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Targeting the Trypanosome Alternative Oxidase (TAO) as Promising Chemotherapeutic Approach for African Trypanosomiasis

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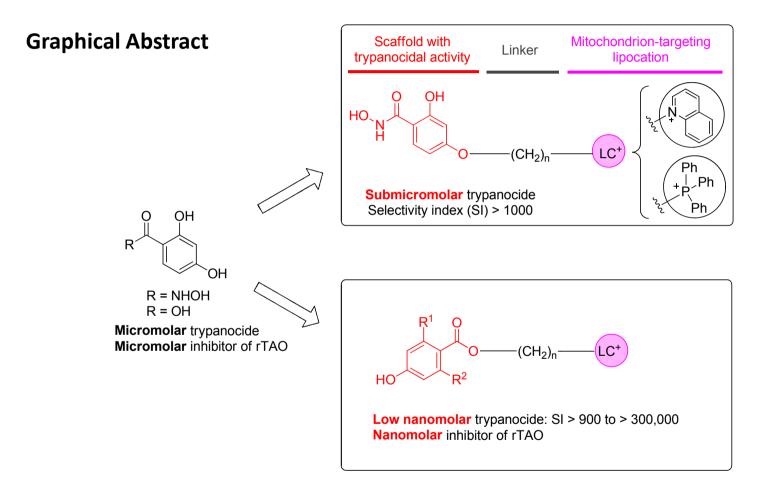


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Targeting the trypanosome alternative oxidase (TAO) as promising chemotherapeutic approach for African trypanosomiasis







Abstract:

In *Trypanosoma brucei*, a parasite that causes African trypanosomiasis in humans (sleeping sickness) and in livestock (nagana) throughout sub-Saharan Africa, the trypanosome alternative oxidase (TAO) is essential for the respiration of bloodstream form parasites (i.e. the human-infective form). Since TAO has no counterpart in mammalian cells and it is conserved among *T. brucei* subspecies, it has been validated as a promising target for the chemotherapy of African trypanosomiasis.

We present here a successful approach to boost the activity of TAO inhibitors based on the conjugation of the inhibitor with lipophilic cations (LC) that can cross lipid bilayers by non-carrier mediated transport, and thus accumulate specifically into mitochondria, driven by the plasma and mitochondrial transmembrane potentials (negative inside). This design afforded several LC–TAO inhibitor conjugates active in the submicromolar to low nanomolar range against wild type and resistant strains of African trypanosomes (*T. b. brucei, T. congolense*), with selectivity over human cells >500.

Keywords: Trypanosome alternative oxidase (TAO) inhibitor; *Trypanosoma brucei*, sleeping sickness, lipocation, mitochondrion





Introduction

African trypanosomiasis

Population at risk: 65 million in sub-Saharan Africa



- Caused by two subspecies of Trypanosoma brucei (T. b.) gambiense (g-HAT; 98% of reported sleeping sickness cases) and T. b. rhodesiense (r-HAT)
- > Transmitted by the tsetse fly
- Occurs in two stages: the early stage (stage 1) with non-specific symptoms, often unor misdiagnosed and the late stage (stage 2) where the parasite crosses the blood-brain barrier, causing serious neurological disorders including sleep cycle disruptions,

neurological manifestations, and progressive mental deterioration

- Mortality without treatment: 100 %
- Economic burden: infection of livestock (nagana)

Source: https://www.dndi.org/wp-content/uploads/2017/08/Factsheet_2016_HAT.pdf





Respiration of T. brucei as target for chemotherapy

African trypanosomes adapt their energy metabolism depending on substrate availability:

• **The procyclic** form of the parasite (present in the tsetse fly vector) has a fully functional cytochrome-dependent respiratory chain.

