



2nd International Electronic Conference on Medicinal Chemistry

1-30 November 2016

chaired by Dr. Jean Jacques Vanden Eynde

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Chiral derivatives of xanthenes: investigation of enantioselectivity as inhibitors of cyclooxygenases (COX-1 and COX-2) and binding interaction with human serum albumin

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Chiral derivatives of xanthenes: investigation of enantioselectivity as inhibitors of cyclooxygenases (COX-1 and COX-2) and binding interaction with human serum albumin

Graphical Abstract



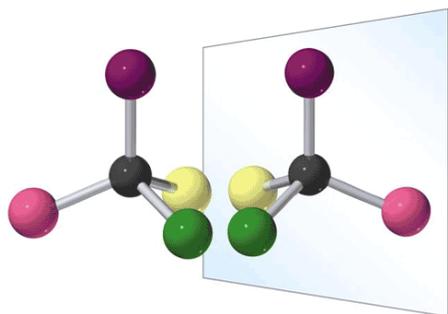
Abstract:

Searching for new enantiomerically pure chiral derivatives of xanthenes (CDXs) with potential pharmacological properties has remained an area of interest of our group, namely those with anti-inflammatory activity. Herein, we describe *in silico* studies and *in vitro* inhibitory assays of different enantiomeric pairs of CDXs. The evaluation of the inhibition of cyclooxygenases (COX-1 and COX-2) activities was performed by using the COX Inhibitor Screening Assay Kit. Docking simulations between the small molecules (CDXs, known ligands and decoys) and the enzyme targets were undertaken with AutoDock Vina embedded in PyRx – Virtual Screening Tool software. All the CDXs evaluated exhibited COX-1 and COX-2 inhibition potential as predicted.

Considering that the (*S*)-(-)-enantiomer of the nonsteroidal anti-inflammatory drug Ketoprofen preferentially binds to albumin, resulting in lower free plasma concentration than (*R*)-(+)-enantiomer, protein binding affinity for CDXs was also evaluated by spectrofluorimetry. For some CDXs enantioselectivity was observed.

Keywords: chiral derivatives of xanthenes, cyclooxygenase, albumin, enantioselectivity





Chirality plays a key role in biochemical events

Chiral molecular recognition strongly influences drug action and efficacy

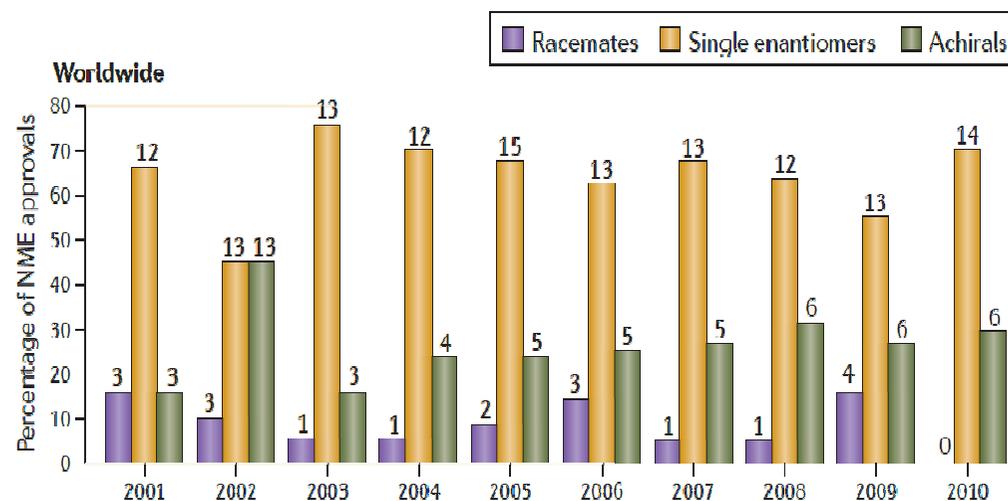
Frequently only one of the two enantiomers exerts the desired effect
Eutomer/Distomer



Major importance in drug discovery and design

De novo design

Chiral switches



Agranat, S. R. Wainschein, and E. Z. Zusman, *Nat. Rev. Drug Discov.*, 2012, 11, 972–973.



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on Medicinal Chemistry
1-30 November 2016

