

1 Article

2 Exploiting carbon and nitrogen compounds for 3 enhanced energy and resource recovery

4 Bahareh Kokabian ¹, Veera Gnanaswar Gude ^{2*}

5 Civil & Environmental Engineering Department, Mississippi State University, Mississippi State, MS 39762

6 * Correspondence: gude@cee.msstate.edu; Tel.: +1-662-325-0345

7

8 **Abstract:** Microbial desalination cells (MDCs), a recent technological discovery, allow for
9 simultaneous wastewater treatment and desalination of saline water with concurrent electricity
10 production. The premise for MDC performance is based on the principles that bioelectrochemical
11 (BES) systems convert wastewaters into treated effluents accompanied by electricity production and
12 the ionic species migration (i.e. protons) within the system facilitates desalination. One major
13 drawback with microbial desalination cells (MDCs) technology is its unsustainable cathode chamber
14 where expensive catalysts and toxic chemicals are employed for electricity generation. Introducing
15 biological cathodes may enhance the system performance in an environmentally-sustainable manner.
16 This study describes the use of autotrophic microorganism such as algae and Anammox bacteria as
17 sustainable biocatalyst/biocathode in MDCs. Three different process configurations of photosynthetic
18 MDCs (using *Chlorella vulgaris*) were evaluated for their performance and energy generation
19 potentials. Static (fed-batch, SPMDC), continuous flow (CFPMDC) and a photobioreactor MDC
20 (PBMD, resembling lagoon type PMDCs) were developed to study the impact of process design on
21 wastewater treatment, electricity generation, nutrient removal, and biomass production and the
22 results indicate that PMDCs can be configured with the aim of maximizing the energy recovery
23 through either biomass production or bioelectricity production. In addition, the microbial
24 community analysis of seven different samples from different parts of the anode chamber, disclosed
25 considerable spatial diversity in microbial communities which is a critical factor in sustaining the
26 operation of MDCs. This study provides the first proof of concept that anammox mechanism can be
27 beneficial in enhancing the sustainability of microbial desalination cells to provide simultaneous
28 removal of ammonium from wastewater and contribute in energy generation.

29

30 **Keywords:** anammox bacteria, microbial desalination, microalgae, photosynthesis, nutrients,
31 bioelectricity

