

Proceedings



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Assessing the Likelihood of *Staphylococcus aureus* Contamination in Bottled Drinking Water Production⁺

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Abstract: This study's objectives were to evaluate the possibility of S. aureus contamination in bot-20 tled drinking water and to determine the elements that affected the level of *S. aureus* in raw water. 21 In two drinking water treatment facilities, samples of raw water, soft water, reverse osmosis (R.O.) 22 water, and finished water were taken at various stages. In addition, raw water samples and risk 23 indicators like pH, temperature, and residual chlorine were gathered at the packaging facility 24 during the washing process. For Factory A (small scale), the pH values for the raw water, soft wa-25 ter, R.O. water, and finished water samples were 7.17, 7.24, 6.69, and 5.92, respectively. For Factory 26 B (medium scale), the pH values were 7.9, 7.44, 6.97, and 6.8. All water samples from Factory A (2 27 CFU/ml) and Factory B (1 CFU/ml) had S. aureus concentrations that were within the acceptable 28 range for human consumption. All water samples from Factory A (2-26 CFU/mL) and Factory B 29 (11-316 CFU/mL) contained total coliforms as well. Our study revealed that S. aureus contamination 30 in water was mostly caused by the pH and processing time. To prevent pathogen contamination in 31 bottled drinking water, it is recommended that raw and finished water be kept at a pH level be-32 tween 6.5 and 7.5. 33

Keywords: Risk Estimation; Food Risk Assessment; Sensitivity Analysis

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1. Introduction

About 50–70% of a person's total weight is made up of water, which is essential for 37 maintaining healthy cells, tissues, and organs (EPA, 2018) [1]. The source of drinking 38 water may contain a variety of radionuclides, microorganisms, and chemicals. These 39 dangers may result in gastrointestinal disorders, brain system or reproductive issues, as 40 well as long-term conditions like cancer. Typhoid fever and cholera are examples of 41 bacteria that cause waterborne disease. Diarrhea is one of the most typical negative 42 effects of contaminated water and can be spread by drinking it [2]. One of the most 43 frequent causes of food poisoning, Staphylococcus aureus, produces enterotoxin from an 44 initial concentration of 100 organisms per milliliter of water. This pathogen has been 45 found in a variety of places, including human nasal passages, skin, clothing, food, flies' 46

digestive tracts, dust, and moisture droplets. As a proven source of their origin in 1 drinking water supplies, S. aureus can be discovered in rural drinking water. Residents 2 exposed to contaminated water may become colonized by S. aureus from drinking water. 3 [3]. Individuals are exposed to avoidable health risks when water and sanitation systems 4 are improperly managed. According to some studies, the prevalence of Escherichia coli 5 and Staphylococcus aureus was 54.17% and 16.67%, respectively, with an average con-6 centration of 1.04 log CFU/ml and 0.26 log CFU/ml in the drinking water filtration dis-7 penser toll machines water in the Mahasarakham province of Thailand [4]. The charac-8 teristics of the contaminants in the water source determine the treatment procedure for 9 disinfecting drinking water to decrease the prevalence of waterborne diseases [1]. Re-10 verse osmosis (R.O.), charcoal filtration, and UV light are used to treat the water. Ac-11 cording to the [5], R.O. can eliminate 99% of the germs in tap water as well as organic and 12 inorganic pollutants. Drinking water may become quickly re-contaminated by other 13 factors, particularly poor process hygiene brought on by erratic maintenance [6]. This 14 can result in the contamination of the water with hazardous chemicals and dangerous 15 microbes. In order to establish the high-risk parameters of processing steps as part of risk 16 assessment, probabilistic modeling and sensitivity analysis of crucial control points in 17 food processing and food safety systems are used [7]. Risk assessors can also utilize these 18 models to help them make management decisions that will lower the risk of contracting 19 foodborne illnesses [8]. In order to clarify the current state of the risk parameters for 20 bottled drinking, it is required to apply modeling of the probability of S. aureus in terms 21 of risk assessment of drinking water. 22

2. Materials and Methods

2.1. Water Sample

2.1.1. Sample collection

The physicochemical and microbiological tests were performed on water samples 26 from one packing house (n = 5) and bottled drinking water (n = 3 at each step) from two 27 factories (Factory A represents a small-scale factory and Factory B represents a medi-28 um-scale plant). The bottled drinking water processing facility took samples of raw wa-29 ter, soft water, reverse osmosis water, and finished water. The samples were maintained 30 in iceboxes after sample collection and transported to the lab (Department of Medical, 31 Regional Medical Medical Sciences Center5, Samut Songkhram) within two hours. In 32 triplicate, each sample was taken and examined. 33

2.2. Physicochemical analysis

The AOAC official technique was used for the physicochemical evaluations of all water samples (AOAC, 2005) [9]. The packing house gathered and evaluated the water sample's pH, residual chlorine (ppm), temperature (°C), and processing time. Using a pH 37 meter (Hanna Instruments, USA), water samples from bottled drinking water were ana-38 lyzed to determine the pH at each stage. 39