- Bloodstream form (BSF) trypanosomes (the human infective form) use the glycolysis as main source of ATP
 - No cytochrome respiratory pathway
 - No oxidative phosphorylation

Respiration of BSF trypanosomes is dependent on a cyanide-insensitive alternative terminal oxidase (TAO)



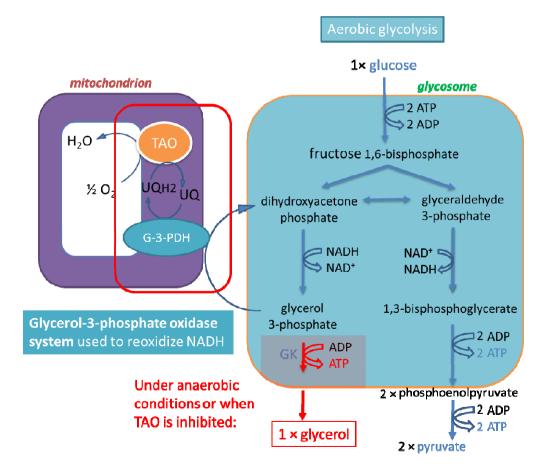






Respiration of T. brucei as target for chemotherapy

As BSF trypanosomes have no functional respiratory chain, the mitochondrial glycerol-3-phosphate oxidase system is used to re-oxidize NADH produced during glycolysis. Specifically, this system oxidizes glycerol-3-phosphate (G3P) using an electron transport system in the inner mitochondrial membrane consisting of G3P dehydrogenase, ubiquinone, and TAO. Thus, aerobic respiration leads to the net production of 2 moles of ATP and 2 moles of pyruvate per glucose molecule.



Under anaerobic conditions, or in the presence of a TAO inhibitor, G3P accumulates inside the glycosome, and it is disposed off by conversion to glycerol by a reverse action of glycerol kinase (GK). This leads to the net production of 1 mole of ATP and equimolar amounts of pyruvate and glycerol.

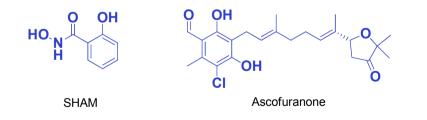
However, BSF trypanosomes do not survive for long time periods under anaerobic conditions: when glycerol accumulates in the cell, mass action induces glycerol kinase to convert glycerol to G3P and the glycolysis stops.

TAO is a validated target of trypanosomes:

- TAO is **essential** for viability of BSF trypanosomes
- TAO is expressed in all subspecies
- TAO is unique (absent in mammals)
- TAO is **sensitive to specific inhibitors** such as salicylhydroxamic acid (SHAM)

or ascofuranone

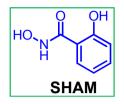
• TAO inhibitors are active in mouse models of *T. brucei* infection (e.g. SHAM, ascofuranone)

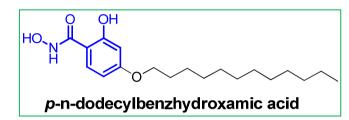


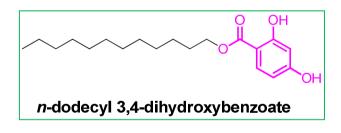


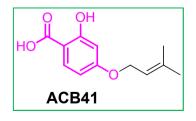


Examples of early TAO inhibitors active against T. brucei









- $K_i = 21 \, \mu M$
- $EC_{50} = 39 \,\mu M$
- Trypanocidal without glycerol
- Glycerol needed to see a therapeutic effect in vivo (Clarkson et al. *Mol. Biochem. Parasitol.* **1981**, 3, 271-291)

