

Proceeding



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Microwave-assisted solid-liquid extraction using propylene glycol as solvent in the recovery of bioactive compounds from umbu seeds

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Abstract: Umbu (Spondias tuberosa) is a Brazilian Caatinga native fruit that has a socioeconomic 12 importance. Pulp is the main product of its agro-industrialization. However, depulping gives rise 13 to about 25% of residue. It is known that fruit peels and seeds are composed of macro and micro-14nutrients as well as bioactive compounds, becoming the use of umbu residue an opportunity to add 15 value to the fruit agro-chain. As the recovery of bioactive compounds from umbu seeds has not yet 16 been optimized, this work aimed to evaluate their recovery through a more sustainable approach, 17 using propylene glycol as solvent in microwave-assisted solid-liquid extraction. For that, an ex-18 perimental design varying the propylene glycol percentage (15-85%) and temperature (59-201 °C) 19 was adopted. The total phenolic compounds (TPC) and antioxidant capacity by DPPH• were 20 evaluated as responses. The highest values were found at 201 °C and 50% propylene glycol (3242 21 mg GAE/100 g and 162 µmol Trolox/g for TPC and DPPH+, respectively). The values observed 22 were, at least, 8 times higher than those obtained in the worst experimental condition (25% pro-23 pylene glycol and 80 °C), showing that the independent variables had effect on the evaluated re-24 sponses (p<0.05). All models were significant, displaying p value <0.05 and no lack of fit was ob-25 served. High adjusted R² values (>0.95) confirm a good fit of the data to the models. Thus, micro-26 wave-assisted extraction employing propylene glycol as solvent can be a more sustainable way for 27 the use of umbu seeds. 28

Keywords: Spondias tuberosa; antioxidant capacity; Brazilian Caatinga

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1. Introduction

The umbu tree (Spondias tuberosa) is a fruit tree native to the Brazilian Caatinga and 32 belongs to the Anacardiaceae family, also known as the "sacred tree of the Sertão" named 33 by Euclides da Cunha. Its name in Tupi-Guarani is "ymbu" which means "drinking tree" 34 due to its ability to store water in its roots [1]. Relevant bioactive compounds, such as 35 quercetin, rutin and vitamin C, are part of its phytochemical composition, as already 36 reported by Ribeiro et al. [2]. Therefore, it has interesting biological properties for dif-37 ferent areas of the industry. In addition, the fruit presents a relevant socioeconomic im-38 portance since it is present in the diet of the population and its cultivation can increase 39 the income of families from semiarid region. The main product of the 40 agro-industrialization of this fruit is the frozen pulp. However, the depulping process 41 can generate a significant amount of residue. A total of 25% in mass can be obtained after 42 processing. This residue is composed of the seeds, peels, and refining cake, which con-43 tains compounds with high added value, making the use of this material very advanta-44

geous [2].

The extraction process of bioactive compounds conventionally uses solvents known 2 for their toxicity and inflammability, which makes the procedure dangerous and requires 3 purification steps before their application. Thus, in order to avoid these problems and make the process more sustainable, safer and more environmentally friendly, green solvents have been evaluated [3]. 6

Propylene glycol is a clear, slightly viscous, water-miscible liquid. It is used as sol-7 vent for aromas, essences and fragrances [4]. These characteristics make it a potential 8 substitute for the most inflammable and toxic conventional solvents. Due to its non-toxic 9 nature, a propylene glycol bioactive extract offers the advantage of being able to be used 10 directly in a formulation [5].

In order to increase extraction efficiency and become more sustainable process, the 12 solid-liquid extraction can be microwave-assisted. When compared with conventional 13 heating methods, it exhibits a shorter processing time as it is possible to reach high tem-14 peratures in a short time as the heat is dissipated evenly in the plant material, reducing 15 the degradation of the target compounds [6]. 16

Therefore, this work aimed at evaluating recovery of bioactive compounds of umbu 17 seeds through a more sustainable approach, using propylene glycol as solvent in mi-18 crowave-assisted solid-liquid extraction. 19

