

# Potential Probiotic *Bacillus* Strains Isolated from Contaminated Soil in North Macedonia: *Salmonella* Growth Inhibition <sup>†</sup>

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**Abstract:** Salmonellosis, which occurs most frequently (85%) as a result of consuming contaminated food, is brought on by salmonellae, which are bacteria that can infect both humans and animals. The aim of this study was to investigate the probiotic properties of three *Bacillus* spp. strains isolated from contaminated soil in North Macedonia and their antimicrobial activity. Additionally, their ability to survive in the presence of bile salts and at low pH, and their susceptibility to antibiotics were examined. This research indicates that new *Bacillus* strains' probiotic qualities are also promising and exhibit strong inhibition activity against *Salmonella enterica* ATCC 10708.

**Keywords:** soil microorganisms; antimicrobial activity; probiotics

## 1. Introduction

Probiotics can be described as live microbial or cultured product feed supplements that have beneficial effects on the host through their production of inhibitory agents, vying for chemicals and adhesion sites, enhancing the microbial stability, and altering and boosting the immune system [1]. Probiotic supplements made from bacterial spore formers are utilized in prescription drugs, dietary supplements for mankind, and animal feeds. They are interesting as food additives because of their heat-resistant properties and capacity to cross the gastrointestinal barrier, and this role has now become advanced. Although sometimes thought of as soil organisms, Bacilli should actually be viewed as commensals in the gut [2]. A viable substitute for conventional antibiotic therapies is the use of probiotics to control illnesses and prevent the increase of antibiotic resistance. Additionally, by regulating the metabolic pathways that lead to the manufacture of hazardous compounds, probiotics can increase the immune system function, activate endogenous enzymes, create antimicrobial substances, and enhance the production of endogenous enzymes. Probiotics can create antimicrobial compounds that limit the growth of infections and the development of toxins [3]. Probiotics that produce spores have grown in popularity recently. Some Bacilli are utilized as probiotics in poultry and are Gram-positive spore-forming bacteria. Studies have demonstrated that *Bacillus* spores can germinate in the digestive system of chickens [4].

Salmonellosis, which occurs most frequently (85%) as a result of consuming contaminated food, is brought on by salmonellae, which are bacteria that can infect both humans and animals [5]. By assisting with the absorption of some critical nutrients, probiotics have the potential to exert growth-promoting effects by competitively excluding pathogens and boosting feed conversion rates. *Salmonella* remains active and can spread on meat and other livestock products that were not properly cooked or kept, making contaminated foods the primary mechanism of transmission. *Salmonella* can survive for years without

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showing any overt clinical symptoms in both humans and animals, and in certain situations, these individuals and animals can develop a chronic infection. Furthermore, an unprecedented increase in *Salmonella* that is resistant to antibacterial medications has been caused by overusing medicines in humans as well as animals [6]. As a result, researchers are currently considering alternative methods of preventing and treating *Salmonella* infections. The aim of this study was to investigate the probiotic properties of three *Bacillus* spp. strains isolated from contaminated soil in North Macedonia and their antimicrobial activity against *Salmonella enterica* ATCC 10708 using the agar-well diffusion method. For identification, isolates were characterized morphologically and physiologically. Additionally, their ability to survive in the presence of bile salts and at low pH, high osmotic concentrations of NaCl and their susceptibility to antibiotics were examined. Each tested strain demonstrated antagonistic activity against *Salmonella enterica* ATCC 10708. The three different *Bacillus* strains were all resilient to an acidic environment (pH 3.0) and a high osmotic pressure (NaCl at 6.5%). This research indicates that new *Bacillus* strains' probiotic qualities are also promising and exhibit strong inhibition activity against *Salmonella enterica* ATCC 10708.

