





1

2

3

4

5

6

7

8

Evaluation of the safety of immobilized microorganisms Lysobacter sp. on inorganic media ⁺ Galina Shaidorova^{1*}, Alexander Vesentsev², Ulyana Krut³, Elena Kuzubova⁴, Alexandra Radchenko⁵ and Marina Potapova⁶

123,4,5,6 Belgorod State University, Russia, Belgorod, 85 Pobedy Street, 308015, 1015artek1015@mail.ru

Correspondence: shaydorova@bsu.edu.ru; Tel.: +7-915-528-73-50.

+ Presented at the title, place, and date.

Abstract: It is known that the immobilization of microorganisms on carriers of various nature in-9 creases their safety. Inorganic matrices: sodium carboxymethyl cellulose technical brand "KMC 10 85/500"; colloidal silicon dioxide in the form of a commercial preparation "Polysorb"; sodium form 11 of montmorillonite from the Podgorenskoye deposit, Voronezh region. Bacterial cells were immo-12 bilized by adding Lysobacter sp. solid sterile carrier with constant mechanical stirring in the ratio 13 "carrier:biomass", equal to 1:(2-4). During the experiment, it was found that the mineral montmo-14 rillonite is a promising material for the immobilization of bacterial cells in order to obtain bio-15 compositions based on them, since a positive trend in the preservation of bacterial cells was re-16 vealed. 17

Keywords: inorganic matrices; immobilization; microorganisms; cell safety

20

21

22

23

26

36

37

18

Citation: Shaidorova, G.; Vesentsev, A.; Krut, U.; Kuzubova, E.; Radchenko, A.; Potapova, M. Evaluation of the safety of immobilized microorganisms *Lysobacter sp.* on inorganic media. **2023**, *5*, x. https://doi.org/10.3390/xxxxx Published: 18 May

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Eithingh of each of the literinors. Submitted of proposition of the terms and pendising faile Creative Commons Attribution (CC BY) license

Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/).

1. Introduction

Currently, a promising direction in biomedicine is the creation of biofilms and compositions based on microorganisms-antagonists of pathogenic microflora on carriers of various nature [1-3].

The aim of the study was to identify a rational inorganic carrier for the immobiliza-tion of *Lysobacter sp.* 25

2. Materials and Methods

As a model bacterial culture for immobilization, a *Lysobacter* culture isolated from 27 the soil of the city of Belgorod using classical biotechnology methods with confirmation 28 of generic affiliation by 16S rRNA sequencing (1484 nucleotides) was used, as a result of 29 which a unique nucleotide sequence of the strain among those presented in GenBank 30 was revealed: the maximum percentage of similarity observed with strain *L*-43 31 (MT229166.1) and *Lysobacter enzymogenes* M497-1 (AP014940.1) - 99.7% each. 32

As a growth substrate, a liquid nutrient medium containing 0.2 wt% casein and 0.1 33 wt% yeast extract was used; T = 30°C; log phase - 24 hours. Metabolic products: chitosan, 34 beta-1,4-glucanase, protease. 35

To study the effect of immobilization of microorganisms and their enzymes, solid carriers were used:

1. Sodium carboxymethyl cellulose (NaCMC) technical brand "KMC 85/500" pro-38duced by LLC "Davos-Trading". TU 2231-001-53535770-2010 (with change No. 1.2): de-39gree of substitution for carboxymethyl groups 80-90; degree of polymerization 500-550;40pH value (pH) of an aqueous solution with a mass fraction of CMC 1% in the range of418-12; dynamic viscosity of a 2% CMC solution at a temperature of 25 °C mPa*s according42to the Brookfield method, not less than 100.43

2. Colloidal silicon dioxide in the form of a commercial preparation "Polysorb" 1 produced by JSC "Polysorb". Polysorb MP (medical oral) is an inorganic, non-selective, 2 multifunctional enterosorbent based on highly dispersed silica with particle sizes up to 3 0.09 mm, with the chemical formula SiO₂. The sorption capacity of the drug for internal 4 use is 300 m²/g. 5

3. Sodium form of montmorillonite (NaMMT) from the Podgorenskoye deposit, 6 Voronezh region [4, 5], obtained by introducing soda ash (4 wt%) into a native rock sus-7 pension (5 wt%), followed by sedimentation enrichment and drying (t=95±3°C). The 8 quantitative content of montmorillonite, determined according to GOST 28177-89 -9 79.35±0.14 wt.%. As a result of the modification, the crystal lattice parameters changed: 10 for the native form of Ca-montmorillonite a = 5.16 Å, b = 8.94 Å, c = 15.02 Å; for modified 11 montmorillonite a = 5.22 Å, b = 9.04 Å, c = 13.82 Å. Specific surface 60 m²/g, specific pore 12 volume 0.083 cm³/g; average pore size 55.5Å. 13

The immobilization of bacterial cells was carried out by adding *Lysobacter sp.* in the 14 logarithmic phase of growth, a solid sterile carrier with constant mechanical stirring in 15 the ratio "carrier : biomass", equal to 1:(2-4), at a temperature of 30°C; the mixture was 16 thoroughly mixed for at least 40 minutes, frozen at minus 40°C and then freeze-dried at 17 minus 40-45°C for 24 hours to a level of 3-7% moisture content of the composition. The 18 dry compositions obtained were then stored in sterile flacons at room temperature. 19

