

Proceedings



Characterization and quantification of phenolic compounds of honeys from Sierra Nevada (Granada)

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Abstract: Antioxidant, anti-inflammatory and antimicrobial effects are some of the effects that have been attributed to honey, especially to its content of bioactive compounds, mainly phenolic compounds (PC), whose content varies greatly depending on the variety, origin, agronomic conditions, harvest season, and climate. The aim of the present study is to characterise 21 honeys from Sierra Nevada (Granada). High performance liquid chromatography coupled to quadrupole-time of flight mass spectrometry (HPLC-ESI-QTOF-MS) was used. The mass accuracy and true isotopic pattern in both MS and MS/MS spectra made possible the tentative identification of 58 PC including flavonoids, phenolic acids and derivatives. The average content of PC was $83.01\pm16.36 \ \mu g/g$, with flavonoids accounting for more than 85%. The most abundant compounds were naringenin ($16.88\pm3.15 \ \mu g/g$), pinocembrin ($12.33\pm2.92 \ \mu g/g$), chrysin ($12.21\pm2.09 \ \mu g/g$), carnosol ($9.52\pm2.90 \ \mu g/g$), galangin ($5.41\pm1.68 \ \mu g/g$) and apigenin ($5.24\pm0.89 \ \mu g/g$). Due to their interesting composition, more studies are necessary to determine if the extreme environmental conditions from Sierra Nevada pose abiotic stress for the plants located there fostering the concentration in PC.

Keywords: honey; phenolic compounds; Sierra Nevada; HPLC-ESI-QTOF-MS

1. Introduction

Honey, a natural product crafted by honey bees (Apis mellifera), is renowned for its nutritive and healthful qualities. Its composition exhibits considerable variability based on factors like botanical and geographical origin [1,2]. Honey primarily consists of sugars (constituting 80-85% of its composition), water (15-17%) and proteins (0.1-0.4%). Additionally, to a lesser extent, it contains enzymes, organic acids, vitamins, minerals and phenolic compounds, all of which significantly contribute to its sensory and functional attributes [3]. Honey has been associated with various beneficial effects, including antioxidant, anti-inflammatory and antimicrobial properties [4]. These health-promoting properties are particularly linked to its content of bioactive compounds, mainly phenolic compounds (PC). The PC content varies substantially depending on factors such as honey variety, origin, agronomic conditions, harvest season, and climate. Plants synthetize PC under both normal and stressful conditions, with functions that encompass attracting pollinating insects and safeguarding against pathogens and ultraviolet (UV) radiation [4]. Recent studies have reported a wide range of total phenolic content (TPC) values in honey, spanning from 6.5 ± 4.2 to $841.7 \pm 304.0 \ \mu g/g$ [5]. While many researchers have analysed the content of bioactive compounds in some Spanish honeys, there are no

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Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/). previous studies on the content of those compounds in honeys from Sierra Nevada (Granada), where the plants are exposed to extreme environmental conditions such as UV radiation, extreme temperatures, and altitude (hypoxia), which could influence the content of PC. Thus, the aim of the present study is to characterise and quantify the PC of 21 honeys from Sierra Nevada, which is a national park with a great variety of vegetation exposed to abiotic stresses that could increase the concentration and/or variety of PC contained in the honeys produced there.

2. Methodology

2.1. Extraction of phenolic compounds

The extraction of PC was performed using a previously described method [6,7] with some modifications. Sixty grams of each of the selected honeys were mixed with 150 ml of acidified water (pH 2.0). The mixture was stirred for 10 minutes at room temperature on a magnetic stirrer and then filtered through cotton wool to remove solid particles. The filtrate was mixed with 40 g Amberlite XAD-2 and stirred for 10 minutes at room temperature on a magnetic stirrer. The Amberlite particles were packed in a 33 cm long, 24 mm inner diameter gravimetric column (Sigma-Aldrich). Then, the column was washed with 100 ml of acidic water (pH 2.0) and then with 300 ml of milliQ water. In this way, PC present in the honey were retained in the column while sugars and other polar compounds were eluted with aqueous solvent. To collect the phenolic fraction, 300 ml of methanol were used. Then the phenolic fraction was dried in a rotary evaporator at a temperature of 30°C. The residue was dissolved in 5 ml of milliQ water and extracted in triplicate with diethyl ether (5 mlx3). The ether extracts were combined and the ether was removed at a rotary evaporator. The residue was dissolved in 1 ml of methanol, filtered through a 45µm membrane filter, transferred to an HPLC vial and frozen at -20°C until analysis.

