *Eng. Proc.* **2023**, *37*, x. <https://doi.org/10.3390/xxxxx> Published: 17 May 2023



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1. Introduction

Any waste that is produced during the diagnosis, treatment, or immunization of people or animals, or in research relevant to such processes, as well as in the manufacturing or testing thereof, is referred to as bio-medical waste. This includes both solid and liquid waste, as well as their containers and any intermediate products. It has a significant environmental problem such as groundwater contamination [1,2]. Phenolic pollutants are found in wastewater from several industries, including biomedical waste, resin production, high-temperature coal conversion, and petroleum refining [3]. These aromatic hydroxyl compounds are regarded as priority pollutants because they are hazardous at low concentrations, damaging to aquatic life, and proven or probable carcinogens at high doses [5]. Phenol enters the environment through industrial effluents and is removed because it is present in many industrial effluents in extremely high concentrations [6]. Due to phenol's high toxicity and challenging biological degradation, there are strict limits on how much phenol is allowed in the environment [7]. The creation

of materials and efficient procedures that enable the removal of these contaminants from water is crucial because these compounds are poisonous and difficult to degrade [4].

2. Methodology

Biomedical model effluent—Stock aqueous solutions of phenol (500 mg/L) were created for the creation of synthetic biomedical solutions by weighing and dissolving the appropriate quantity of the respective component in distilled water. These solutions were then stored and subjected to spectrophotometric analyses to determine their quality. When used, a stock solution will be diluted to a lower concentration than its original concentration. 0.467 mL of phenol is dissolved in 1000 mL of distilled water to create a phenol stock solution with a 500 ppm concentration, and the stock solution was then diluted to create other concentrations.

Synthesis of modified activated carbons from potato peels—Potato peel would be an advanced green adsorbent since it is rich in high-value elements, particularly polysaccharides, and lignin, which may be burned to produce carbonaceous material. Either Greek cultivars or leftovers from restaurants were used to get potato skins. The potato peels were thoroughly cleaned with distilled water before being baked in an oven for 24 h at 393 K to eliminate dust and other inorganic contaminants [9]. The dry material was crushed, and then a uniform particle size of 0.45-0.15mm was obtained using sieving.

Estimation of Sample—A UV-Visible Spectrophotometer was used to analyze the estimate of phenol. At 270 nm, phenol's maximum wavelength was discovered [10]. To determine different optical densities at various concentrations, the sample was calibrated. The calibrated findings, which display a graphical depiction of the calibration of phenol, are highly useful for determining the specific phenol removal capabilities of various adsorbents. To calculate the amount of phenol absorbed (q_e , mg/g), the following relationship was used [11]:

$$q_e = \frac{C_o - C_e}{M} V$$

where V is the volume of the solution (L), M is the weight of the adsorbent utilized (g), and C_o is the initial concentration of phenol (mg/L). C_e is the equilibrium liquid phase concentration of phenol [8]. The following equation was used to express the percentage of phenol elimination;

$$\text{phenol removal} = \frac{C_o - C_e}{C_o} \times 100$$

where C_e denotes the final concentration and C_o denotes the initial concentration of phenol.

3. Results and Discussion

The phenol sample was calibrated to determine different optical densities at different concentrations. The calibrated results are highly useful for determining each adsorbent's phenol elimination capabilities. The standard stock solution is made in various concentrations, and a spectrophotometer is used to measure the absorbance of each concentration.

Effect of pH—The degree of ionization of the adsorbate during the reaction is influenced by pH, which also affects the solubility of phenol ions in the solution and replaces some of the positive ions present at the active sites. Within the pH range of 2 to 10, the initial pH's impact on phenol's adsorption was determined. At pH 4, the maximum adsorption was found to be 88.07 %. The difference in the concentrations of H^+ and OH^- in the solutions may be the cause of this.

Effect of adsorbent dosage—The dosage of the adsorbent is a crucial parameter since it controls the system's sorbent-sorbate equilibrium as well as the adsorbent's capacity for a particular phenol concentration. Within the adsorbent dosage range of 1–5 g/L, the adsorbent dose's impact on phenol's adsorption was evaluated. Adsorption effectiveness

was shown to rise with increasing adsorbent dosage, however, adsorption volume was found to decrease. The adsorption efficiency did not considerably increase after a particular dosage. Therefore, 2 g/L is chosen as the ideal concentration for subsequent adsorption trials.