## 2. Materials and Methods

### 2.1. Antimicrobial Activity

The determination of the antimicrobial activity of the isolates against the test microorganisms was conducted using the agar well diffusion method. The test bacteria *Salmonella enterica* ATCC 10708 used in this analysis is included in the microorganism collection of the Microbiology Laboratory at the Faculty of Natural Sciences and Mathematics, Skopje, North Macedonia. The test bacteria was inoculated in nutrient broth (NB) and incubated at 37 °C for 24 h. The test microorganism was inoculated onto each sterile nutrient agar (NA) Petri dish using a sterile swab, and then wells with a diameter of 6 mm were placed on the nutrient medium. Into these wells, cultures of *Bacillus* spp. were added which were allowed to pre-incubate at 37 °C for 24 h. The plates thus inoculated were incubated at 37 °C for 24 h, and then the diameter of the zone of inhibition was measured for each isolate.

### 2.2. Bile Salt Tolerance

To determine bile salt tolerance, Gilliland et al.'s approach [7] was modified in certain ways. Nutrient Broth (NB) containing 0%, 0.3%, 0.5%, 1%, 1.5% and 2% of bile salts was inoculated with each probiotic strain and incubated for 24 and 48 h at 37 °C. Growth in control (no bile salts) and test cultures was evaluated at 24 and 48 h by measuring the turbidity (FAU).

### 2.3. Antibiotic Resistance

By inoculating the isolates of choice on nutrient agar and then applying antibiotic discs with sterile tweezers, it was possible to test the isolates' resistance to various antibiotics, for antibiotic sensitivity using the Kirby-Bauer methodology. A zone of inhibition was observed after incubation.

### 2.4. Evaluation of Resistance in Intestinal Tract Conditions: Low pH and Sodium Chloride

The Nguyen et al. [8] approach was used to evaluate the tested isolates' resistance to acidic pH values. Testing was done on isolates to see how sensitive they are to a nutrient broth containing 6.5% NaCl. The tubes were incubated at 37 °C for the tests on pH and concentrations of NaCl. At the time points evaluated, each sample was streaked on NA for presence or absence of growth, to confirm livability of the strains and the turbidity of the tubes was assessed.

## 3. Results

### 3.1. Antimicrobial Activity

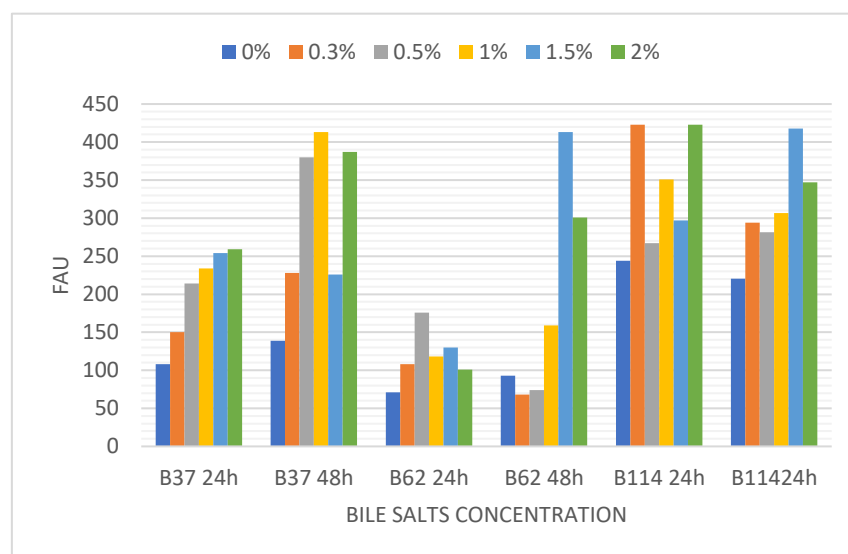
All isolates were found to have inhibitory activity against *Salmonella enterica* ATCC 10708. The isolate B114 showed the greatest inhibitory effect on the test bacteria, with a zone of inhibition of 15.18 mm, while the isolates B37 and B62 showed zones of inhibition of 14.22 mm and 14.83 mm (Figure 1).



**Figure 1.** Antibacterial activity (zones of inhibition) of *Bacillus* spp. from soil against *Salmonella enterica* ATCC 10708.

### 3.2. Bile Salt Tolerance

Evaluating bile salts tolerance of the vegetative cells of our selected strains, we found that all strains were able to grow when cultured at 0%, 0.3%, 0.5%, 1%, 1.5% and 2% of bile salts at 24 and 48 h of incubation (Figure 2).