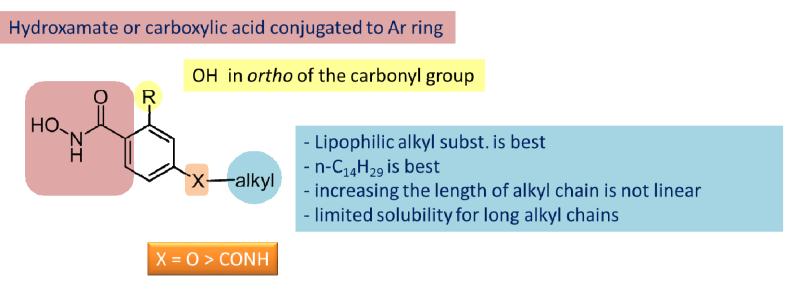
• $K_i = 1.1 \, \mu M$

- $EC_{50} = 1.5 \ \mu M$ (with glycerol)
- Trypanocidal in vitro when combined with glycerol
- Inactive in vivo → poor water solubility
 (Grady et al. *Mol. Biochem. Parasitol.* **1986**, *19*, 231-240)
- $IC_{90} = 0.9 \ \mu M$
- MIC = 1 -10 μM (10 mM glycerol)
- Reduces parasitaemia in mice when combined with glycerol (Grady et al. *Mol. Biochem. Parasitol.* **1986**, 21, 55-63)
- K_i = 5 μM
- $EC_{50} = 16.5 \ \mu M$
- Trypanocidal without glycerol (Ott et al. *Acta Trop.* **2006**, 172-184)





TAO inhibition: SAR of benzhydroxamic and benzoic acids reported in the literature

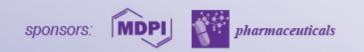


Grady et al. *Mol. Biochem. Parasitol.* **1986**, *19*, 231-240 Grady et al. *Mol. Biochem. Parasitol.* **1986**, *21*, 55-63

Drawbacks of these inhibitors:

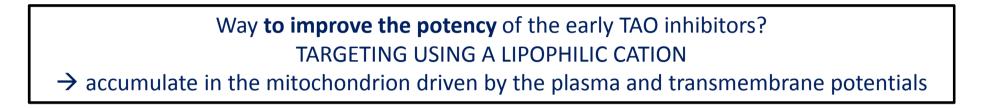
- Low potency of inhibition of TAO and T. brucei
- > Many compounds require glycerol (i.e. inhibit anaerobic pathway) to be trypanocidal
- Limited solubility

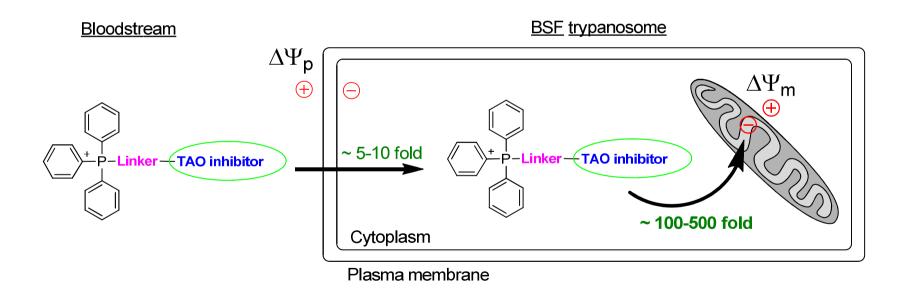




Results and discussion

Mitochondrion targeting with lipocations



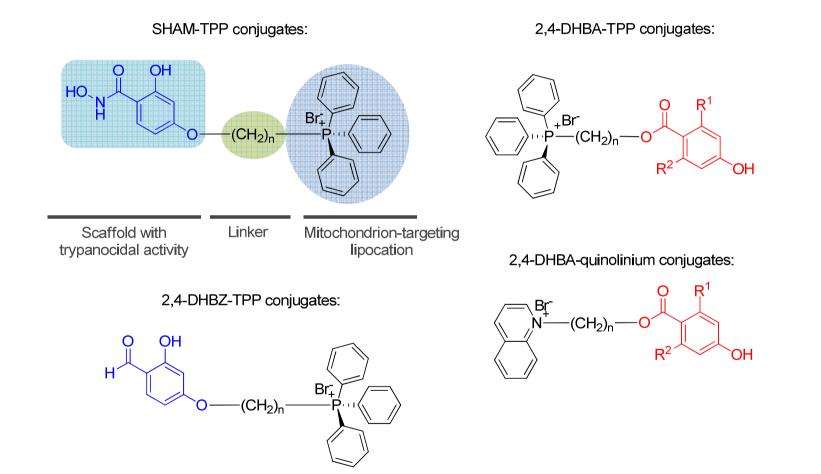






Results and discussion

1. Design and general structure of the SHAM, 2,4-dihydroxybenzaldehyde (DHBZ), and 2,4-dihydroxybenzoic acid (DHBA) conjugates