Effect of initial concentration—The initial concentration acts as a crucial driving factor to go around all of phenol's mass transfer resistance between the aqueous and solid phases. The initial phenol concentration is controlled between 0.5 and 2.5 mg/L. The starting phenol content was increased up to 1 mg/L, and this modestly improved the equilibrium adsorption efficiency of activated carbon.

Effect of time—The properties of activated carbon and the number of sorption sites it offered influenced how long it took to reach equilibrium. Time is restricted to 15 to 75 min for this experiment. In the first 30 min, significant amounts of phenol were eliminated, and equilibrium was established in 45 min.

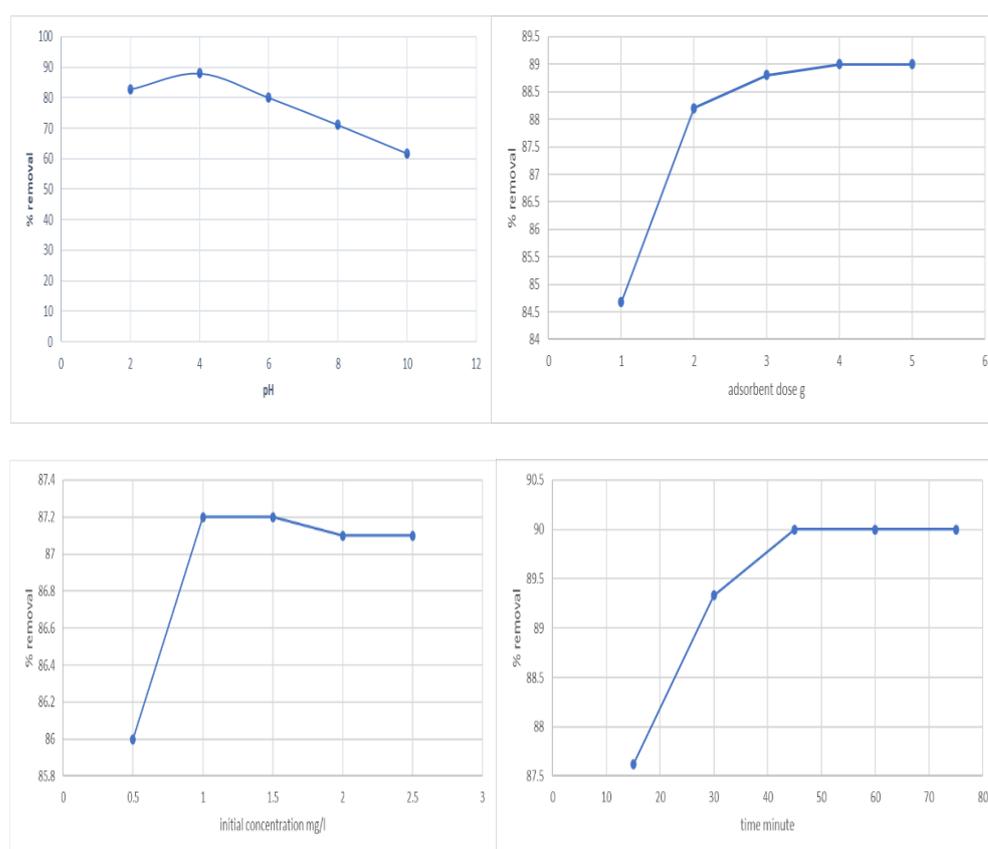


Figure 1. Effect of pH adsorbent dose, initial concentration, and time v/s removal of phenol.

Isotherm—To best utilize adsorbents, it is essential to understand how solutes interact with them by using the term “adsorption isotherm.” The link between the volume of phenol adsorbed and its equilibrium concentration in solutions was modeled by the Langmuir and Freundlich isotherms. Langmuir isotherm is given by the following equation;

$$q_e = \frac{q_o k_L C_e}{1 + k_L C_e}$$

where C_e (mg/L) denotes the equilibrium concentration of the adsorbate, q_e (mg/g) is the quantity of adsorbate adsorbed per unit mass of adsorbent, q_o denotes the Langmuir constants linked to adsorption capacity, and k_L denotes the constant relating to the free energy of adsorption. The following equation can be used to express the Langmuir isotherm in its linearized version;