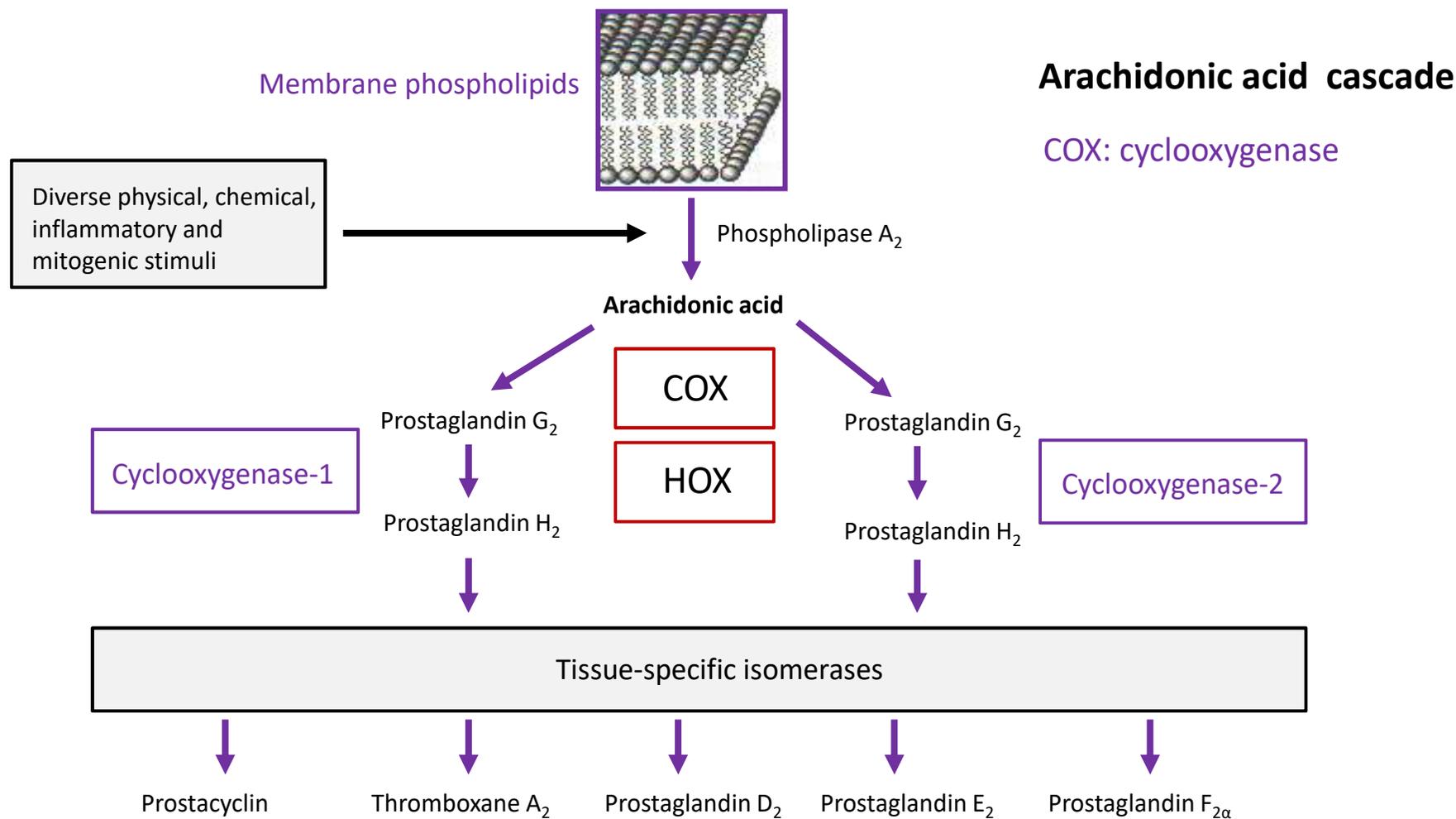
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pharmaceuticals

Introduction

INFLAMMATORY PATHWAY AND ITS MEDIATORS



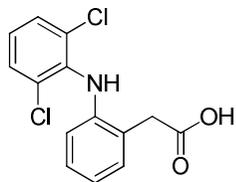
FitzGerald, G. A., *Nat Rev Drug Discov*, 2003, 2(11), 879-890.



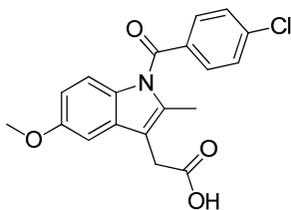
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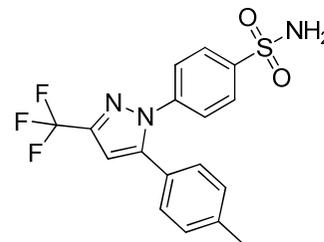
Examples of nonsteroidal anti-inflammatory drugs (NSAIDs):



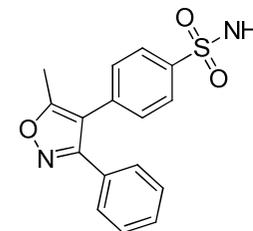
Diclofenac



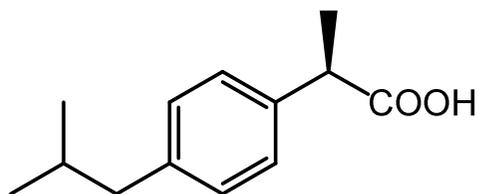
Indomethacin



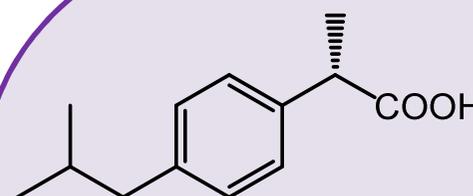
Celecoxib



Valecoxib



(R)-(-)-Ibuprofen

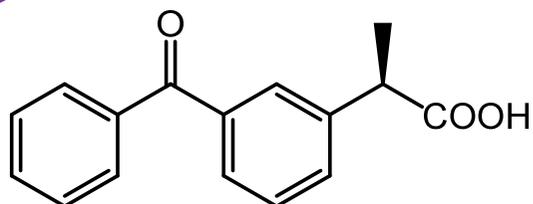


(S)-(+)-Ibuprofen

Eutomer



NSAID

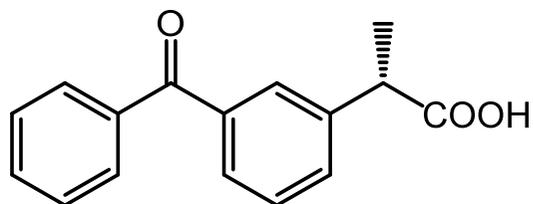


(R)-(+)- Ketoprofen

Higher affinity



Human Serum Albumin
(HSA)



(S)-(-)- Ketoprofen



higher free plasma
concentration

Evans, S. E. *et al. Trends Env Anal Chem*, 2014, 1(0), e34-e51.