**Figure 2.** Growth of the isolates B37, B62 and B114 at different % of bile salts at 24 and 48 h of incubation.

### 3.3. Antibiotic Resistance

The antibiotic resistance and susceptibility of the three *Bacillus* isolates to twenty four antibiotics were analyzed. The isolate B37 was sensitive to all test antibiotics, while the isolate B62 was resistant to cefotaxime (Figure 3). The isolate B114 was resistant to ampicillin, cefotaxime, penicillin G and cephalosporins. The diameter of the zones of inhibition is presented in Table 1.

**Table 1.** Antibiotic sensitivity test for *Bacillus* spp. isolates.

Antibiotic	B37	B62	B114
Rifampicin	21.21	19.28	17.12
Ampicillin	22.19	21.74	/
Gentamicin	18.63	18.59	15.07
Doxycycline	25.76	27.99	22.38
Chloramphenicol	26.43	30.88	24.09
Norfloxacin	28.29	30.09	22.60
Amoxicillin/Clavulanic acid	27.46	27.65	9.60
Ceftriaxone	27.76	26.72	9.89
Ticarcillin	26.10	30.31	8.87
Pipemidic acid	26.09	25.80	17.83
Nitrofurantoin	25.45	24.57	19.01
Cefotaxime	33.07	/	/
Ampicillin/Sulbactam	33.07	32.19	9.20
Cefalexin	32.64	33.19	/
Cefotaxime 5	21.47	/	/
Erythromycine	28.98	25.66	9.46
Penicillin G	24.46	19.13	/
Tetracycline	19.48	23.16	9.65
Clarithromycin	33.17	35.67	28.53
Streptomycine	30.23	31.94	17.12
Clindamycine	28.22	24.85	24.09
Mupirocin	28.72	29.87	24.13
Bacitracine	14.19	17.21	/
Penicillin	7.63	16.83	18.50

### 3.4. Evaluation of Resistance in Intestinal Tract Conditions: Low pH and Sodium Chloride

Growth was seen on the control plates inoculated with the isolates as well as on the plates with medium with a pH value of 3, indicating that the isolates have the capacity to grow at pH 3 i.e., tolerance to low pH values after the incubation period of 24 h. This quality is crucial in potential probiotics since the bacterium needs to be tolerant to acidic pH in order to thrive both in the stomach and on food. All strains were also able to tolerate high osmotic concentrations of NaCl.

## 4. Discussion

Before entering the intestinal tract, probiotic bacteria must make it through the stomach, where the pH can drop as low as 1.5 to 2.0 [9], and remain alive for at least 4 h [10]. Probiotic microorganisms that exhibit resistance to a particular antibiotic may be administered along with antibiotic therapy [11]. Since conjugative plasmids frequently carry antibiotic resistance genes, they can spread to other bacteria [12] and potentially give rise to enteropathogenic bacteria that are resistant to antibiotics. Determining whether antibiotic-resistant genes are located on chromosomes or plasmids is therefore crucial [11]. In general, it has been thought that tolerance to bile salts is a need for bacterial colonization and metabolic activity in the host's intestine. Depending on the person and the kind and quantity of food consumed, the small intestine's bile salt content ranges from 0.2% to 0.3% on average and may reach 2% (*w/v*) [11,12]. However, bile levels in the colon fluctuate and are often low until a fatty meal is consumed. Bile secretion's primary goal is to emulsify and dissolve ingested lipids. Bile salts can rupture the lipid barrier, enter the bacterial cell, denature proteins, chelate ions, and harm DNA, but they also have bactericidal effects [13].

## 5. Conclusions

Current probiotic products, which have been demonstrated to prevent gastrointestinal disorders in both humans and animals, contain bacterial spore formers, particularly those of the genus *Bacillus*. Numerous uses for these probiotic-based spores have been demonstrated, including the treatment of immunosuppressive disorders and diarrhea brought on by antibiotics. The findings of this investigation demonstrated the probiotic *Bacillus* spp. strains' endurance of various physiological circumstances as well as *Salmonella enterica* suppression. Additionally, the techniques employed to screen isolates could be crucial in determining if *Bacillus* spp. are suitable for use as probiotics in both people and animals.

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