$$\frac{1}{q_e} = \frac{1}{q_o} + \frac{1}{k_L q_o} \times \frac{1}{C_e}$$

To determine the Langmuir constants, a graph between C_e/q_e and C_e is drawn. The dimensionless separation factor (R_L) can be used to express the key features of the Langmuir isotherm. what is referred to as;

$$R_L = \frac{1}{1 + k_L C_o}$$

where the concentration of phenol is highest in C_o . The value of R_L indicates whether the isotherm is unfavorable ($R_L > 1$), irreversible ($R_L = 0$), linear ($R_L = 1$), or favorable ($0 < R_L < 1$). The graph illustrates a linear relationship between C_e/q_e and C_e based on experimental data, demonstrating the Langmuir model's applicability ($R^2 = 0.999$). The Langmuir isotherm is favorable for the adsorption of phenol onto the activated carbon, as demonstrated by the value of R_L , which was discovered to be 0.00678.

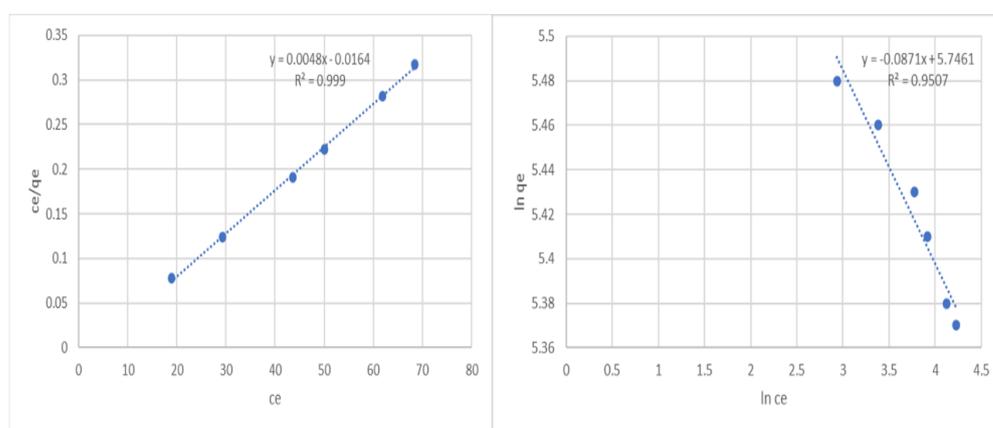


Figure 2. Langmuir and Freundlich adsorption isotherm of phenol onto an adsorbent.

Freundlich isotherm is given by the following equation;

$$q_e = K_f C_e^{\frac{1}{n}}$$

where $(1/n)$ is a measure of the adsorption intensity and K_f generally represents the adsorption capacity. The Freundlich isotherm model's linearized form is;

$$\ln q_e = \ln K_f + \frac{1}{n} \ln C_e$$

A straight line with a slope of $1/n$ and an intercept of $\ln K_f$ is produced by plotting $\ln q_e$ versus $\ln C_e$. The size of the exponent n determines how favorable adsorption is. According to conventional wisdom, n numbers between 2 and 10 indicate favorable, 1-2 highly challenging, and less than 1 undesirable adsorption quality. This n value of 11.48 indicates good adsorption properties. The R^2 value, 0.9507, is lower than the Langmuir isotherm model's value. The Freundlich isotherm is therefore not the ideal isotherm model to describe the phenol adsorption mechanism.

Kinetics studies – The rate of the adsorption process and potential rate-controlling phase are examined using kinetics models. These investigations provide information on the rate of change of the reaction. Pseudo-first-order and pseudo-second-order models have been used to analyze the kinetic data gathered from studies. The conformance between the kinetic models and the experimental data was expressed using the correlation coefficient (R^2).