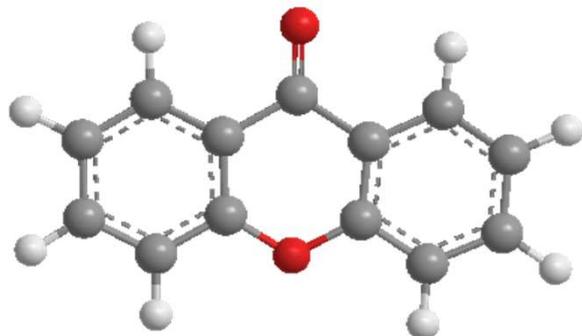


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Introduction

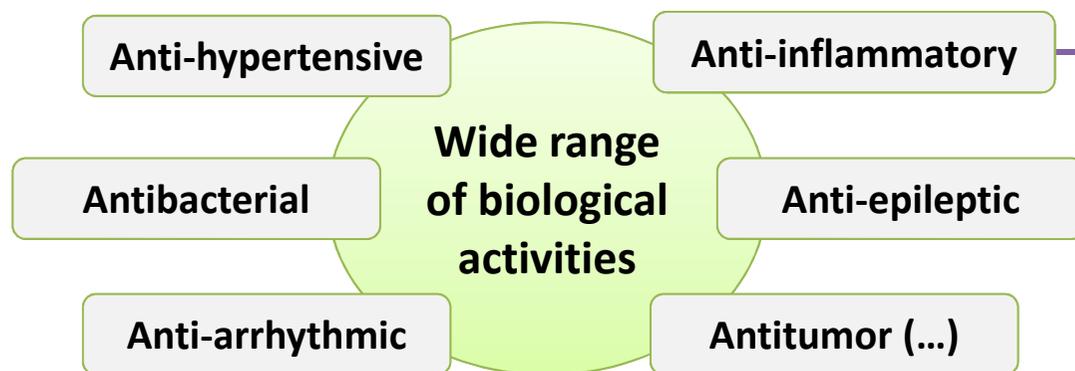
XANTHONE – A PRIVILEGED STRUCTURE



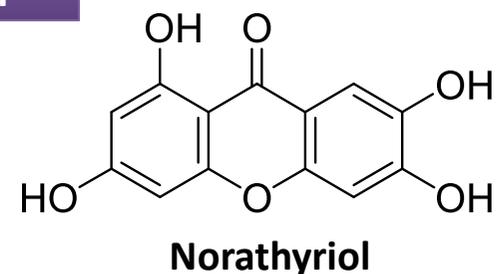
Capable of binding to multiple targets, and consequently, with appropriate structure modifications, could exhibit multiple activities



Different substituents
Natural and synthetic origin



Example



Pinto, M., E. Sousa, *et al.* *Curr Med Chem*, 2005, 12(21), 2517-2538.
Shagufta and I. Ahmad, *Eur J Med Chem*, 2016, 116, 267-280.



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New chiral derivatives of xanthenes: Synthesis and investigation of enantioselectivity as inhibitors of growth of human tumor cell lines

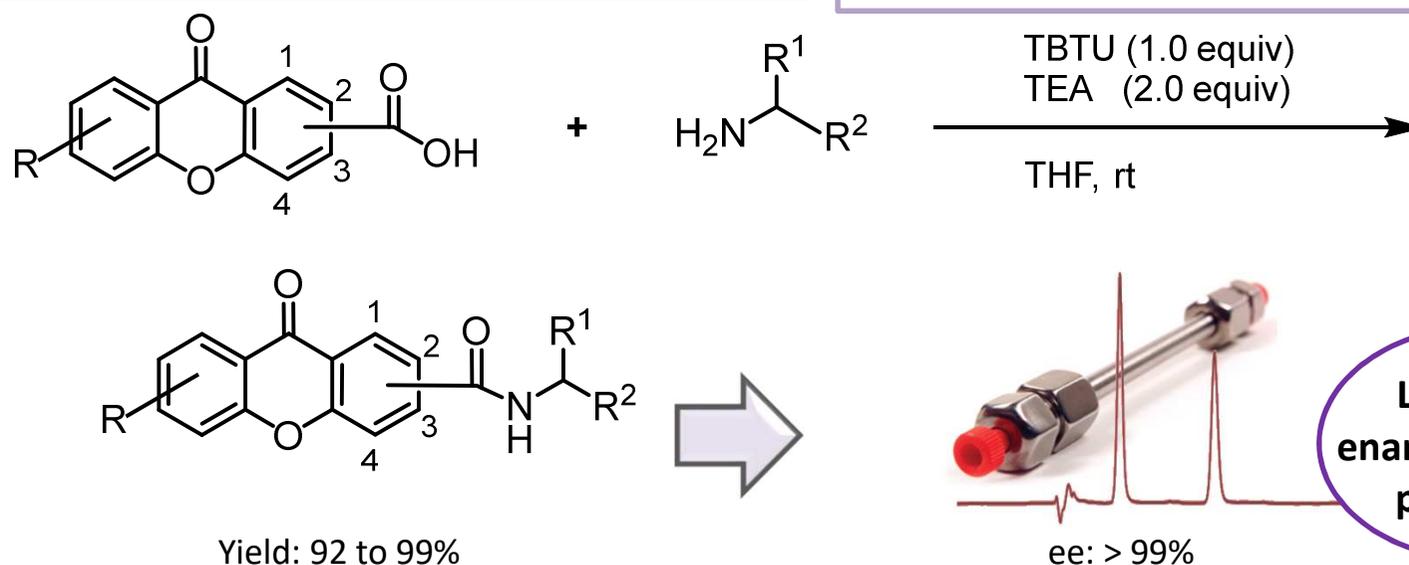
Bioorganic & Medicinal Chemistry 22 (2014) 1049–1062