32 **PACS:** J0101

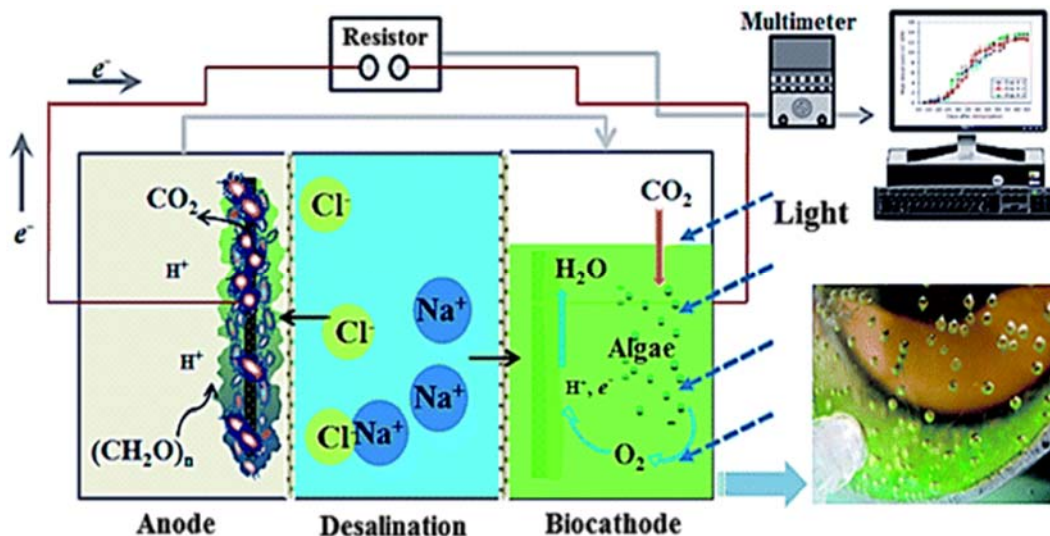
33

34 1. Introduction

35 The energy and water production issues are intertwined and cannot be addressed in isolation
36 [1-4]. Wastewater treatment and desalination, in particular, are energy consuming processes which
37 can have detrimental effects on the environment [3]. Integrated solutions that utilize waste sources
38 to generate energy, which in turn, can be used to produce freshwater are attractive options to address
39 current energy and water issues [5]. In this context, bioelectrochemical systems have evolved as a

40 novel technology to convert wastes into valuable energy [6]. Bioelectrochemical systems can be
 41 employed to generate clean electricity, or high value energy or chemical products from various
 42 wastewater sources and organic or inorganic wastes that can serve as fuel feedstock for electroactive
 43 bacteria [7]. Microbial desalination cells (MDCs) are based on an integrated configuration in which,
 44 wastewater and saline water sources can be treated simultaneously without any external power input
 45 or mechanical energy or pressure application [8]. This process offers multiple benefits of energy and
 46 resource (water and nutrients) recovery while eliminating environmental pollution.

47 Photosynthetic microorganisms can be used in PMDCs for accomplishing proper utilization
 48 of carbon and nutrient compounds (Figure 1)[9]. Their role in PMDCs can be further controlled
 49 specifically for bioelectricity production or biomass production which depends on the process
 50 configuration. Microalgae provide in-situ oxygen production which can serve as an electron acceptor
 51 in the electron transfer process while utilizing organic carbon, nitrogen and phosphorous compounds
 52 for growth [10]. On the other hand, conventional removal of nitrogenous compounds by nitrification-
 53 denitrification process from waste water in waste water treatment plants requires considerable
 54 amount of energy and costs. Anammox process which comes from Anaerobic ammonia oxidation
 55 (anammox) is an emerging microbial process for conversion of ammonium to nitrogen gas under
 56 anaerobic condition with potential energy and cost savings [11]. Autotrophic bacteria create a
 57 bypass to oxidize ammonia to nitrogen gas by nitrite omitting the need for organic carbon source.
 58 Partially nitrification of ammonia to nitrite instead of nitrate, allows for about 40% saving on energy
 59 used for aeration. In addition, due to the autotrophic nature of these bacteria their growth rate is
 60 slow and thus, less biosolids are produced during this process. All of these benefits, make Anammox
 61 based nitrogen removal process more cost effectiveness (cost reduction of up to 60%) and less
 62 greenhouse gas emission compared to conventional nitrification-denitrification process [12].
 63