The pseudo-first-order model of Lagergren has been widely used to predict the phenol kinetics and its formula is given as;

$$\frac{dq_t}{dt} = K_1(q_e - q_t)$$

where K_1 is the equilibrium rate constant of pseudo first order adsorption (1/h), q_e denotes the quantity of phenol adsorbed (mg/g) at equilibrium, and q_t is the amount of phenol adsorbed (mg/g) at any time. The linearized form of the equation is given as;

$$\log(q_e - q_t) = \log q_e - \frac{K_1 t}{2.303}$$

The slope and intercept can be calculated from the plot of $\log(q_e - q_t)$ over time, which is typically used to calculate the equilibrium adsorption density q_e and first-order rate constants K_1 and R_2 . It was discovered that there was a significant disparity between the calculated (q_e , cal) and experimental (q_e , exp) adsorption capacity values and that the R_2 value derived for the pseudo-first-order kinetic model exhibits a consistent trend. The computed q_e is substantially lower than the experimental q_e (225mg/g), at 25.45 mg/g.

The behavior is predicted by the pseudo-second-order model for the whole adsorption range. It is consistent with the rate-controlling step being an adsorption mechanism.

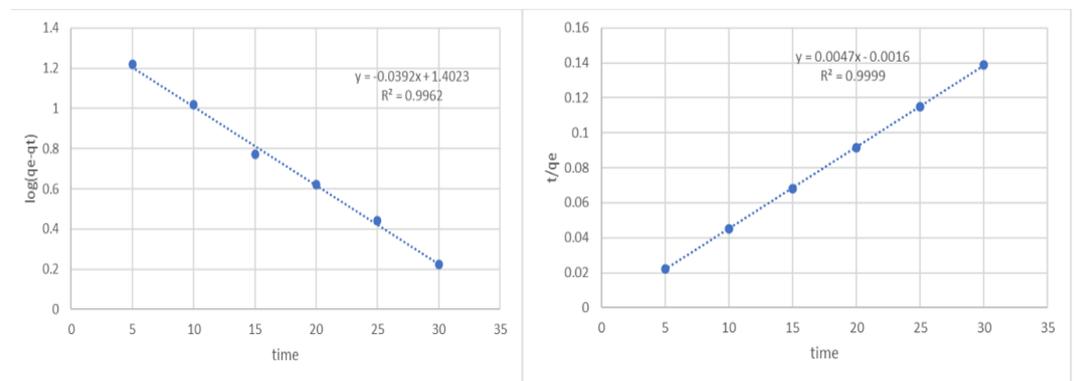


Figure 3. Pseudo first-order and second-order kinetics of phenol on potato peel adsorbent.

The equation is as follows;

$$\frac{dq_t}{dt} = K_2(q_e - q_t)^2$$

K_2 is the equilibrium rate constant of pseudo-second-order adsorption. The linearized form of the equation is given as;

$$\frac{t}{q_t} = \frac{1}{K_2 q_e^2} + \frac{1}{q_e} t$$

The values of q_e and the second-order rate constant K_2 were determined using the slope and intercept of the plot of t/q_t vs t , respectively. The linear relationship between the plot of t/q_t versus t indicates that second-order kinetics is appropriate in this situation. As the initial phenol concentration rose, the rate constant generally dropped. The second-order kinetic model’s correlation coefficient was higher than 0.99. The experimental findings (225 mg/g) and the value computed by the q_e (212.76 mg/g) do correspond extremely well.

4. Conclusions

The elimination of phenol from biomedical waste using potato peel has been proven to be effective (removal efficiency 97%). The Phenol-activated carbon system performed best when the time $t = 45$ min, $pH = 4$, starting concentration $C_0 = 1$ mg/L, and adsorbent dose $m = 2$ g/L were met. The adsorption capacity of activated carbon significantly increased as the pH of the solution decreased. The results obtained demonstrated that activated carbon has a high capacity for phenol adsorption from biomedical waste. Activated carbon has a maximum adsorption capacity of 225 mg/g. The kinetics study’s findings show that the pseudo-second-order kinetics may accurately model the

adsorption of phenol on a solution of biomedical waste. Because it demonstrates linear regression of the experimental data and suggests monolayer adsorption of phenol on the surface of activated carbon, over-equilibrium model analysis shows the fitness of the Langmuir isotherm model to the Phenol-activated carbon adsorption system. Because of its accessibility, affordability, high surface area, and adsorption capacity, activated carbon generated from the chemical activation of potato peel appears to be an efficient adsorbent for the removal of phenol from biomedical waste.

Author Contributions: Conceptualization, methodology, experiments, writing—original draft preparation, review and editing had done by A.P., A.K., A.A. and A.S., P.A.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

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