





1

2

3

4

5

6 7

8

9

10

11

12

13

14

24

25

# An insight into chayote (*Sechium edule*) peels valorization: phytochemical characterization and bioactive potential <sup>+</sup>

Elsa F. Vieira <sup>1,\*</sup>, Manuela M. Moreira <sup>1</sup>, Rebeca Cruz <sup>2</sup>, Aline Boatto da Silva <sup>2</sup>, Susana Casal <sup>2</sup> and Cristina Delerue-Matos <sup>1</sup>

REQUIMTE/LAQV, Polytechnic of Porto—School of Engineering (ISEP/IPP), Rua Doctor António Bernardino de Almeida, 4249-015 Porto, Portugal; elsa.vieira@graq.isep.ipp.pt; manuela.moreira@graq.isep.pt;
REQUIMTE/LAQV, Laboratório de Bromatologia e Hidrologia, Faculdade de Farmácia da Universidade

do Porto, Rua de Jorge Viterbo Ferreira 228, 4050-313 Porto, Portugal; rcruz@ff.up.pt<u>.</u> alineboatosilva@gmail.com<u>;</u> sucasal@ff.up.pt

- \* Correspondence: elsa.vieira@graq.isep.pt; Tel.: +351-228340500
- + Presented at the The 4th International Electronic Conference on Foods, 15–30 October 2023; Available online: https://foods2023.sciforum.net/.

Abstract: A Box-Behnken Design was applied to investigate the influence of ethanol %, time, temperature, and ultrasonic power on the Ultrasound-Assisted Extraction (UAE) of phenolic compounds, carotenoids, and antioxidant capacity from chayote peel. The recovery of total phenolics15(406 mg GAE/100 g dw) and antioxidant compounds (FRAP value of 82.83 mg AAE/100 g dw and18ABTS value of 319 mg AAE/100 g dw) were maximized using 37% ethanol, 55 °C and 224 W, for 3019min. The extraction of carotenoids (17.14 mg/100 g dw) was maximized using 75% ethanol, 30 °C20and 200 W, for 61 min.21

Keywords: Sechium edule; chayote peel; ultrasound-assisted extraction (UAE); Box-Behnken Design22(BDD); phenolics and carotenoids profile23

# 1. Introduction

Chayote, scientifically known as Sechium edule (Jacq.) Swartz, is an underutilized veg-26 etable in many regions, including Portugal. It is a member of the Cucurbitaceae family, 27 which includes cucumbers, pumpkins, and squashes. Chayote is known for its interesting 28 nutritional and phytochemical composition, and it has been associated with various po-29 tential health benefits [1]. Literature reports that chayote peels extracted with 100% water 30 have a Total Phenolic Content (TPC) of 746.46 ± 58.73 mg GAE/ 100 g dry weight (dw), 31 mostly represented by the phenolic acids (e.g., caffeic acid, phenylacetic acid, 4-hy-32 droxybenzoic acid, protocatechuic aldehyde and dihydroxybenzoic acid isomer IV) and 33 flavonoids (Hispidulin, apigenin 7-O-apiosyl-glucoside, apigenin, chrysoeriol 7-O-api-34 osyl-glucoside and neohesperidin) [2]. Chayote peels are also rich in carotenoids (1.7 mg/ 35 100 g dw), with a  $\beta$ -carotene content of 0.36 mg/ 100 g dw [3]. Hence, the application of 36 green-extraction techniques to recover these valuable bioactive compounds and further 37 investigation on the phytochemical profile of chayote peels is required to promote its sus-38 tainable use in food, cosmetic and pharmaceutical sectors. 39

This study aimed to determine the optimal ultrasound-assisted extraction (UAE) 40 conditions for recovering carotenoids, phenolic compounds, and antioxidant capacity 41 from chayote peel. A response surface methodology using the Box-Behnken Design (BDD) 42 was employed to investigate the impact of ethanol percentage, extraction time, temperature, and ultrasonic power on the recovery of these valuable compounds. 44