Carla Fernandes^{a,b,c,†}, Kamonporn Masawang^{a,d,†}, Maria Elizabeth Tiritan^{a,c,e}, Emília Sousa^{a,b,c},
 Virgínia de Lima^f, Carlos Afonso^{a,b,c}, Hassan Bousbaa^{a,c,e}, Wanwisa Sudprasert^d, Madalena Pedro^{a,e,*},
 Madalena M. Pinto^{a,b,c,*}

Synthesis of new chiral xanthone derivatives acting as nerve conduction blockers in the rat sciatic nerve

Carla Fernandes^{a,b,1}, Laura Oliveira^{c,1}, Maria Elizabeth Tiritan^{b,d}, Luís Leitao^c, Angelo Pozzi^c,
 José Bernardo Noronha-Matos^c, Paulo Correia-de-Sá^{c,*}, Madalena M. Pinto^{a,b,**}

European Journal of Medicinal Chemistry 55 (2012) 1–11



Library of
 enantiomerically
 pure CDXs

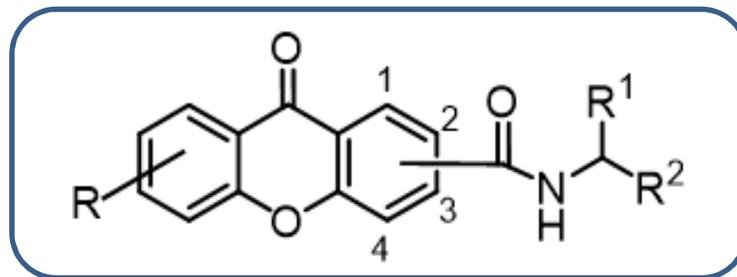
CDX: Chiral derivative of xanthone; TBTU: *O*-(Benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium tetrafluoroborate; TEA: Triethylamine; THF: Tetrahydrofuran; ee: enantiomeric excess.



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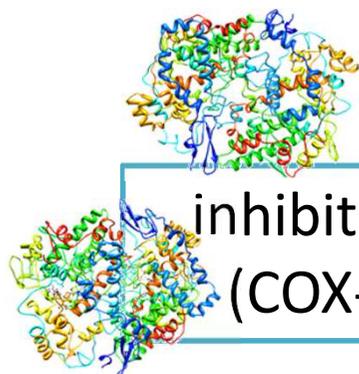
Aims



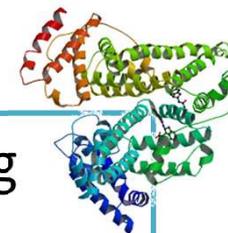
enantiomeric pairs of CDXs



in silico studies and *in vitro* assays



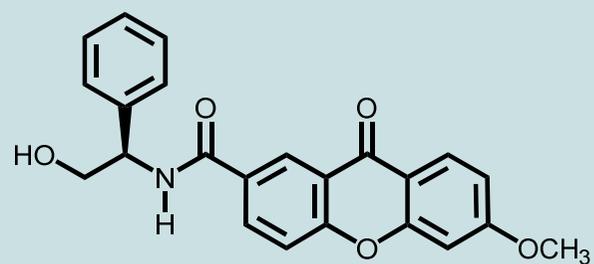
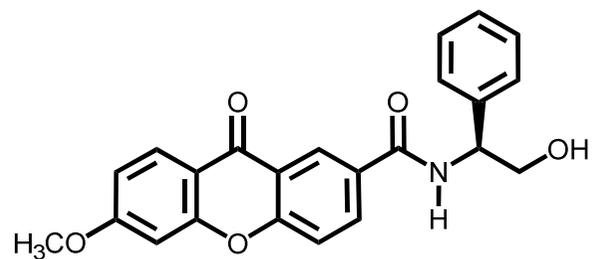
inhibition of cyclooxygenases
(COX-1 and COX-2) activity



protein binding
affinity

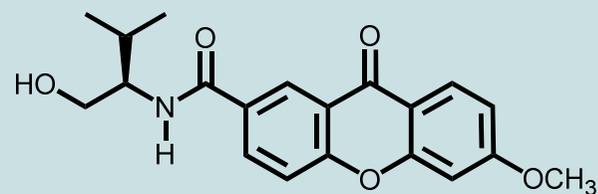
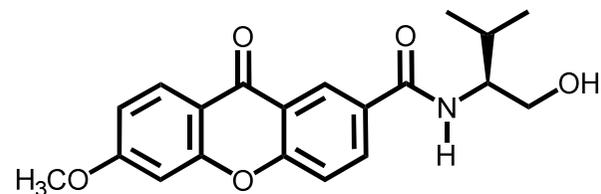