64
 65 **Figure 1.** A photosynthetic microbial desalination cell (PMDC)
 66

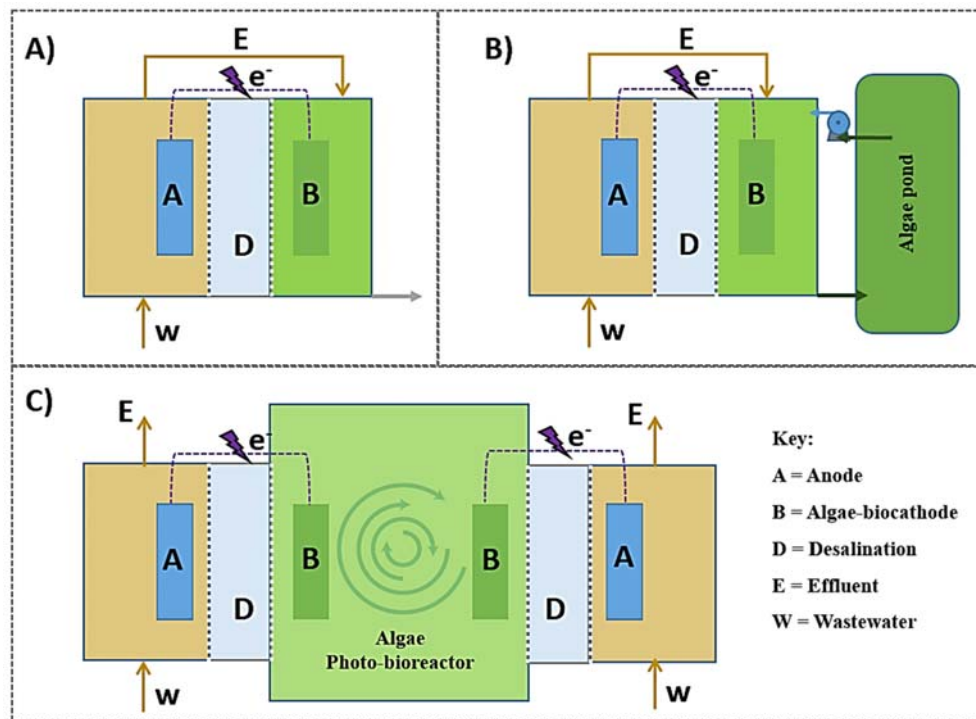
67 This research article presents the preliminary and proof-of-concept studies of photosynthetic
 68 (microalgal) and autotrophic (anammox bacteria) as biocathodes in microbial desalination cells. The
 69 following sections describe the experimental details and preliminary results.

70 2. Materials and Methods

71 2.1. MDC configuration and operation

72 Three-chamber inside circular shaped MDCs with 7.2 cm diameter were made using plexi-
 73 glass. Anion exchange membrane (AEM, AMI7001, Membranes International) separated the anode
 74 and the desalination chambers, while cation exchange membrane (CEM, CMI7000, Membranes
 75 International) separated the cathode and the desalination compartments. Both membranes were
 76 preconditioned by immersing in 5% NaCl solution at 40 °C for 24 h and rinsed with distilled water
 77 (DI) water prior to use, to allow for membrane hydration and expansion as recommended by the
 78 supplier. Carbon cloth covered with stainless steel mesh were used as electrodes with 16 cm² surface
 79 area. Prior to use, both electrodes were washed first with 1 N HCl solution and then with 1 N NaOH
 80 and finally rinsed with deionized water. The electrodes are then soaked in DI water over a night prior
 81 to use to remove any excess residues [13]. Anode and cathode electrodes were connected through a
 82 titanium wire. The working volume of anode, desalination, and cathode chambers after inserting the
 83 electrodes were 37, 28, and 37 mL respectively. Three different operational modes, namely, static
 84 (SPMDC, Figure 2a), continuous flow (CFPMDC, Figure 2b), and photobioreactor (PBMDC, Figure
 85 2c) were used for photosynthetic MDC to assess its performance in terms of electricity generation,
 86 biomass production and nutrient removal capacities. SPMDC was run in batch cycles. In each test,
 87 new wastewater, fresh algae medium and fresh salt solution were used in PMDCs. In the continuous
 88 mode, the algae catholyte was circulated using a peristaltic pump. Two MDC biocathodes were
 89 assembled to the large photo-bioreactor (5 liter volume). This configuration was called Photo-
 90 bioreactor MDC (PBMDC).