2.3. Microbiological analysis

All samples were collected for measurement of the quantification and risk of S. au-41 reus and total coliforms by following the FDA BAM, 2019 (Chapter 12) 10] and FDA BAM 42 (Appendix 2) [11] procedures. 43

2.3.1. Plating media method of S. aureus and Total Coliforms

A quantity of 25 ml of samples was measured, rinsed in a 250 mL bottle containing 45 225 mL of sterile 0.1% peptone water, and then the suspension diluted from 1:10-2 to 46 1:10-4. A volume of 0.1 mL of each dilution was spread on Braid Parker agar plates 47 (BPA) (Merck, Germany) for the enumeration of *S. aureus* and Five tubes should be used 48for each dilution of the multiple-dilution MPN series used to enumerate of total coliforms 49

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using Violet Red Bile Agar (VRBA) (Merck, Germany). The plates were incubated at 37°C for 24 h for isolation of bacteria (FDA BAM 2019 [10] and FDA BAM 2020 [12]). The colony-forming units of characteristic S. aureus and total coliforms were counted and reported as CFU.

2.4. Sensitivity analysis

To explain how *S. aureus* became contaminated during the various processing steps, a probabilistic risk assessment (quantitative model) was developed. Using the analysis 7 tool program in Microsoft ExcelTM, the results of the microbiological analysis were 8 transformed into log10 units and put through a straightforward regression analysis. It 9 often entails fitting a relationship between inputs and outputs, such as the following 10 linear one [13]. The equation is as follows: 11

$$Yi = bo + b1X1, i + b2X2, i + ... + bmXm, i + ei$$
 (1)

Yi is the ith output data point for the ith input data point, Xj,i is the ith input data 12 point for the jth input, bj is the jth input's regression coefficient, and ei is the ith data 13 point's error. The basis function for each term in the regression model may be distinct 14and either linear or nonlinear. According to Frey et al. (2003) [13], the regression coeffi-15 cient, bj, for a linear model can be understood as the change in the output Yi when the 16 17 input Xj,i for a given value of j increases by one unit and the values of all other inputs remain constant. Rank order correlations, a non-parametric statistic for estimating the 18 correlational link between two means, were employed in the regression sensitivity anal-19 ysis based on Spearman's rank correlation calculations The analysis involved running 20 simulations with probability distributions allocated to the inputs and determining the 21 impact of input variance on the output distribution. Then, tornado graphs might be 22 produced. The various input factors were represented by horizontal bars on the graph, 23 and the length of the bars indicated the degree of association with the mean numbers and 24 the amount of S. aureus discovered in the samples (output variables). The nominal rage 25 sensitivity analysis equation was as follows [13]: 26

Sensitivity =	Output at maximum in	put value - Out	put at minimum in	put value (2)

Output at nominal inp	out value
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2.5. Statistical analysis

All measurements were carried out in triplicate. The means and standard deviation 29 (± SD) were calculated using Data Analysis in the Excel platform. 30

3. Results

3.1. Physicochemical and microbiological properties of the water samples

Table 1 displays the conditional factors for the bottling plant (Plant B) and the 33 packing facility (Plant A). The packing house had water with pH values between 10.33 34 and 10.81, residual chlorine concentrations between 0.23 and 1.67 ppm, and temperatures 35 between 17.3 and 19.1°C. In order to generate the simulation models, the quantity of S. 36 aureus (3.40 LogCFU) discovered in the water samples was evaluated. Raw, soft, R.O., 37 and finished water had pH values of 7.17, 7.24, 6.69, and 5.92, respectively, for plant A. 38 For plant B, the corresponding values were 7.60, 7.44, 6.97, and 6.80. S. aureus concen-39 trations at Plant A and Plant B ranged from 1-3 CFU. Both Factory A (2-26 CFU/mL) and 40Factory B (11-316 CFU/mL) provided water samples that also included total coliforms. 41

Table 1. Physicochemical	l and microbiologica	il analysis results c	of water sample.

Plants	Packing House	Drinking Water Factory A			Drinking Water Factory B				
Samples	Water	Raw	Soft	R.O.	Finished	Raw	Soft	R.O.	Finished
Time (min)	0-20	-	-	-	-	-	-	-	-

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Temperature (°C) pH	17.3–19.1 10.33–10.81	- 7.17±0.25	- 7.24±0.26	- 6.69±0.21	- 5.92±0.32	- 7.60±0.33	- 7.44±0.29	- 6.97±0.22	- 6.80±0.21
Residual chlorine (ppm)	0.23-1.67	-	-	-	-	-	-	-	-
Total coliform (CFU)	-	189 ± 8.60	316±44.22	30±44.40	145±7.72	26±3.89	2±3.13	20±1.48	13±1.47
S. aureus (CFU)	3.40 LogCFU	3±1.00	1 ± 0.00	1 ± 0.00	1 ± 0.00	1 ± 0.00	1 ± 0.00	1 ± 0.00	1 ± 0.00
Nute The second second CD (second second sec									

Note: The numbers are means±S.D. from three independent replicates.