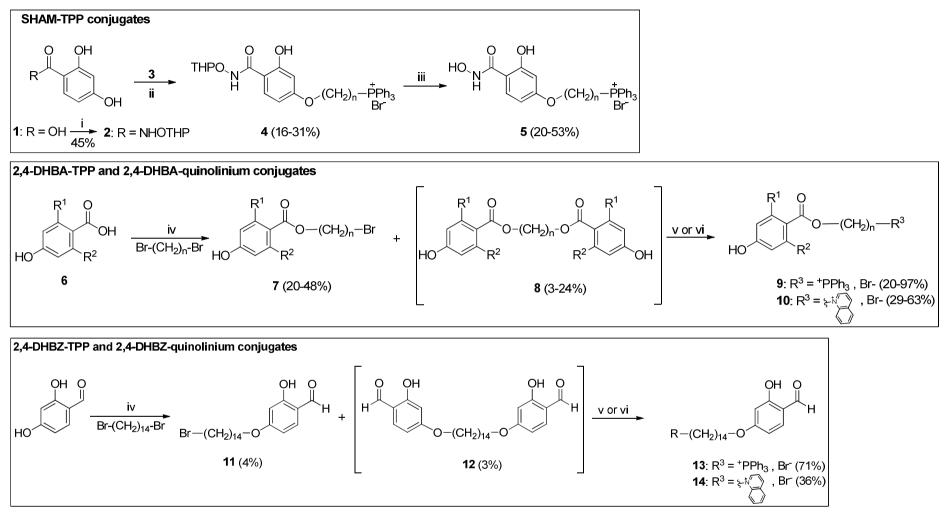


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2. Synthesis of the inhibitors



Reagents and conditions: (i) THPONH₂, EDC, NMM, HOBt, DMF, MWI, 120 $^{\circ}$ C, 30 min; (ii) Br-(CH₂)_n-PPh₃⁺Br⁻(**3**), NaHCO₃, NaI, CH₃CN, 65 $^{\circ}$ C, 3 days; (iii) TsOH (cat.), MeOH, rt; (iv) NaHCO₃, CH₃CN or DMF, Δ ; (v) Ph₃P, CH₃CN, 80 $^{\circ}$ C, 10 days; (vi) quinoline, CH₃CN, 80 $^{\circ}$ C, 10 days.





3. Inhibition of recombinant TAO enzyme

Compound series (conjugates)	Structure	<i>rT</i> ΑΟ IC ₅₀ (μΜ)
5 (SHAM-TPP)	HO N H O $(CH_2)_n$ $-PPh_3$ Br	> 5
7	$R^1 O (CH_2)_n - Br$	0.007 – 0.45
9 (2,4-DHBA-TPP)	$HO \qquad R^{1} O \qquad Br \\ O - (CH_{2})_{n} - PPh_{3}$	0.030 - 1.46
10 (2,4-DHBA-quinolinium)	$HO \xrightarrow{R^1 O (CH_2)_n - \overset{+}{N}}$	0.030 - 1.36
11	H O OH H O O OH O O OH O OH O OH O OH O	0.073
13 (2,4-DHBZ-TPP)	H H O-(CH ₂) ₁₄ -PPh ₃	0.22
14 (2,4-DHBZ-quinolinium)	$H \xrightarrow{O OH} Br_{+}$	1.23
SHAM Ascofuranone		5.93 0.002

In general, the addition of a mitochondrion-targeting lipocation barely affected the inhibitory potency against rTAO, showing that the lipocation does not participate in the interaction with the binding pocket (or, at the very least, does not interfere with binding to TAO) when a C14 linker is used.