91



92

93 **Figure 2.** PMDC process configurations: a) PMDC with an algae biocathode under fed batch (static) operational
 94 mode (SPMDC); b) PMDC with an algae biocathode under continuous flow operational mode (CPMDC); c)
 95 PMDC with an algae biocathode connected to a photo-bioreactor (PBMDC)

96 Anammox biomass was provided by Hampton Roads Sanitation District in Virginia and was
97 divided in three bottles under anaerobic condition in the shaker incubator at 35°C and 150 rpm. The
98 culture contained NH_4Cl , 382 mg L⁻¹; NaNO_2 , 493 mg L⁻¹; KHCO_3 , 200 mg L⁻¹; KH_2PO_4 , 27 mg L⁻¹;
99 $\text{FeSO}_4 \times 7\text{H}_2\text{O}$, 9.0 mg L⁻¹; EDTA, 5.0 mg L⁻¹; $\text{MgSO}_4 \times 7\text{H}_2\text{O}$, 240 mg L⁻¹; $\text{CaCl}_2 \times 2\text{H}_2\text{O}$, 143 mg L⁻¹ and
100 300 µl of trace metal solution. The trace solution contained $\text{ZnSO}_4 \times 7\text{H}_2\text{O}$, 1,247 mg L⁻¹; $\text{MnSO}_4 \times \text{H}_2\text{O}$,
101 1,119 mg L⁻¹; $\text{CuSO}_4 \times 5\text{H}_2\text{O}$, 44 mg L⁻¹; $\text{Al}_2(\text{SO}_4)_3 \times 14\text{H}_2\text{O}$, 201.5 mg L⁻¹; $\text{Na}_2\text{MoO}_4 \times 2\text{H}_2\text{O}$, 129 mg L⁻¹;
102 $\text{CoCl}_2 \times 6\text{H}_2\text{O}$, 30 mg L⁻¹; KCl, 100 mg L⁻¹; EDTA, 975 mg L⁻¹ that provides micronutrients needed for
103 microbial growth of anammox bacteria [14]. After about two months reactivation process, this sludge
104 was transferred to the cathode chamber of MDC. The anode chamber of MDC was inoculated with
105 30 ml of acclimatized anaerobic sludge.

106

107 2.2. Analyses and calculations

108 The voltage across a 1 kΩ external resistor was recorded every 15 min by a digital multimeter
109 (Fluke, 287/FVF). The current was calculated using Ohm's law, $I = V/R$. The power density was
110 calculated ($P = V/I$) as per the volumes of the anode/cathode chambers. COD tests were carried out
111 using standard methods. The nitrogen, as nitrate ($\text{NO}_3\text{-N}$), and phosphorus ($\text{PO}_4^{3-}\text{-P}$) concentrations
112 were measured by colorimetric methods according to the method of Hach (Methods 8039 & 8114).
113 Electrical conductivity, total dissolved solids (TDS) removal, and salinity removal were recorded
114 using a conductivity meter (Extech EC400 ExStik Waterproof Conductivity, TDS, Salinity, and
115 Temperature Meter). The algae concentration was determined by measuring the absorbance of the
116 cell suspension at a wavelength of 620 nm and then converting it to dry weight of biomass in volume
117 by a calibration curve. pH was measured using a pH meter (Orion 720A+ advanced
118 ISE/pH/mV/ORP). Dissolved oxygen was measured using YSI 5100 system. Continuous illumination
119 on the algae cathode chamber was provided by CFL white light at 60W (276 mmol per m² per second).

120

121

122 3. Results and Discussion

123 3.1. Photosynthetic microbial desalination cells

124 The COD removals in the anode along with pH changes in the cathode chamber for each cycle are
125 shown in Table 1. Low COD removal rates (less than 30%) were observed in these tests. The low COD
126 removal rates in all cycles suggest that substrate limitation was not the reason for voltage drop in the
127 cells, but the decrease in the conductivity of the solution in the middle chamber which increases the
128 ohmic resistance of the cells could be the reason for voltage drop [8]. The other reason could be due
129 to the increase in pH of the cathode solution and increased pH imbalance between the anode and
130 cathode chambers. pH is an important factor in performance of bioelectrochemical systems since it
131 affects the biological activity of the microorganisms [15]. The increase in pH which is typically caused
132 by consumption of the protons and photosynthetic activity of the algae, slows down the ORR rate
133 and is often reported as the limiting factor in power production [16]. In addition to pH, ORR rate is
134 affected by fouling and biofouling which may hinder the transfer of oxygen to the electrode surface.
135 It can be concluded that the performance of the PMDC depends on the photosynthetic activity of the
136 algae and the bioelectrochemical function of the biofilm on the cathode electrode which was reported
137 in our previous study [10]. Due to the high buffer concentration in the anode chamber, the pH did
138 not change significantly; however, the pH in the cathode chamber increased.