(S)-XEGOL2



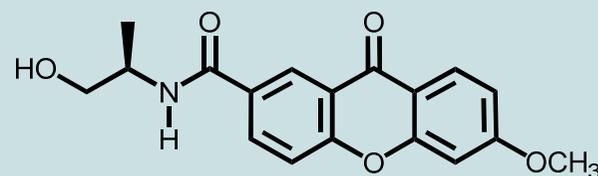
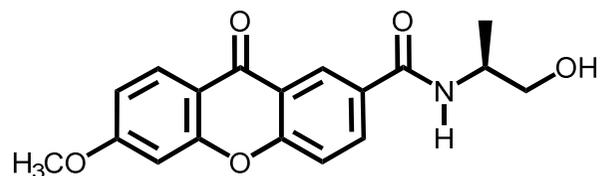
(R)-XEGOL2

(S)-XEVOL



(R)-XEVOL

(S)-X2AP1



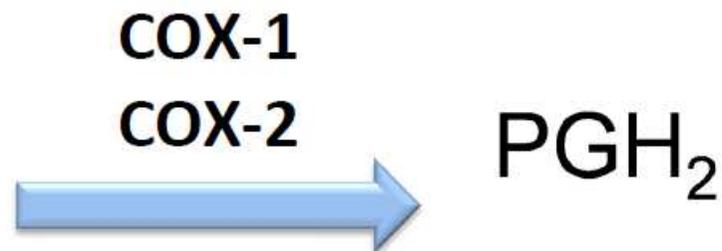
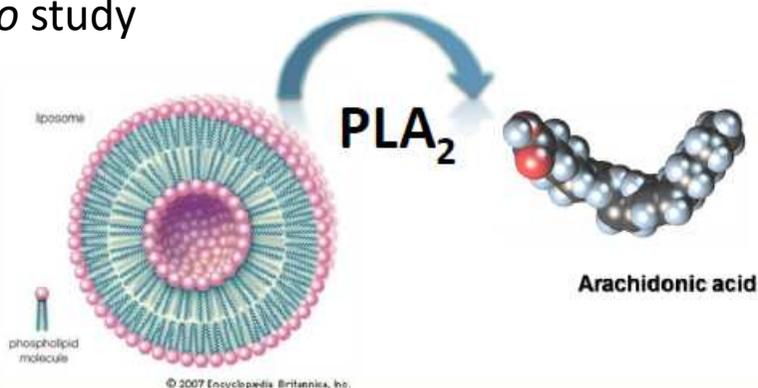
(R)-X2AP1



Results and discussion

INHIBITION OF CYCLOOXYGENASES

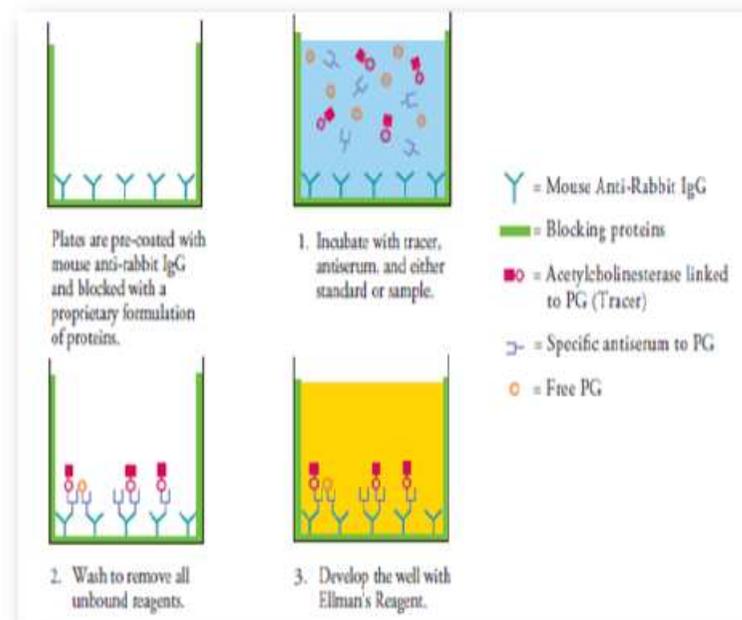
In vitro study



LUVs → ADIFAB → PLA2+DRUG

ADIFAB:
AcryloDated
Intestinal fatty
acid binding
protein

The presence of an inhibitor leads to a decrease in the release of fatty acid which generates a minor decrease of the fluorescence of ADIFAB ($\lambda_{em}=432$ nm). This allows the calculation of the relative inhibition (%).



Gelb, H., Jain, M. K., Berg, O., *Bioorg. Med. Chem. Lett.*, 1992, 1335.
Dixon D.A., et al., *J Biol Chem*, 2000, 275, 11750–11757.