2. Materials and Methods

2.1. Samples

The umbu seeds used in this study were obtained from the depulping process car-22 ried out in the pilot plant of Embrapa Agroindústria de Alimentos (Guaratiba, Rio de 23 Janeiro). The residue was dried at 50 °C in an oven with forced air ventilation and disin-24 tegrated to obtain a pulverized material. By means of the granulometric analysis of the 25 sample, it was possible to identify that approximately 54% of the material is composed of 26 particles with a diameter between 1.00 and 1.68 mm. 27

2.2. Microwave-assisted solid-liquid extraction

In order to optimize the extraction process, an experimental design was used (Table 29 1). Different propylene glycol concentrations (15-85%) and temperatures (59-201 °C) were 30 evaluated as independent variables. The extraction time and solid-liquid ratio were fixed 31 at 10 min and 1:30 (m/v), respectively. The extraction was performed in microwave Ethos 32 1, Milestone. At the end of each extraction, the extracts were vacuum filtered and stored 33 under freezing until analysis for total phenolic compounds (TPC) and antioxidant ca-34 pacity by DPPH assay. Experimental data were analyzed by response surface method-35 ology using a second-order polynomial equation. Analysis of variance (ANOVA), test to 36 determine the lack of fit of the models, and coefficient of determination (R²) were used to 37 verify the significance of the model considering a significance level of 5%. 38

2.3. Methods

2.3.1. DPPH• assay

The DPPH method was performed according to the methodology proposed by Hi-41 dalgo, Sanchez-Moreno and Pascual-Teresa [7]. For the reactions, 100 µL of each extract 42 was added to 2900 µL of DPPH· solution (6×10-5 M in methanol and diluted to obtain an 43 absorbance of 0.700 at 517 nm) and allowed to react for 30 minutes at room temperature. 44 Then, absorbance was read at 517 nm using methanol as a blank. The results were ob-45 tained from the elaboration of a standard curve of Trolox with different concentrations in 46 a range of 80-700 µmol. Results were expressed as µmol Trolox/g. 47

2.3.2. Total phenolic compounds (TPC)

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This analysis was performed with the Folin-Ciocalteu reagent according to the 1 method described by Georgé et al. [8] For that, 250 µL of each extract were mixed with 2 1250 µL of 10% Folin-Ciocalteu (v/v) and 1000 µL of 7.5% Na₂CO₃ (w/v). Subsequently, 3 the mixtures were heated to 50 °C for 15 minutes, being cooled in an ice bath to read the 4 absorbance at 760 nm. The results were obtained with the aid of a calibration curve pre-5 pared from gallic acid solutions with concentrations ranging from 10 to 100 mg/L. The 6 content of TPC in the extracts was expressed as mg gallic acid equivalents per 100 g of 7 sample (mg GAE/100 g). 8

2.4. Statistical analysis of data

The experimental data obtained was evaluated on Statistica software version 13 10 (Dell Inc.), through analysis of variance (ANOVA) and Pareto chart. The test to determine the model's lack of fit and determination of R^2 were employed to verify the model significance, considering a confidence interval of 95%. 13

3. Results and Discussions

By means Table 1 it is possible to verify that both solvent composition and the pro-15 cessing temperature influenced the evaluated responses, since the antioxidant capacity and the content of TPC varied considerably among the obtained extracts. The analysis of 17 the antioxidant capacity of the extracts by the DPPH[•] method shows that this potential 18 ranged from 16 to 162 µmol Trolox/g reached a value ten times higher in the trial 8, 201 °C and 50% propylene glycol. 20

The content TPC in the extracts ranged from 406 to 3242 mg GAE/100 g, resulting in 21 a value eight times higher compared to the worst experimental condition (trial 1: 25% 22 propylene glycol, 80 °C), corroborating the results for antioxidant capacity. It is worth 23 noting that the best response is twice as high as the result reported by Petchsomrit et al. 24 [5] (1503 mg GAE/100 g), which evaluated the extraction of bioactive compounds from 25 Buchanania siamensis, in a microwave-assisted extraction process at a temperature of 60 26 •C, using a solvent of 50% propylene glycol in water for 20 min. 27