139

140 Table 1. Organic carbon removal and pH changes during the four batch tests of SPMDC

141

	Batch 1	Batch 2	Batch 3	Batch 4
COD removal	31% ± 3.4	22.5% ± 0.18	29% ± 1.63	28% ± 1.08
Initial Cathode pH	6.6	6.9	6.7	7.3
Final Cathode pH	10.3	11	10.6	10.83

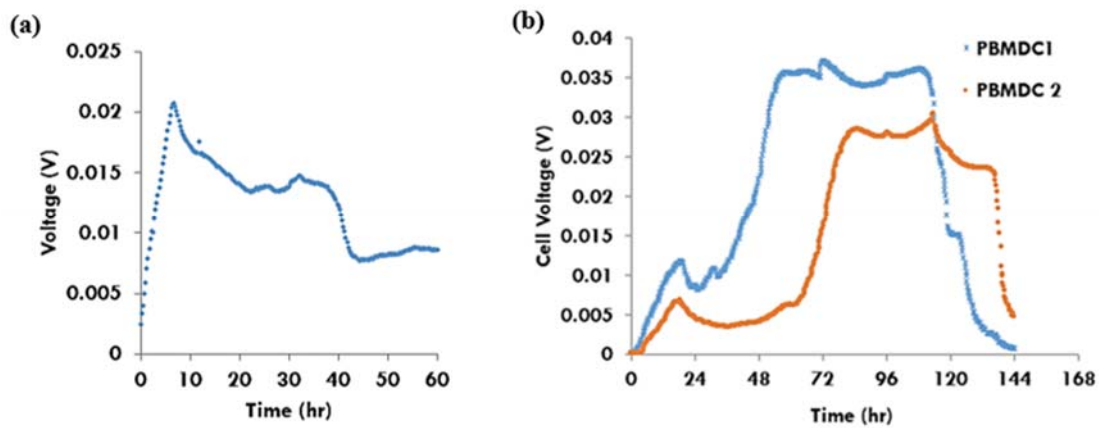
142

143

144

145

The voltage generation profiles for continuous flow PMDC and photobioreactor PMDCs are shown in **Figure 3**.



146

147

148

149

Figure 3. Voltage generation profiles in (a) PMDC with an algae biocathode under continuous flow operational mode (CPMDC); b) PMDC with an algae biocathode connected to a photo-bioreactor (PBMD)

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

Due to the potential limitation for algae growth in static PMDCs, the cathode chamber of PMDCs was connected with a peristaltic pump to a 1000 mL bioreactor containing algae cells in suspension (Figure 2b). About 100 mg of sodium bicarbonate was added to the algae container with 1000 mL of growth medium to provide the inorganic carbon source for photosynthetic activity by algae cells. The electricity generation for continuous flow PMDC (CFPMDC) is shown in Figure 3a. The Maximum voltage obtained during this cycle (20 mV) was lower than SPMDC. It has been reported before that immobilized cells produce higher electricity than suspended algae [17]. This could be due to the improved oxygen reduction with easy electron transfer when algae cells deposit on the surface of the electrode whereas in the continuous mode less amount of cells settled at the surface due to the suspension. This indicates the catalytic role of algae for oxygen reduction beside its role as an oxygen supplier. The catalytic role of algae cells was reported by Cai et al. [18] in a previous study where photosynthetic biocathode generated higher electricity in comparison to the abiotic that was aerated to have the same level of dissolved oxygen (DO). Walter et al. [19] performed cyclic voltammetry analysis of a photo-biocathode and abiotic control electrode and found reduction peak for biocathode whereas no peak was observed for abiotic control electrode.