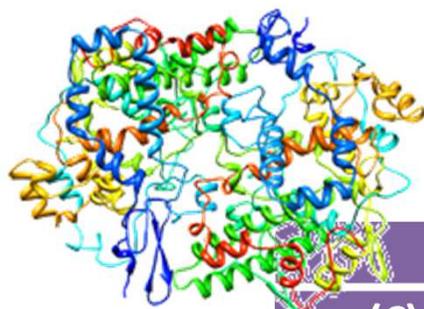


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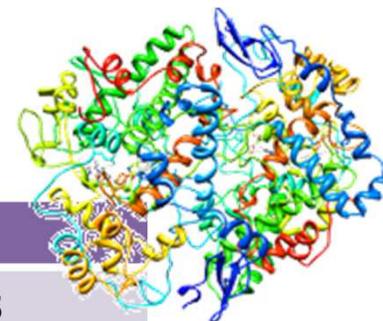


pharmaceuticals



COX-1

COX-1 and COX-2 inhibitory effects of CDXs



COX-2

CDX	COX-1	COX-2
(S)-XEGOL2	87.6 ± 2.1	80.1 ± 12.8
(R)-XEGOL2	79.6 ± 5.0	84.7 ± 5.7
(S)-XEVOL	82.9 ± 5.2	85.7 ± 4.5
(R)-XEVOL	66.8 ± 1.6	73.2 ± 0.4
(R)-X2A1P	75.2 ± 9.0	75.1 ± 7.2
(S)-X2A1P	91.7 ± 10.7	93.4 ± 11.4
Indomethacin	83.2 ± 6.4	80.7 ± 9.5

Results are given as % of inhibition and are expressed as mean ± standard deviation of two independent experiments.

The concentration of CDXs was 20 $\mu\text{mol.L}^{-1}$.
Indomethacin 1 $\mu\text{mol.L}^{-1}$ was used as positive control.



In vitro study

Solutions of increasing concentrations of CXD (Phosphate buffer, pH 7.4, I=0,1 M)

Adding HSA (final concentration 2 μ M)



$$\% \text{QUENCHING} = \frac{y_{\max}}{K_d} \frac{1}{1 + [\text{ligand}]}$$

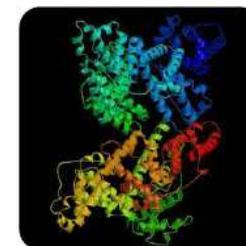
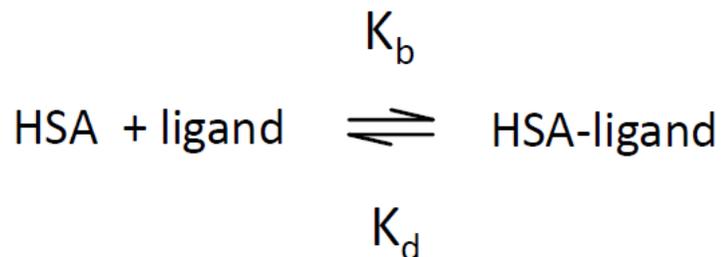
Langmuir model

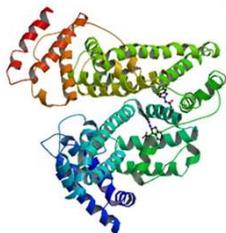
$$\% \text{QUENCHING} = \frac{y_{\max} n}{K_d} \frac{1}{1 + [\text{ligand}]}$$

Number of the binding sites (n)

$$\Delta G = -RT \ln K$$

Fluorimetric analysis
 $\lambda_{\text{exc}} = 280 \text{ nm}$, $\lambda_{\text{em}} = 330 \text{ nm}$





Results of the binding parameters

CDX	K_d (μM)	Y_{max}	n	ΔG binding (Kcal/mol) 25 °C
(S)-XEGOL2	23.6 ± 0.8	105.3 ± 0.4	1	-1.9 ± 0.1
(R)-XEGOL2	61.8 ± 6.5	109.6 ± 1.6	1	-2.4 ± 0.2
(S)-XEVOL	24.7 ± 1.1	107.4 ± 5.4	1	-1.9 ± 0.1
(R)-XEVOL	29.2 ± 0.9	108.2 ± 0.2	1	-2.0 ± 0.1
(R)-X2A1P	26.4 ± 1.2	113.2 ± 1.4	1	-1.9 ± 0.1
(S)-X2A1P	31.4 ± 2.0	116.2 ± 0.6	1	-2.0 ± 0.2

K_d (μM) < 100 μM



Ligands

CDXs and positive controls

Drawn and minimized

Decoys and known ligands

From DUD - *a directory of useful decoys*

Negative controls

NCI compound database - based on structural parameters of CDXs

Targets

Protein Data Bank of Brookhaven

ovine COX-1 (PDB code: 3n8x)

murine COX-2 (PDB code: 1cx2)

human albumin (PDB code: 2bxg)