2. Material and Methods

45

**Citation:** To be added by editorial staff during production.

Academic Editor: Firstname Lastname

Published: date



**Copyright:** © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/).

1

8

# 2.1. Sampling

Samples of chayote green variety at maturity stage were supplied by a local farm 2 located at Cinfães, Douro (Portugal), and were collected in October 2021 from 10 plants 3 (random sampling) to obtain a representative set of fruits. Then, peels were separated 4 from pulp, dried for 18 h at 52 °C in processed food (Excalibur 9 Tray Dehydrator, Model 5 4926 T, USA), grounded (Moulinex A320), sieved through 0.75 mm stainless steel sieve, 6 thoroughly mixed and stored at 8 °C under light-free conditions until extractions. 7

#### 2.2. BBD Optimization and Validation

The optimization of UAE of phenolics and carotenoids from chayote peels was done 9 by Box-Behnken Design (BBD). Four independent variables were considered: X1 - 25-75% 10 of ethanol; X2 - 30-80 minutes; X3 - 35-55°C; and X4 - 60-80% ultrasonic power amplitude. 11 Total phenolic content, TPC (Y1); Ferric reduction antioxidant power, FRAP (Y3); 2,2'-az-12 ino-bis (3-ethylbenzothiazoline-6-sulphonic acid radical scavenging activity, ABTS (Y2) 13 and total carotenoid content, TC (Y4), were taken as the dependent variables (Table 1). 14 Desirability indices were constructed to obtain the optimum experimental conditions to 15 maximize the bioactivities of chayote peel. 16

## 2.3. Ultrasound Assisted Extraction (UAE)

For UAE procedures, 1 g of freeze-dried powder of chayote peels were mixed with 18 30 mL extraction solvent in a 3.5 cm inner diameter cylindrical flask. After that, the flask 19 was covered with aluminum foil and placed in the ultrasonic bath (Bandelin SON-20 OREXTM Digital 10 P Ultrasonic baths DK 102 P, Bandelin Electronic GmbH, Berlin, Ger-21 many). The extraction was then carried at conditions defined by the BBD (Table 1). All 22 extractions were performed in triplicate, using a solid to sample ratio of 1:30 g/mL, with 23 occasional stirring. The obtained extracts were filtered, centrifuged (5000 rpm for 15 min 24 at 4 °C), lyophilized for 48 h and stored at 4 °C until further use. 25

## 2.4. Characterization of Chayote Peel Extracts

The total phenolic content (TPC), total carotenoid content (TC), and antioxidant ac-27 tivity evaluated by FRAP and ABTS assays were performed as previously described [4]. 28 TPC results were expressed as milligrams of gallic acid equivalents (GAE) per 100 grams 29 of dry weight (dw); TC results as mg/ 100 g dw; and FRAP and ABTS results as mg of 30 ascorbic acid equivalents (AAE)/ 100 g dw. 31

#### 2.5. HPLC-PDA Polyphenol Composition Profile

The phenolic profile of the optimal extract was characterized by HPLC with a photo-33 diode array detector and a C18 column as previously described [4]. The extract was ana-34 lyzed three times, and the results were expressed as mg/ 100 g dw. 35

#### 2.6. HPLC Vitamin A, Vitamin E, Carotenoids, and Chlorophylls Composition Profile

The vitamin A, vitamin E, carotenoids, and chlorophylls composition profile of the 37 optimal UAE-chayote peel extract was characterized by HPLC with a photodiode array 38 detector and a C18 column as previously described [4]. The extract was analyzed three times, and the results were expressed as mg/ 100 g dw. 40