165

166

167

168

A new PMDC configuration integrating two PMDCs with one large algae biocathode chamber was developed (Figure 2c). The algae cells were maintained in suspension with a mechanical mixer. This new configuration was named as photo-bioreactor MDC (PBMD). The advantage of this system is that two MDCs could work at the same time with one common photosynthetic biocathode

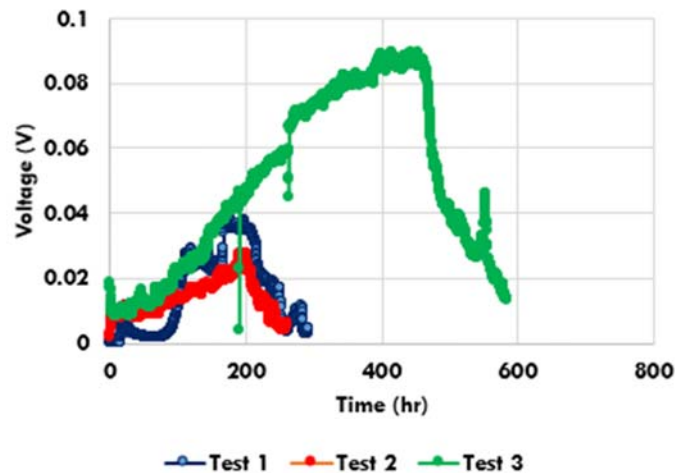
169 chamber which may increase the overall efficiency of the system. The cell voltage for this new
 170 configuration is depicted in Figure 3b. The electricity generation profiles for the two cells followed a
 171 similar pattern. The system reached its maximum cell voltage after 50 hours and could maintain its
 172 maximum voltage for almost 50 hours. The maximum cell voltage was still lower than the SPMDC
 173 due to the aforementioned reasons in the previous section. The other reason could be due to the very
 174 large ratio of the cathode volume to electrode surface which may decrease the efficiency of this
 175 system. It has been reported before that scaled MFCs could not generate power as well as smaller
 176 scale MFCs due to the higher internal resistance that was created [20]. Our new system however,
 177 operated very well in large volume algae production. The concentration of algae cells increased from
 178 135 mgL⁻¹ to 362 mgL⁻¹. The system could work longer due to the better design of the system for
 179 growth of algae cells compared to CFPMD. pH and DO profiles are similar to CPMDC.

180

181 3.2. Anammox microbial desalination cells

182 Voltage profiles generated by Anammox MDC (AnxMDC) for three batch experiments are
 183 shown in Figure 4. Since we did not provide any chemical catalyst or aeration in the cathode chamber,
 184 the production of electricity indicates the effective role of anammox bacteria as biocathode and
 185 Nitrite/Nitrate as electron acceptor. Increase of maximum power for the third batch experiment
 186 compared to the first and second test demonstrates an improvement in the catalytic activity of the
 187 biofilm. The maximum produced voltage was 0.0896 V which is equal to power density of 0.114 W/m³.
 188 These data highlights the fact that electricity generation by these cells has the potential to improve by
 189 several batch tests and better formation of the biofilms on the electrodes.

190



191

192 Figure 4. Voltage generation by AnxMDC during three batch tests

193

194 Only 29% of the organic carbon in the anode chamber in the third batch was removed to generate
 195 electricity. Coulombic efficiencies and salt removals increased over the three batch tests. The
 196 coulombic efficiencies for glucose oxidation were 3.4%, 6.02% and 52.7% respectively for the three
 197 batch tests while the coulombic efficiencies for nitrite/nitrate reduction were 17.5%, 35.6% and 99%
 198 respectively for the three tests. The improvement in coulombic efficiency over the three batch tests
 199 indicates the improvement of microbial growth on electrodes after several batches. Due to the higher
 200 electricity production and longer operating time, salinity removal was also higher for the third test.