Water molecules and crystal ligands removed

Flexible and adaptable

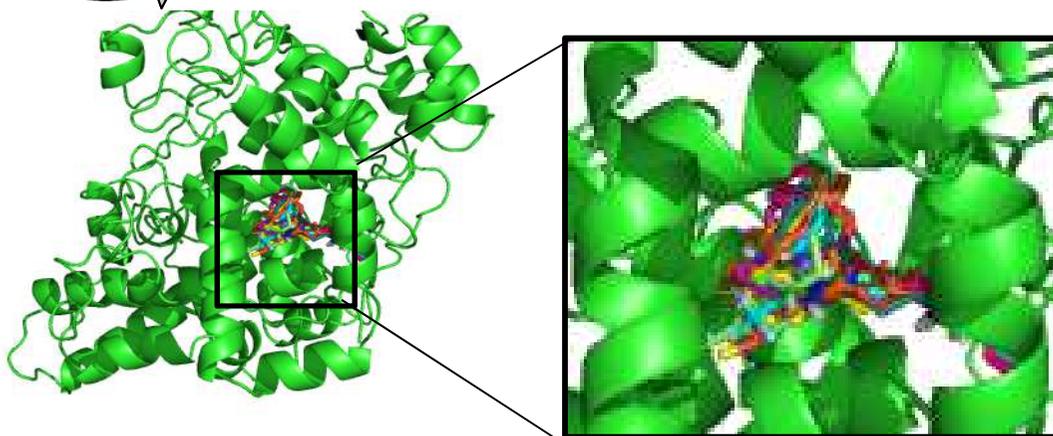
AutoDock Vina

Rigid units

Lowest binding energy docking poses of each compound were chosen



- ✓ Positive controls
- ✓ Known ligands
- ✓ Decoys
- ✓ CDXs



COX-1 binding energy (Kcal/mol)		
Known ligands	Diclofenac	-6.1
	Indomethacin	-5.1
	Naproxen	-7.8
	Piroxicam	-5.2
Ligands from database		-7.8
Decoys from database		-7.3
(R)-XEGOL2		-4.2
(S)-XEGOL2		-4.5
(R)-X2A1P		-5.3
(S)-X2A1P		-5.6
(R)-XEVOL		-3.4
(S)-XEVOL		-5.4