2.2. Analysis of phenolic compounds

The analytical technique used was High Performance Liquid Chromatography coupled to Mass Spectrometry (HPLC-MS-MS/MS). Specifically, an Acquity HPLC system (Waters Corporation, Milford, MA, USA) coupled to an electrospray ionisation (ESI) source operating in negative mode and a quadrupole time-of-flight (QTOF) mass spectrometer (Waters) was used. To separate the compounds of interest effectively, it was used an ACQUITY UPLC BEH Shield RP18 column (1.7 μ m, 2.1 x 100 mm; Waters Corporation, Milford, MA, USA). Acidified H₂O with 1% acetic acid and methanol as solvents A and B, respectively, were used for the mobile phases. A linear gradient was applied so that at the start 95% was solvent A and 5% solvent B; at minute 30, 23.9% was solvent A and 76.1% solvent B; and at minute 33, 100% was solvent B. The initial conditions were maintained for 6 minutes before each analysis. The column temperature was maintained at 40°C and the injection volume was 0.5ml/min.

2.3. Characterisation of phenolic compounds

MassLynx 4.1 software (Waters Corporation, Milford, MA, USA) was used to process the chromatographic data. The characterisation strategy was based on the exact mass and fragment information provided by the compound-specific MS and MS/MS spectra determined by the QTOF mass analyser. For the acquisition of information on the chemical structure of the compounds, in addition to consulting previously published research, the following databases were used: SciFinder Scholar (http://scifinder.cas.org), FoodDb (https://foodb.ca/), MassBank (http://massbank.jp), Pubchem (https://pubchem.ncbi.nlm.nih.gov), METLIN Metfrag (https://msbi.ipb-halle.de/MetFrag/), (http://metlin.scripps.edu), National Institute of Standards and Technology (https://www.nist.gov/), and the National Institute of Health (NIH) database. Additionally, SIRIUS 4 (https://bio.informatik.uni-jena.de/sirius/) was used to obtain metabolite

structure information. Six analytical standards (catechin, chlorogenic acid, ferulic acid, rutin, phloridizin, quercetin, phloretin and vanillic acid) were employed to estimate the amount of phenolic compounds present in the honeys.

3. Results and discussion

Fifty-eight phenolic compounds were characterized in the 21 honeys, including flavonoids, phenolic acids and derivatives (Table 1).

| [M-H] ⁻ | RT | Molecular formula | Proposed compound | Fragments | References |
|--------------------|--------|----------------------|--|--------------------------------|------------|
| 135.043 3 | 2.189 | C8H8O2 | Phenylacetic acid | 117 | [18] |
| 135.043 8 | 2.01 | C8H8O2 | Vinylcatechol | 134, 133, 105 | [19] |
| 137.022 2 | 4.014 | C7H6O3 | Hydroxybenzoic acid | 93 | [20] |
| 153.020 8 | 4.852 | C7H6O4 | Protocatechuic Acid | 109, 137 | [21] |
| 163.038 6 | 3.225 | C9H8O3 | Cumaric acid (Isomer 1) | 145, 119 | [20] |
| 163.039 6 | 3.341 | C9H8O3 | Cumaric acid (Isomer 2) | 119, 117 | [20] |
| 165.054 7 | 2.458 | C9H10O3 | 4-hydroxicinnamic acid | 161, 133, 132, 122 | [22] |
| 165.055 8 | 3.448 | C9H10O3 | L-(-)-phenylactic acid | 147 | [23] |
| 167.033 7 | 2.065 | C8H8O4 | Homogentisic acid | 134, 137, 131, 117, 108 | [24] |
| 177.018 1 | 1.769 | C9H6O4 | Esculatin | 145, 125, 120, 144 | [25] |
| 193.049 5 | 2.095 | C10H10O4 | Coniferic/ferulic acid | 133 | [26] |
| 195.065 9 | 1.995 | C10H12O4 | 4-methoxyphenylactic acid | 133, 177, 149 | [27] |
| 197.044 2 | 2.16 | C9H10O5 | Siringic acid (Isomer 1) | 106 | [20] |
| 197.045 5 | 2.262 | C9H10O5 | Siringic acid (Isomer 2) | 121, 123 | [20] |
| 211.059 9 | 4.875 | C10H12O5 | Methylsyringate | 181 | [28] |
| 221.080 4 | 3.45 | C12H14O4 | 3-hydroxy-1-(2-methoxyphenyl)penta-1,4-dione (Isomer 1) | 133 | [29] |
| 221.081 1 | 3.605 | C12H14O4 | 3-hydroxy-1-(2-methoxyphenyl)penta-1,4-dione (Isomer 2) | 133 | [29] |
| 223.060 9 | 3.228 | C11H12O5 | Sinapic acid | 144, 116, 142, 160 | [21] |
| 223.096 8 | 6.851 | C12H16O4 | Vanillin 1,2-butylene glycol | 151, 136, 108 | [30] |
| 253.049 7 | 12.928 | C15H10O4 | Chrysin | 209, 143, 145, 119,195 | [31] |
| 255.065 9 | 12.128 | C15H12O4 | Pinocembrin | 171, 133, 213, 134, 169 | [32] |
| 269.043 9 | 10.498 | C15H10O5 | Apigenin | 117,149, 201, 145, 183, 107 | [33] |