#### 2.7. Statistical Analysis

Results were expressed as means ± standard deviation. Design-Expert software ver-42 sion 7.0 (Stat-Ease Inc., Minneapolis, MN, USA) was used for establishing the experi-43 mental design of the optimization process. IBM SPSS Statistics 22.0 software (SPSS Inc., 44 Chicago, IL, USA) was employed to analyze data HPLC analyses. Tukey's multiple range 45 test, at a significance level of p < 0.05, was used for the comparisons of the mean values. 46

39

36

41

17

26

	Inde	pendent	variable	ariables Dependent variables								
	V/4		N2	V/A	Y1 -	TPC	Y2 -	FRAP	Y3 -	ABTS	Y4	- TC
Run	X1 EtOH	X2 Time	ХЗ Т	X4 Power	(mg GA	E/100 g	(mg AA	E/100 g	(mg AA	E/100 g	(mg/	100 g
Kull	(%)	(min)	(°C)	(%)	dw)		dw)		dw)		dw)	
					Exp.ª	Pred. <sup>b</sup>	Exp.ª	Pred. <sup>b</sup>	Exp.ª	Pred. <sup>b</sup>	Exp.ª	Pred. <sup>b</sup>
1	25	30	55	60	275.85	305.02	87.39	77.76	289.25	293.69	4.89	5.06
2	50	55	45	70	237.87	250.78	49.77	54.03	244.73	290.57	16.99	16.50
3	50	55	45	90	386.60	357.69	81.65	74.69	364.20	328.99	10.47	10.03
4	50	55	45	70	237.87	250.78	49.77	54.03	244.73	290.57	16.99	16.50
5	50	55	45	70	276.06	282.58	62.21	54.03	293.53	290.57	15.43	16.50
6	25	30	35	60	302.22	257.17	80.36	76.05	341.74	302.18	5.44	6.87
7	75	80	35	80	255.19	257.17	52.13	65.05	214.57	257.00	16.65	17.87
8	100	55	45	70	69.29	86.64	10.06	8.18	129.39	87.17	17.94	17.12
9	75	80	55	80	229.89	259.18	53.79	59.21	225.38	260.98	16.93	16.23
10	75	30	55	60	237.50	239.11	55.70	58.36	264.96	254.38	4.44	4.97
11	50	55	45	70	215.49	250.78	41.19	54.03	274.40	290.57	16.20	16.50
12	25	80	35	80	321.33	329.36	76.42	74.87	325.51	332.13	4.72	4.92
13	50	55	25	70	411.13	361.82	89.90	85.69	356.41	344.36	15.35	12.78
14	75	80	35	60	283.85	277.67	71.60	61.72	252.84	239.01	14.91	16.44
15	50	105	45	70	332.41	293.79	72.33	63.72	327.35	271.57	15.10	13.86
16	75	30	35	60	225.49	228.69	34.32	38.84	186.26	217.80	15.47	15.90
17	50	55	45	70	268.68	250.78	60.62	54.03	343.51	290.57	15.69	16.50
18	75	30	55	80	384.89	331.94	76.04	68.02	234.64	239.90	5.53	6.05
19	25	30	35	80	261.55	296.52	65.01	66.29	310.33	314.40	7.13	6.52
20	25	30	55	89	390.92	406.74	69.02	80.01	299.39	309.27	8.28	7.49
21	75	80	55	60	195.71	191.89	41.84	43.86	205.84	239.63	10.01	12.02
22	25	80	55	60	290.80	267.21	40.70	43.28	237.74	239.63	8.34	6.86
23	25	80	35	60	256.88	340.98	67.62	78.95	251.48	284.09	1.27	2.15
24	50	5	45	70	310.33	308.16	65.42	69.63	246.73	268.59	9.29	8.40
25	50	55	65	70	377.75	386.27	81.75	81.56	361.73	339.86	8.89	9.33
26	75	30	35	80	200.50	233.73	37.96	36.48	205.84	199.97	11.99	14.21
27	25	80	55	80	315.16	343.39	52.44	51.22	284.71	291.05	11.44	12.41
28	0	55	45	70	282.90	224.75	39.93	37.41	193.31	201.62	1.58	2.27
29	50	55	45	70	268.68	250.78	60.62	54.03	343.51	290.57	16.69	16.50
30	50	55	45	50	288.36	276.47	66.55	69.11	294.12	295.42	7.86	6.17