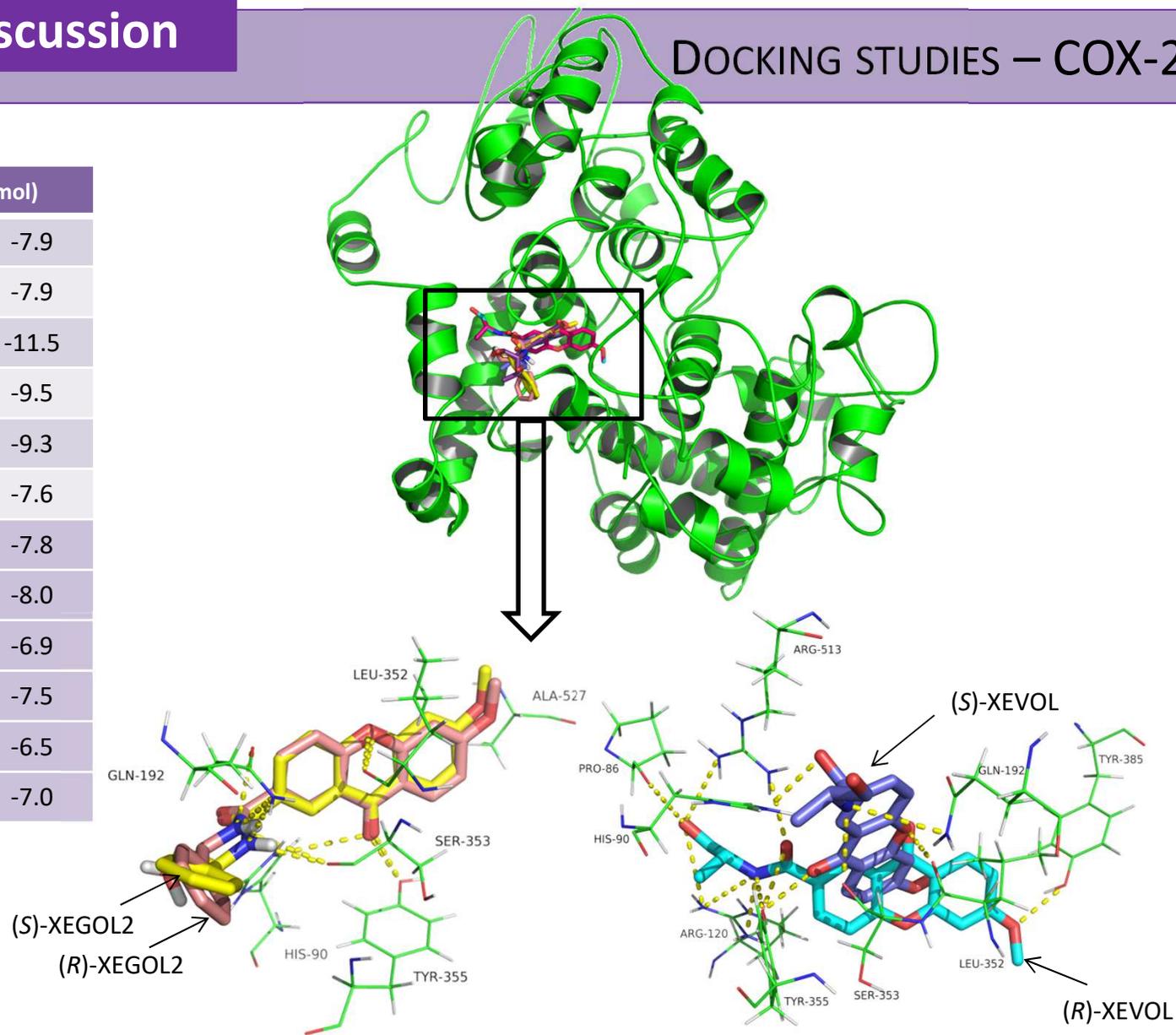


Results and discussion

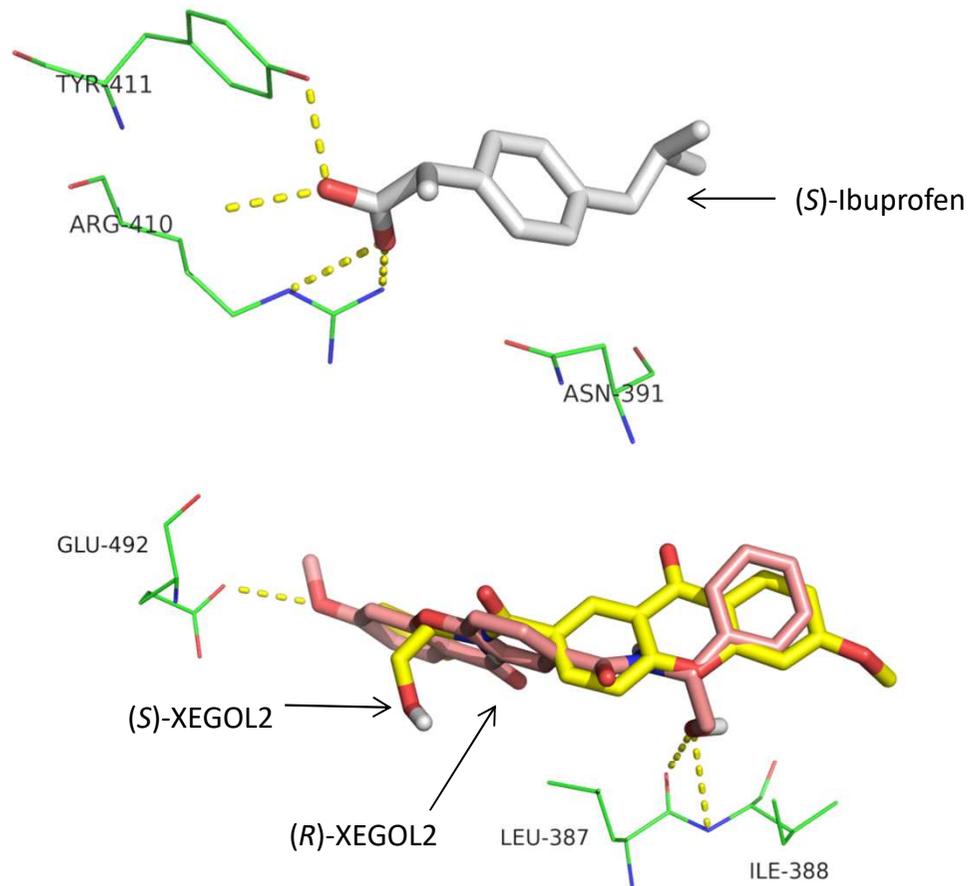
DOCKING STUDIES – COX-2

COX-2 binding energy (Kcal/mol)

Known ligands examples	Diclofenac	-7.9
	Indomethacin	-7.9
	Celecoxib	-11.5
	Valecoxib	-9.5
Ligands from database		-9.3
Decoys from database		-7.6
(<i>R</i>)-XEGOL2		-7.8
(<i>S</i>)-XEGOL2		-8.0
(<i>R</i>)-X2A1P		-6.9
(<i>S</i>)-X2A1P		-7.5
(<i>R</i>)-XEVOL		-6.5
(<i>S</i>)-XEVOL		-7.0



Albumin binding energy (Kcal/mol)		
Known ligands	Azaprozone	-5.9
	Diazepam	-7.1
	Fusidic acid	-5.8
	Ibuprofen	-7.3
	Ilophenoxid acid	-4.4
	Naproxen	-7.9
	Warfin	-8.5
(R)-XEGOL2	-7.3	
(S)-XEGOL2	-7.0	
(R)-X2A1P	-7.2	
(S)-X2A1P	-7.0	
(R)-XEVOL	-7.2	
(S)-XEVOL	-7.2	



Conclusions

Considering the inhibition of cyclooxygenases (COX-1 and COX-2):

- all the CDXs evaluated exhibited COX-1 and COX-2 inhibition potential in *in vitro* assays,
- the inhibitory effects were very similar for the same enantiomeric pair as well as for both COXs,
- no significant difference was found between known ligands and decoys docking scores on COX-1; therefore, no reliable conclusions can be taken from test ligands binding affinity to COX-1,
- XEGOL2 enantiomeric pair is predicted to show more affinity towards COX-2, presenting docking scores similar to known ligands, such as diclofenac and indomethacin.

Considering the HSA binding affinity:

- all CDXs demonstrated to bind with high affinity to HSA potential in *in vitro* assays,
- XEGOL2 enantiomeric pair exhibited enantioselectivity,
- *in silico* studies CDXs confirmed that they bind to albumin serum protein, as they have docking scores similar to positive controls such as ibuprofen and diazepam.



Acknowledgments

This research was partially supported by the Structured Program of R&D&I INNOVMAR –Innovation and Sustainability in the Management and Exploitation of Marine Resources (reference NORTE-01-0145-FEDER-000035, Research Line NOVELMAR), funded by the Northern Regional Operational Programme (NORTE2020) through the European Regional Development Fund (ERDF) and by Foundation for Science and Technology (FCT) and COMPETE under the projects PTDC/MAR-BIO/4694/2014 (POCI-01-0145-FEDER-016790) and COXANT–CESPU- 2016.



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