Table 1. Phenolic compounds characterised in the 21 honeys.

| 269.044 1 | 9.803 | C15H10O5 | Baicalein | 129, 143, 151 | [25] |
|--------------|--------|----------|---|---------------------------------|------|
| 269.044 5 | 13.681 | C15H10O5 | Galangin | 211, 239, 195,167, 151 | [34] |
| 271.060 0 | 7.979 | C15H12O5 | Pinobanksin | 253, 197,225, 209, 125 | [35] |
| 271.060 3 | 8.406 | C15H12O5 | Naringenin | 253,197, 161,125, 225 | [20] |
| 283.059 7 | 11.218 | C16H12O5 | Prunetin | 211, 238, 167, 165 | [36] |
| 283.060 4 | 13.878 | C16H12O5 | Biochanin A | 268, 211, 239, 269, 195 | [37] |
| 283.060 5 | 13.469 | C16H12O5 | Genkwanin | 134, 175, 168, 148, 159 | [38] |
| 283.096 9 | 13.161 | C17H16O4 | Phenylethyl caffeate | 135, 133, 161, 134 | [34] |
| 285.038 1 | 10.787 | C15H10O6 | Kaempferol (Isomer 1) | 151, 184, 245, 255, 273 | [39] |
| 285.039 2 | 6.831 | C15H10O6 | Luteolin (Isomer 1) | 151, 257 | [40] |
| 285.039 4 | 13.682 | C15H10O6 | Kaempferol (Isomer 2) | 269, 268, 211, 239 | [39] |
| 285.039 6 | 8.691 | C15H10O6 | Luteolin (Isomer 2) | 255, 133, 283, 151 | [40] |
| 285.040 4 | 7.646 | C15H10O6 | Kaempferol (Isomer 3) | 255, 227, 211, 284 | [39] |
| 285.040 8 | 9.135 | C15H10O6 | Luteolin (Isomer 3) | 241, 133 | [40] |
| 285.077 2 | 12.291 | C16H14O5 | 5-O-Methylnaringenin | 188, 191, 255, 243, 158 | [20] |
| 287.055 5 | 5.52 | C15H12O6 | Eriodictyol | 161, 269, 251 | [41] |
| 299.054 5 | 11.479 | C16H1206 | Kaempferide (Isomer 1) | 284, 227, 256, 165, 269 | [42] |
| 299.054 7 | 11.155 | C16H1206 | Kaempferide (Isomer 2) | 284 | [42] |
| 301.032 2 | 4.236 | C15H1007 | Quercetin | 175, 183, 201, 225, 245 | [42] |
| 301.033 9 | 4.077 | C15H1007 | Quercetin | 255, 273, 213, 151 | [42] |
| 301.035 4 | 9.343 | C15H1007 | Morin | 273, 151, 257, 178, 255 | [34] |
| 301.069 6 | 9.9400 | C16H14O6 | Hesperetin (Isomer 1) | 164 | [43] |
| 301.071 3 | 5.3190 | C16H14O6 | Hesperetin (Isomer 2) | 151, 177 | [43] |
| 301.071 7 | 5.4440 | C16H14O6 | Hesperetin (Isomer 3) | 177, 286 | [43] |
| 301.200 8 | 8.5460 | C15H10O7 | Tricetin | 255, 151 | [44] |
| 315.048 7 | 10.875 | C16H12O7 | 3-Methylquercetin/Isorhamnetin (Isomer 1) | 241, 242, 270, 313, 300 | [39] |
| 315.050 0 | 10.157 | C16H12O7 | 3-Methylquercetin/Isorhamnetin (Isomer 2) | 300 | [39] |
| 315.050 6 | 9.136 | C16H12O7 | 3-Methylquercetin/Isorhamnetin (Isomer 3) | 241, 242, 270, 271, 300, 313 | [39] |
| - | | | | | |