**Table 1.** Experimental conditions and results of total phenolics content (Y1), antioxidant activity (Y2, Y3) and total carotenoids (Y4)1obtained by UAE of chayote peels.2

<sup>a</sup> Experimented values are expressed as average of triplicate determinations from different experiments. <sup>b</sup> Predicted valued based on BBD evaluation. 4

# 3. Results and Discussion

# 3.1. Analysis of BBD

7 Table 1 shows the experimental extraction conditions and the experimental and predicted values of TPC, FRAP, ABTS and TC of chayote peel extracts. For all the responses, 8 there was a close agreement between the experimental values and the theoretical values 9 predicted by BBD. TPC varied between 69.29 and 411.13 mg GAE/ 100 g dw, FRAP from 10 10.06 to 89.90 mg AAE/100 g dw, ABTS from 129.39 to 364.20 mg AAE/ 100 g dw, and TC 11 from 1.27 to 17.94 mg/ 100 g dw. The lowest values of Y1-Y3 responses were recorded at 12 45°C, 100% ethanol, 55 min and 70% (200 W) of ultrasound power (run 8), while the lowest 13 value of TC (Y4 response) was achieved at 80°C, 25% ethanol, 35 min and 60% (170 W) of 14 ultrasound power (run 23). 15

4 5

## 3.2. Validation of the BBD model

The optimal UAE conditions to maximize the phenolic, antioxidant and carotenoid, 2 composition of chayote peel were predicted by BBD. For this purpose, individual desira-3 bility's of the three responses were combined into a single number and then searched the 4 greatest overall desirability. With a desirability of 89.9% (Figure 1a), the optimum condi-5 tions predicted by the BBD model to maximize the combined Y1, Y2 and Y3 responses 6 were 37% ethanol, 55 °C and 224 W, for 30 min. The experimental values agreed within a 7 95% confidence interval with the predicted values, p = 0.104. Regarding Y4 response, the 8 carotenoids extraction from chayote peel was maximized using 75% ethanol, 30 °C and 9 200 W, for 61 min (desirability of 100%, Figure 1b). The experimental value agreed within 10 a 95 % confidence interval with the predicted value, p = 0.203. Thereby, the adequacy of 11 the models in predicting the optimum UAE conditions of phenolics, carotenoids and an-12 tioxidant compounds from chayote peels was confirmed. 13



**Figure 1.** Desirability index for combined responses Y1 (TPC), Y2 (FRAP) and Y3 (ABTS) (a) and for 15 Y4 (TC) response (b).

The TPC values obtained in this work (406 mg GAE/ 100 g dw) are in the same range 17 than those reported by [2], ~ 500 mg GAE/ 100 g dw, when a UAE probe device (20 kHz, 18 375 W) was used and extractions were performed with 25% ethanol, 140 rpm, 30 min, at 19 25 °C and a sample/solvent ratio of 100 mg/mL. Nevertheless, these authors showed that 20 extraction with 100% water stood out as the most potent in obtaining phenolic compounds 21  $(746.46 \pm 58.73 \text{ mg GAE}/100 \text{ g dw})$ . The TC values obtained in this work are significantly 22 higher than values reported by [3], 1.7 mg/ 100 g dw, when powdered samples were ex-23 tracted in acetone and transferred to petroleum ether phase for carotenoids estimation. 24