Extract obtained in the best condition of the experimental design (trial 8) also 28 showed a higher concentration of TPC (3242 mg GAE/100 g) when compared to extracts 29 obtained using microwaves as in the case of red onion (1239 GAE/100 g) [9], different 30 pepper species (82-2939 mg GAE/100 g) [10], cherry processing residue (1414 GAE/100 g) 31 [11], and umbu seeds by conventional extraction using acetone as solvent (947 g GAE/ 32 100 g) [12]. 33

The Pareto diagram (Figures 1 and 2) presents the effects of the independent varia-34 bles on responses. It is possible to observe that the temperature and propylene glycol had 35 an effect on TPC and DPPH[•]. The linear effect of the temperature showed a positive value 36 (p<0.05), demonstrating that the increase in the temperature favors the recovery of bio-37 active compounds. Furthermore, the quadratic effect was also significant, proving that 38 there is a limit to this behavior. Higher temperatures favor the extraction of biocom-39 pounds, by reducing the solvent's viscosity, making easier the solvent-sample interac-40 tion, increasing the solubility of the target compounds and the diffusion coefficient [13]. 41 However, very high temperatures can result in the degradation of certain substances, 42 reducing the efficiency of the extraction process, which requires studies of process opti-43 mization [14,5]. The positive linear effect of propylene glycol concentration in the solvent 44 was also significant, showing that increase of propylene glycol concentration in the ex-45 tractive solution improves the efficiency of the process. This behavior is explained by the 46 chemical affinity of propylene glycol with the sample's compounds, that is, a solvent 47 with an intermediate polarity interacts well with a wide range of bioactive compounds, 48such as phenolic compounds. Furthermore, the combination of the propylene glycol with 49 water is required in order to reduce the viscosity of the solvent system [15]. Regarding 50 the statistical analysis, all models were significant for predicting the behavior of the re-51

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sponses, since the calculated F values (36 and 295 for TPC and DPPH[•], respectively) were 1 higher than the listed F value ($F_{5,5}$ =5.05) at α =0.05. Adjusted R² values were higher than 2 0.95, which explains at least 95% of the data variability. 3

Thus, these results show the potential of the umbu seeds to obtain an antioxidant 4 extract and that the more sustainable approach by combining microwaves and propylene 5 glycol provided promising results. 6

Table 1. Experimental design for optimization of microwave-assisted extraction of antioxidant7compounds from umbu seeds using propylene glycol as solvent and results for content of total8phenolic compounds (TPC) and antioxidant capacity by DPPH• assay.9

Trial	Propylene glycol (%)	Temperature (°C)	DPPH· (µmol Trolox/g)	TPC (mg GAE/100 g)
1	25	80	16	406
2	25	180	99	2289
3	75	80	21	801
4	75	180	143	3187
5	15	130	22	970
6	85	130	56	1159
7	50	59	20	551
8	50	201	162	3242
9	50	130	71	1603
10	50	130	65	1498
11	50	130	64	1413

GAE: Gallic acid equivalent.



Figure 1. Effect of the independent variables on the antioxidant capacity by DPPH• assay and response surface. 12



Figure 2. Effect of the independent variables on the total phenolic compounds (TPC) and response15surface.16

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4. Conclusion

From the results, it is possible to conclude that using propylene glycol as a green 2 solvent in microwave-assisted solid-liquid extraction favored the recovery of bioactive 3 compounds from the umbu seeds. Also, the condition that produces extract with higher values of antioxidant capacity and content of total phenolic compounds was at 201 °C, 5 using a 50% propylene glycol solution as solvent. Thus, microwave-assisted extraction 6 employing propylene glycol as solvent can be a more sustainable way to add value to the 7 umbu seeds, considered a residue in this agro-chain. 8