| 329.065 0 | 9.824 | C17H14O7 | Quercetin dimethyl ether (Isomer 1) | 314 | [45] | |
|--------------|--------|---------------|--|-------------------------|------|--|
| 329.066 4 | 11.956 | C17H14O7 | Quercetin dimethyl ether (Isomer 2) | 314 | [45] | |
| 329.174 4 | 15.415 | C20H26O4 | Carnosol (Isomer 1) | 285 | [35] | |
| 329.174 5 | 15.068 | C20H26O4 | Carnosol (Isomer 2) | 285 | [35] | |
| 329.175 3 | 15.708 | C20H26O4 | Carnosol (Isomer 3) | 285 | [35] | |
| 329.175 8 | 14.856 | C20H26O4 | Carnosol (Isomer 4) | 285 | [35] | |
| 431.097 4 | 7.651 | C21H20O1 0 | Kaempferol-rharmnoside | 285, 255, 227 | [46] | |
| 461.106 5 | 9.049 | C22H22O1 1 | 8-Methoxykaempferol 7-rhamnopyranoside | 287, 299, 315, 259, 139 | [46] | |

RT: retention time.

The average TPC from the Sierra Nevada honeys analysed was $83.01\pm16.36 \ \mu g/g$, being above the average of other Spanish honeys such as Galician honeys, which contain an average of $38 \mu g/g$ [8]. Flavonoids accounted for more than 85% of the TPC. Figure 1 shows the content of phenolic acids, flavonoids, other phenolic compounds and total phenolic compounds of each honey. The most abundant compounds were naringenin (16.88±3.15 μg/g), pinocembrin (12.33±2.92 μg/g), chrysin (12.21±2.09 μg/g), carnosol $(9.52\pm2.90 \text{ }\mu\text{g/g})$, galangin $(5.41\pm1.68 \text{ }\mu\text{g/g})$ and apigenin $(5.24\pm0.89 \text{ }\mu\text{g/g})$. Naringenin is noted for its positive effects on the cardiovascular system through antioxidant, anti-inflammatory, anti-atherogenic and anti-apoptotic actions [9]. Pinocembrin is a flavanone with antioxidant, antimicrobial and anti-inflammatory properties, and has recently been studied for its potential to inhibit histidine decarboxylase as a new natural anti-allergic drug candidate [10]. Chrysin has shown significantly greater antiproliferative activity on cancer cell growth than other compounds [11]. It also has antioxidant, anti-obesity, antiinflammatory, anti-diabetic and neuroprotective activity [12]. Carnosol is a phenolic diterpene with demonstrated antioxidant, anti-inflammatory and anticancer activity [13,14]. Positive effects of carnosol have also been reported in ischaemic stroke, by inhibiting apoptosis and attenuating oxidative damage and cellular inflammation [15]. The main positive effects of galangin are attributed to its anti-inflammatory, antioxidant, anticancer and antineoplastic properties [16]. Apigenin has shown therapeutic functions through cell cycle arrest, apoptosis, anti-inflammatory. In addition, apigenin contributes to counteracting oxidative stress by enhancing the expression of anti-oxidant enzymes such as glutathione synthase, catalase and superoxide dismutase. After its absorption into the digestive tract, apigenin is able to reach the brain and could have antidepressant and antianxiety effects [17].



Figure 1. Content of phenolic compounds of Sierra Nevada honeys.

4. Conclusion

Due to their interesting composition, more studies are necessary to determine if the peculiar environmental conditions from Sierra Nevada, such as UV radiation, extreme temperature, or altitude (hypoxia) pose abiotic stress for the plants located there fostering the concentration in PC and thus increasing the antioxidant, antimicrobial, anti-inflammatory and anticarcinogenic activity of these honeys.

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