3.2. Probabilistic distribution

The pH of the water and the concentration of S. aureus and total coliforms in the water of Plants A and B were simulated to have probability distribution values of Nor-4 mal(1.5,1.0) and Normal(1,0) and Normal(163.09,89.10) and Normal(13.97,6.70), respec-5 tively. Water from Plants A and B had normal pH distributions of 6.75, 0.61 and 7.20, 6 0.38, respectively. The concentration of S. aureus and total coliforms in water samples of 7 factory A and B was in the range of 0-4 CFU and 0-2 CFU and 0-316 CFU and 0-30 CFU, 8 respectively, with the highest probabilities of S. aureus and total coliforms contamination 9 in the water of Plant A and Plant B being 0.42 and 0.35 and 4.47E-03 and 0.0514, respec-10 tively. The pH values in the water of Plants A and B had the maximum probabilities of 11 0.6 at pH ranging from 5-8 and 1 at pH ranging from 6-8, respectively. 12





Figure 1. (a) The probability distribution functions of S. aureus concentrations in water (b) The13probability distribution functions of total coliforms concentrations in water samples. (c) The14probability distribution functions of pH values of water samples.15

3.3. Modeling with multiple regression equations

The sensitivity analysis of water was developed using the equation for multiple regression, and the equations are as follows for the multiple regression of S. aureus contamination in water (eq. 3) and S. aureus contamination in bottled drinking water (eq. 4): 19

$$Y = -233.9910 + (1.8471 \times \text{Time}) - (9.1343 \times \text{Temperature of Wa-}$$
(3)
ter)-(5.0746 × Residual Chlorine in Water)+(40.5970 × pH of Water)

$$Y = -0.1260 + (0.1972 \times pH \text{ of Water})$$
(4)

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The model's input parameters included the water's temperature, pH, residual chlo-1 rine content, and temperature. The model's result was the detection of S. aureus water 2 contamination. 3

3.4. Sensitivity Analysis

The Excel Platform was used to do the sensitivity analysis. For each input variable, 5 horizontal bars on a tornado chart were drawn, with the length of the bars indicating the 6 strength of the association with the output variable. 7



Figure 2. Sensitivity analysis of the risk factors of S. aureus contaminated in water.

The sensitivity analysis results (Figure 2) show that the temperature of the water 10 and residual chlorine concentration were the conditional parameters affecting a decrease 11 in the number of *S. aureus* in the water and an increase in the concentration of *S. aureus* in 12 the water. Water temperature and residual chlorine concentration were the main intervention measures that helped control the effect on the water. 14

4. Discussion

To assess the health risk of drinking water, the study of Wibuloutai and coleagues, 16 which assesses the microbiological quality of drinking water produced by drinking water 17 filtration dispenser toll machines (DFTMs) was compared to this study. 210 samples in 18 total were chosen at random from 70 DFTMs that were spaced 500 meters apart from 19 Mahasarakham University. The DFTM water had high levels of Escherichia coli and 20 Staphylococcus aureus, with prevalence rates of 54.17% and 16.67% and average values 21 of 1.04 log cfu/ml and 0.26 log cfu/ml, respectively. The risk assessment used the @Risk 22 tool to calculate the likelihood of exposure as 1.67 E-01 and the odds of getting sick from 23 S. aureus and E. coli as 2.08 E-03 and 1.58, respectively (@Risk is a software tool to gen-24 erate a distribution of possible outcomes and estimate the probability of different levels 25 of pathogen exposure). [4] It has been established that the findings of our investigation 26 may be safely consumed. The suggested interventions keep the water's pH between 6.5 to 27 7.5 since doing so could help to lessen the amount of microbial contamination in the 28 water [15]. According to Pratum and Khananthai (2017) [14], the quality of drinking wa-29 ter also depends on the tap water or water source used throughout the manufacturing 30 process. The local government must issue local laws for the control of such businesses in 31 the area of responsibility in accordance with the Ministry of Public Health's notification 32 on hazardous health activities [4]. 33

5. Conclusions

This study was successful in identifying the potential risk of S. aureus contamination 35 in water as well as the recommended pH levels for processing bottled drinking water at 36 small- and medium-sized facilities. The findings of the sensitivity study revealed that the 37 pH of the water samples played a significant role in the procedure. This result implies 38 that preserving the pH of the water throughout the procedure, as shown by this study, is 39

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a helpful technique for lowering the risk of contamination. However, harmful bacteria 1 have the potential to taint finished goods if the water quality is inadequate. 2

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