201 4. Conclusions

202 Photosynthetic MDCs (PMDCs) can be operated either in fed-batch, batch or continuous flow
203 conditions for maximizing the energy recovery from wastewater. The findings of this study
204 demonstrate the beneficial use of photosynthetic microorganisms as biocathodes or biocatalysts in
205 microbial desalination cells to produce oxygen, algae biomass and nutrient removal from wastewater.
206 Different efficiencies of PMDC (Static, Continuous and Photobioreactor PMDC) observed in this
207 study show that the design and process configuration play a critical role in the overall efficiency of
208 the system. If harvesting higher biomass is the major target, open large scale systems are more
209 suitable whereas for small systems, closed and static systems are more beneficial. The nutrient
210 removal capability of PMDCs provides the opportunities to utilize agricultural, food and other
211 industrial wastewaters that are rich in nutrients for use as catholyte medium.

212 The study demonstrated the feasibility of using an autotrophic microbial culture containing
213 anammox bacteria as the biocathode of MDC to contribute in simultaneous energy generation and
214 wastewater treatment. Batch experiments improved the coulombic efficiency of the system as well as
215 the nitrite and ammonium removal of the wastewater. A maximum power of 0.114 W/m³ with more
216 than 90% removal of ammonium was achieved in this system. The finding of this research showed
217 that this system is more useful for wastewaters with low C/N ratio to suppress the possibility for
218 growth of heterotrophic bacteria. The proposed MDC configurations demonstrated great potential to
219 replace conventional energy intensive nutrient removal process while at the same time generating
220 clean energy and water and possibly valuable microalgae biomass.

221
222 **Acknowledgments:** This research was supported by the Office of Research and Economic Development
223 (ORED), Bagley College of Engineering (BCoE), and the Department of Civil and Environmental Engineering
224 (CEE) at Mississippi State University. The author would like to acknowledge the funding support from
225 the United States Environmental Protection Agency (USEPA) under P3 (People, Planet, and Prosperity) Awards
226 program through the grants [SU835721](#) and [SU835722](#).

227 **Author Contributions:** Gude V.G. conceived and designed the experiments; B.K. performed the experiments;
228 B.K analyzed the data; V.G.G and B.K. wrote the paper.”

229

230 **Conflicts of Interest:** The authors declare no conflict of interest. The founding sponsors had no role in the design
231 of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the
232 decision to publish the results.

233

234 References

- 235 1. Gude, V. G., Nirmalakhandan, N., & Deng, S. (2010). Renewable and sustainable approaches for
236 desalination. *Renewable And Sustainable Energy Reviews*, 142641-2654.
- 237 2. Gude, V. G. (2011). Energy consumption and recovery in reverse osmosis. *Desalination & Water Treatment*,
238 36(1-3), 239-260.
- 239 3. Gude, V. G. (2015a). Energy and water autarky of wastewater treatment and power generation systems.
240 *Renewable and Sustainable Energy Reviews*, 45, 52-68.
- 241 4. Gude, V. G. (2015b). Energy storage for desalination processes powered by renewable energy and waste
242 heat sources. *Applied Energy*, 137, 877-898.
- 243 5. Gude, V. G., Kokabian, B., Gadhamshetty, V. (2013). Beneficial Bioelectrochemical Systems for Energy,
244 Water, and Biomass Production. *Journal of Microbial & Biochemical Technology*6: 005 doi: 10.4172/1948-
245 5948.S6-005