## 3.3. Phenolic Composition Profile of Optimal Extract

HPLC-DAD was employed to evaluate the phenolic composition profile of chayote 26 peel extracts; Table 2 summarizes the identified phenolic compounds by the chromato-27 graphic analysis, which could contribute to the antioxidant activity observed in the opti-28 mal extract. The phenolic composition determined by HPLC-DAD revealed the presence 29 of compounds belonging to different families, with 4-hydroxyphenilacetic acid (33.32  $\pm$ 30 1.67 mg/100 g dw), gallic acid ( $15.09 \pm 0.75 \text{ mg}/100 \text{ g dw}$ ), protocatechuic acid ( $14.99 \pm$ 31 0.75 mg/100 g dw), ferulic acid (14.9  $\pm$  0.75 mg/100 g dw) and p-coumaric acid (11.2  $\pm$ 32 0.56 mg/100 g dw) being the major contributors to the demonstrated antioxidant proper-33 ties of the produced UAE-chayote peel extract. These phenolic compounds have been pre-34 viously identified in chayote peel extracts [2]; however, different amounts have been 35 quantified depending on the variety, as well as from the extraction conditions employed. 36 Similar as reported by [2], the phenolic acid caffeic acid and the flavonone naringin were 37 also quantified in the UAE- chayote peel extract. The phenolic acids accounted for 71% of 38 the total compounds quantified, followed by 21% of flavonols (mostly represented by my-39 ricetin). 40

1

14 15

Compounds	UAE (mg/ 100 g dw)		
Gallic acid	15.09 ± 0.75		
Protocatechuic acid	14.99 ± 0.75		
4-hydroxyphenilacetic acid	33.32 ± 1.67		
4-hydroxybenzoic acid	ND		
4-hydroxybenzaldehyde	1.03 ± 0.05		
Chlorogenic acid	4.91 ± 0.25		
Vanillic acid	4.28 ± 0.24		
Caffeic acid	0.91 ± 0.05		
Syringic acid	0.35 ± 0.18		
<i>p</i> -coumaric acid	11.2 ± 0.56		
, Ferulic acid	14.9 ± 0.75		
Sinapic acid	2.21± 0.11		
Cinnamic acid	6.47 ± 0.32		
$\Sigma$ Phenolic acids	118.93 ± 5.95		
(+)-Catechin	1.80 ± 0.09		
(-)Epicatechin	4.50 ± 0.23		
$\Sigma$ Flavanols	6.30 ± 0.31		
Naringin	5.98 ± 0.30		
Naringenin	1.06 ± 0.05		
Pinocenbrin	2.13 ± 0.11		
$\Sigma$ Flavanones	9.16 ± 0.46		
Rutin	1.03 ± 0.05		
Quercetin-3-O-glucopyranoside	2.05 ± 0.10		
Quercetin-3-O-galactoside	ND		
Myricetin	20.96 ± 1.05		
Kaempferol-3- <i>O</i> -glucoside	4.28 ± 0.21		
Kaempferol-3-O-rutinoside	1.34 ± 0.06		
Quercetin	2.13 ± 0.11		
Tiliroside	0.91 ± 0.04		
Kaempferol	2.29 ± 0.11		
$\sum$ Flavonols	34.92 ± 1.75		
$\Sigma$ Stilbenes (Resveratrol)	1.56 ± 0.08		
Phloridzin	0.56 ± 0.02		
Phloretin	0.34 ± 0.02		
$\sum$ Others	0.90 ± 0.05		
$\sum$ All phenolic compounds	167.03 ± 8.35		

**Table 2.** Content (mg/100 g dw) of the identified phenolic compounds in the optimal UAE-chayote1peel extract. Results were expressed as mean  $\pm$  standard deviation (n = 3).2

3.4. Vitamin A, Vitamin E, Carotenoid, and Chlorophyll Composition Profile of Optimal Extract 3

The HPLC analysis of the UAE-chayote peel extract showed that tocopherol esters 4 were the main class of carotenoids ( $1047.20 \pm 121.41 \mu g/g$  dw extract), followed by retinol 5 esters ( $245.89 \pm 34.85 \mu g/g$  dw extract) and  $\alpha$ -tocopherol ( $219.13 \pm 12.80 \mu g/g$  dw extract). 6 The content of  $\alpha$ -tocopherol was higher than that reported for spinach, 75-88  $\mu g/g$  DW, 7 suggesting that chayote peel might have the potential to supply nutritionally relevant vitamin E in the diet. 9