Survival after immobilization of microorganisms *Lysobacter sp.* on solid carriers was 20 determined by the Pour Plate method, in which the samples were suspended in a Petri 21 dish using molten agar cooled to about 40-45°C (just above the solidification point to 22 minimize heat-induced cell death). After the nutrient agar solidified, the plates were in-23 cubated for 24 hours and the number of colony forming units (CFU) was determined by 24 the serial dilution method. 25

The degree of preservation (α , %) was determined by the formula:

$$\alpha = \left(1 - \left(\frac{CFU_{ref} - CFU_n}{CFU_{ref}}\right) \times 100\%\right),\tag{1}$$

where: CFU_{ref} - the number of colony-forming units in the biocomposition immediately after immobilization; 28

 CFU_n - the number of colony-forming units in the biocomposition after storage on 29 the n-th day. 30

3. Results

The results of assessing the viability of immobilized *Lysobacter sp.* are presented in 32 Table 1, and Figure 1 shows the dynamics of the preservation of bacterial culture. 33

Table 1. Viability of immobilized Lysobacter cells.

Biomass : carrier ratio	CFU * (g/l) after storage			
	Day 2	Day 15	Day 31	Day 92
Freeze culture	2,4±0,04 · 10 ⁵	2,3±0,06 ·	2,0±0,04 ·	0,9±0,05 · 10 ⁵
	105	105	105	10°
NaCMC 1:2	$3,4\pm0,06$ ·	$3,7{\pm}0,02\cdot$	$3,9{\pm}0,05{}$	2,1±0,02 ·
	105	105	105	105
NaCMC 1:3	$3,5\pm0,04$ ·	3,8±0,03 ·	4,1±0,07 \cdot	2,7±0,07 ·
	105	105	105	105
NaCMC 1:4	3,3±0,07 ·	3,5±0,03 ·	$3,6\pm 0,05$ ·	1,9±0,03 ·
	105	105	105	10^{5}

33 34

31

26

Polysorb 1:2	3,5±0,02 ·	3,2±0,02 ·	2,7±0,01 ·	1,8±0,02 ·
	105	105	105	105
Polysorb 1:3	3,2±0,03 ·	3,1±0,01 ·	2,5±0,05 ·	1,7±0,01 ·
	105	105	105	105
Polysorb 1:4	3,3±0,03 ·	$3,1\pm0,05$ ·	2,4±0,07 ·	1,5±0,04 ·
	105	105	105	105
NaMMT 1:2	3,6±0,03 ·	3,8±0,02 ·	4,0±0,04 ·	4,2±0,06 ·
	105	105	105	105
NaMMT 1:3	3,5±0,05 ·	3,7±0,06 ·	3,9±0,07 ·	4,1±0,03 ·
	105	105	105	105
NaMMT 1:4	3,4±0,01 ·1	3,8±0,04 ·	4,0±0,06 ·	4,1±0,03 ·
	05	105	105	105

*Std. Deviation



Figure 1. Dynamics of preservation of the bacterial culture Lysobacter sp.

After three months (92 days) of storage of lyophilizates, the following results were obtained:

1. Lyophilization of the bacterial culture of Lysobacter sp. without immobilization on the matrix leads to a decrease in safety to 37.5%.

2. Cell immobilization on sodium carboxymethyl cellulose allows to increase safety up to 65%; on colloidal silicon dioxide (Polysorb) - up to 50%.

3. When immobilized on the mineral montmorillonite, not only the preservation of microorganisms is manifested, but also an increase in the number of cells by 18%.

 $3 \ of \ 4$

 3.

	4. Conclusions	1		
	Thus, a positive trend in the preservation of bacterial cells during immobilization on	2		
	solid carriers was revealed.	3		
	It has been established that the most effective matrix for immobilization of Lysobacter	4		
	<i>sp.</i> is the sodium form of montmorillonite.	5		
	The obtained research results can be used to create biocompositions based on bacte-	6		
	rial cultures for various purposes.	7		
		8		
	Funding: The work was carried out within the framework of the state task FZWG-2023-0007.	9		
	Adaptive reactions of microorganisms: theoretical and applied aspects.	10		
Ref	ferences	11		
		12		
1.	Efremenko, E.N. Immobilized cells: biocatalysts and processes. M.: RIOR, 2018; 499 p.	13		
2.	Milivojevic, M.; Pajic-Lijakovic, I.; Bugarski, B.; Levic, S.; Nedovic, V. Alginic acid: sources, modifications and main applica-	14		
	tions. Alginic Acid: Chemical Structure 2015, pp. 45-88.			
3.	Niyazbekova, Z.T.; Nagmetova, G.Z., Kurmanbayev A.A. An overview of bacterial cellulose applications. Eurasian Journal of	16		
	Applied Biotechnology 2018 , № 2. pp. 17-25.	17		
4.	Shaidorova, G.M.; Vezentsev, A.I.; Trufanov D.A. Obtaining the sodium form of clays from the Podgorenskoe deposit of the	18		
	Voronezh region. Bulletin of the Technological University 2022, V.25. No. 11. pp. 101-105.	19		
	https://doi.org/10.55421/1998-7072_2022_25_11_101	20		
5.	Shaidorova, G.M.; Vezentsev, A.I.; Trufanov, D.A.; Sokolovsky, P.V. Sorption activity of sodium-modified bentonite-like clays	21		
	of the Podgorensky deposit of the Voronezh region. Actual physical and chemical problems of adsorption and synthesis of	22		
	nanoporous materials: All-Russian symposium with international participation, dedicated to the memory of corr. RAS V.A.	23		
	Avramenko 2022, Moscow. Russia. Collection of proceedings of the symposium. M.: IFCHE RAN. pp. 177-179.	24		