- 246 6. Logan, B. E., & Rabaey, K. (2012). Conversion of wastes into bioelectricity and chemicals by using microbial
247 electrochemical technologies. *Science (Washington)*, 337(6095), 686-690.
- 248 7. Kokabian, B., & Gude, V. G. (2015a). Role of membranes in bioelectrochemical systems. *Membrane water*
249 *treatment*, 6(1), 53-75
- 250 8. Cao, X., Huang, X., Liang, P., Xiao, K., Zhou, Y., Zhang, X., & Logan, B. (2009). A new method for water
251 desalination using microbial desalination cells. *Environmental Science And Technology*, 43(18), 7148-7152.
- 252 9. Kokabian, B., & Gude, V. G. (2013). Photosynthetic microbial desalination cells (PMDCs) for clean energy,
253 water and biomass production. *Environmental Science. Processes & Impacts*, 15(12), 2178-2185.
254 doi:10.1039/c3em00415e
- 255 10. Kokabian, B., & Gude, V. G. (2015b). Sustainable photosynthetic biocathode in microbial desalination cells.
256 *Chemical Engineering Journal*, 262, 958-965. doi:10.1016/j.cej.2014.10.048
- 257 11. Terada, A., Zhou, S., & Hosomi, M. (2011). Presence and detection of anaerobic ammonium-oxidizing
258 (anammox) bacteria and appraisal of anammox process for high-strength nitrogenous wastewater
259 treatment: a review. *Clean Technologies & Environmental Policy*, 13(6), 759-781. doi:10.1007/s10098-011-
260 0355-3
- 261 12. Siegrist, H., Salzgeber, D., Eugster, J., & Joss, A. (2008). Anammox brings WWTP closer to energy autarky
262 due to increased biogas production and reduced aeration energy for N-removal. *Water Science &*
263 *Technology*, 57(3), 383-388.
- 264 13. Pandit, S., Nayak, B. K., & Das, D. (2012). Microbial carbon capture cell using cyanobacteria for simultaneous
265 power generation, carbon dioxide sequestration and wastewater treatment. *Bioresource Technology*, 10797-
266 102. doi:10.1016/j.biortech.2011.12.067
- 267 14. Rothrock, M. J., Vanotti, M. B., Szögi, A. A., Gonzalez, M. G., & Fujii, T. (2011). Long-term preservation of
268 anammox bacteria. *Applied Microbiology And Biotechnology*, 92(1), 147-157.
- 269 15. Zhang, F., Jacobson, K., Torres, P., & He, Z. (2010). Effects of anolyte recirculation rates and catholytes on
270 electricity generation in a litre-scale upflow microbial fuel cell. *Energy And Environmental Science*, 3(9),
271 1347-1352.
- 272 16. Commault, A., Weld, R., Lear, G., & Novis, P. (2014). Photosynthetic biocathode enhances the power output
273 of a sediment-type microbial fuel cell. *New Zealand Journal Of Botany*, 52(1), 48-59.
274 doi:10.1080/0028825X.2013.870217
- 275 17. Zhou, M., He, H., Jin, T., & Wang, H. (2012). Power generation enhancement in novel microbial carbon
276 capture cells with immobilized *Chlorella vulgaris*. *Journal Of Power Sources*, 214216-219.
277 doi:10.1016/j.jpowsour.2012.04.043
- 278 18. Cai, P., Xiao, X., He, Y., Li, W., Zang, G., Sheng, G., & ... Yu, H. (2013). Reactive oxygen species (ROS)
279 generated by cyanobacteria act as an electron acceptor in the biocathode of a bio-electrochemical system.
280 *Biosensors & Bioelectronics*, 39(1), 306-310.
- 281 19. Walter, X. A., Greenman, J., & Ieropoulos, I. A. (2013). Oxygenic phototrophic biofilms for improved cathode
282 performance in microbial fuel cells. *Algal Research*, 2(3), 183-187.
- 283 20. Logan, B. E. (2010). Scaling up microbial fuel cells and other bioelectrochemical systems. *Applied*
284 *Microbiology & Biotechnology*, 85(6), 1665-1671.
- 285