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References

- Barreto: L. S.; Castro, M. S. Good management practices for sustainable umbu extraction. Brasilia: Embrapa Genetic Resources 1. and Biotechnology, 2010. 15
- 2. Ribeiro, L. O.; Viana, E.S.; Godoy, R. L. O.; Freitas, S.C.; Freitas, S.P.; Matta, V.M. Nutrients and compounds of pulp, peel and seed from umbu fruit. Cienc. Rural. 2019, v.49.
- Oreopoulo, A.; Tsimogiannis, D.; Oreopolou, V. Extraction of polyphenols from aromatic and medicinal plants: an overview of 3. the methods and the effect extraction parameters. In: Watson, R.R (Ed), Polyphenols in Plants. Academic Press, 2019, pp. 243-259.
- Santos, J. S. B.; Misael, C. G. A.; Fernandes, C. V.; Chaves, F. J. F.; Cavalcante, J. N. A.; Vasconcelos, S. F. Conversion Analysis of 4. the Propylene Glycol Reaction. In: V Regional Chemistry Meeting & IV National Chemistry Meeting. São Paulo, Brasil. 2015.
- 5. Petchsomrit, A.; Chanthathamrongsiri, N.; Manmuan, N.; Leelakanok, N.; Wangpradit, N.; Vongsak, B.; Sirirak, T.Green extraction of Buchanania siamensis and water-based formulations. Sustainable Chemistry and Pharmacy 2022, v.30.
- 6. Tsukui, A.; Rezende, C. M.. Microwave Assisted Extraction and Green Chemistry. Rev. Virtual Quim 2014, v.6.
- 7. Hidalgo, M.; Sánchez-Moreno, C.; de Pascual-Teresa, S.: Flavonoid – Flavonoid interaction and its effect on their antioxidant activity. Food Chem 2010, v.121, 691-696.
- Georgé, S.; Brat, P.; Alter, P.; Amiot, M.J. Rapid Determination of Polyphenols and Vitamin C in Plant-Derived Products. J. 8. Agric. Food Chem 2005, v.53, 1370-1373.
- 9. Dairi, S.; Dahmoune, F.; Belbahi, A.; Remini, H.; Kadri, N.; Aoun, O.; Bouaoudia, N.; Madani, K.: Optimization of microwave extraction method of red onion phenolic compounds using response surface methodology and inhibition of low-density lipoprotein oxidation. Journal of Applied Research on Medicinal and Aromatic Plants 2021, v.2.
- Gallo, M.; Ferracane, R.; Graziani, G.; Ritieni. A.; Fogliano, V.; Microwave Assisted Extraction of Phenolic Compounds from 10. Four Different Spices. *Molecules* **2010**, *v*.15, 6365-6374.
- Simsek, M.; Sumnu, G.; Sahin. S.. Microwave Assisted Extraction of Phenolic Compounds from Sour Cherry Pomace. Separation 11. Science and Technology 2012, v.47, 1248-1254.
- Freitas, B.P.; Oliveira, A.H.; Kunigami, C.N.; Novo, A.A.; Matta, V.M.; Jung, E.P.; Ribeiro, L.O. Effect of the solvent on the 12. recovery of bioactive compounds from Umbu residue. Molecules 2022, v. 27,410.
- Ribeiro, L.O.; Freitas, B.P.; Lorentino, C.M.A.; Frota, H.F.; Santos, A.L.S.; Moreira, D.L.; Amaral, B.S.; Jung, E.P; Kunigami, C.N. 13. Umbu Fruit Peel as Source of Antioxidant, Antimicrobial and α-Amylase Inhibitor Compounds. Molecules 2022, v.27, 410.
- Neves-Brito, B. S.; Láscaris, M. P. S.; Moreira, J. J. S.; Nunes, T. P.; Pagani, A. A. C.; Silva, G. F.. Influence of rota-evaporation 14. temperature and type of residue on the extraction of bioactive compounds from jamelão (Syzygium cumini). Research, Society and Development 2021, v.10, n. 2.
- Nikolic, V.G.; Troter, D.Z.; Savic, I.M.; Gajic, I. M.S.; Zvezdanovic, J.B.; Konstantinovic, I. B.; Konstantinovic, S.S.. Design and 15. 44 optimization of "greener" and sustainable ultrasound-assisted extraction of valuable bioactive compounds from common 45 centaury (Centaurium erythraea Rafn) aerial parts: A comparative study using aqueous propylene glycol and ethanol. Industrial 46 Crops & Products 2023, v.192. 47

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