4. Conclusions

This study successfully applied a BBD as a practical approach to optimize the UAE 1 conditions of phenolics and carotenoids from chayote peels, which can be further safely 2 applied to food or cosmetic industries creating an added value to this residue. 3

Author Contributions: E.F.V.: Conceptualization, Supervision, Investigation, Methodology, Writ-<br/>ing—original draft, review & editing. M.M.M.: Formal analysis, Data curation, Writing—review &<br/>editing. A.B.d.S.: Formal analy- sis, Data curation, Writing—review & editing. R.C.: Formal analysis,<br/>Data curation, Writing—review & editing. S.C.: Supervision, Formal analysis, Data curation, Writ-<br/>review and editing. C.D.-M.: Supervision, Resources, Writing—review and editing. All au-<br/>thors have read and agreed to the published version of the manuscript.4

Funding: This work was supported through the projects UIDB/50006/2020 and UIDP/50006/2020,10funded by FCT/MCTES through national funds. Elsa F. Vieira and Manuela Moreira thank FCT11(Fundação para a Ciência e Tecnologia) for funding through the Scientific Employment Stimulus-12Individual Call (Ref. CEECIND/03988/2018 and CEEC- IND/02702/2017, respectively).13

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors also thank the project SYSTEMIC "an integrated approach to the<br/>challenge of sustainable food systems: adaptive and mitigatory strategies to address climate change<br/>and malnutrition". The Knowledge hub on Nutrition and Food Security has received funding from<br/>national research funding parties in Belgium (FWO), France (INRA), Germany (BLE), Italy<br/>(MIPAAF), Latvia (IZM), Norway (RCN), Portugal (FCT), and Spain (AEI) in a joint action of JPI<br/>HDHL, JPI-OCEANS and FACCE-JPI, launched in 2019, under the ERA-NET ERA-HDHL (no<br/>22<br/>2323

**Conflicts of Interest:** The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

# References

- Vieira, E.F.; Pinho, O.; Ferreira, I.M.; Delerue-Matos, C. Chayote (*Sechium edule*): A review of nutritional composition, bioactivities, and potential applications. *Food Chem* 2019, 275, 557–568. https://doi.org/10.1016/j.foodchem.2018.09.146
- de Souza Medina, T.; D'Almeida, C.T.d.S.; Nascimento, T.P.d.; de Abreu, J.P.; de Souza, V.R.; Kalili, D.C.; Teodoro, A.J.; Cameron, L.C.; Koblitz, M.G.; Ferreira, M.S.L. Food Service Kitchen Scraps as a Source of Bioactive Phytochemicals: Disposal Survey, 30 Optimized Extraction, Metabolomic Screening and Chemometric Evaluation. *Metabolites* 2023, 13, 386. 31 https://doi.org/10.3390/metabol3030386
- 3. Bellur Nagarajaiah, S.; Prakash, J. Chemical composition and bioactive potential of dehydrated peels of *Benincasa hispida, Luffa acutangula*, and *Sechium edule*. *J Herbs Spices Med Plants* **2015**, 21, 193–202. https://doi.org/10.1080/10496475.2014.940437
- Vieira, E.F.; Souza, S.; Moreira, M.M.; Cruz, R.; Silva, A.B.d.; Casal, S.; Delerue-Matos, C. Valorization of Phenolic and Carotenoid Compounds of *Sechium edule* (Jacq. Swartz) Leaves: Comparison between Conventional, Ultrasound- and Microwave Assisted Extraction Approaches. *Molecules* 2022, 27, 7193. https://doi.org/10.3390/molecules27217193.

38

33

34

14

15

16

24

25