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Compound series (conjugates)	Structure	7. b. brucei s427 (WT) EC ₅₀ (μM) Selectivity index (SI)	Τ. congolense EC ₅₀ (μΜ) (SI)	<mark>Cytotoxicity</mark> Human cells CC ₅₀ (μM)
5 (SHAM-TPP)	HO OH HO N H O $(CH_2)_n - PPh_3$ Br	0.14 – 0.4 (SI > 1000)	27 – 46 (SI > 8)	>200
7	$HO \xrightarrow{R^1 O} O \xrightarrow{(CH_2)_n} Br$	14.4 – 45.7	>50	>200
9 (2,4-DHBA-TPP)	$HO \xrightarrow{R^1 O G} O \xrightarrow{Br} O \xrightarrow{PPh_3} O \xrightarrow{HO G} O \xrightarrow{R^2} O \xrightarrow{HO G} O \xrightarrow{R^2} O $	0.0012 –0.073 (SI > 500)	<mark>0.03 – 3.9</mark> (SI: 5 to > 3000)	>200
10 (2,4-DHBA-quinolinium)	$HO \xrightarrow{R^1 O (CH_2)_n - \overset{H}{N}}$	0.14 – 0.33 (SI > 600)	3.0 – 7.3 (SI > 34)	>200
11	H O OH O	17.6	42.6	>200
13 (2,4-DHBZ-TPP)	$H \xrightarrow{O OH} Br \xrightarrow{+} Br O-(CH_2)_{14}$ -PPh3	0.133 (SI > 1500)	0.27 (SI > 740)	>200
14 (2,4-DHBZ-quinolinium)	$H \xrightarrow{O OH} Br_{+}$	1.75 (SI > 114)	2.1 (SI > 95)	>200
SHAM Pentamidine Diminazene Phenylarsine oxide		38.7 0.003 0.065 0.001	0.20	0.29

4. In vitro activity against T. b. brucei and T. congolense



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5. Mechanism of action studies

Most of these TAO inhibitors were significantly more effective in the presence of 5 mM glycerol, and against aquaglyceroporin-null trypanosomes which have glycerol efflux defects, consistent with TAO being the principal target of these inhibitors in the parasite cell.

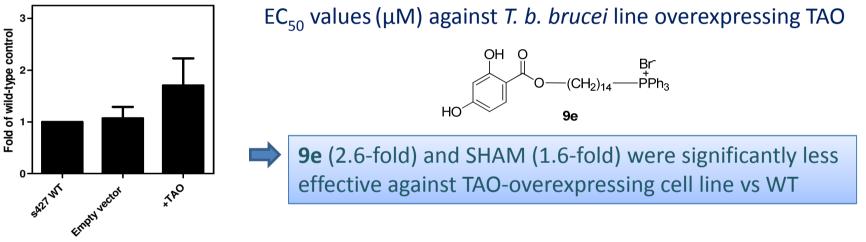


Figure 2. Expression of TAO in *T. b. brucei* trypomastigotes. Relative levels of TAO expression were determined by qPCR in wild-type Lister 427, in the same cell line transfected with the 'empty vector' pHD1336 (no insert) and with the TAO open reading frame in pHD1336. Average and SEM of 3 determinations.

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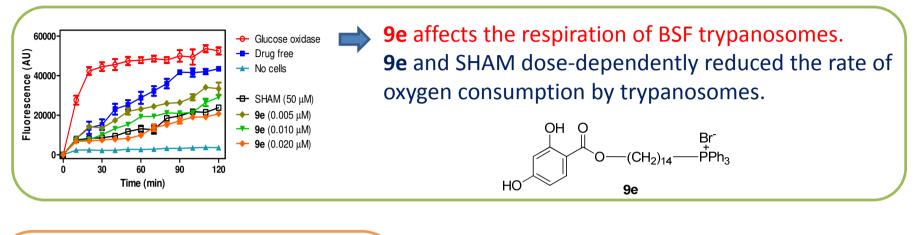
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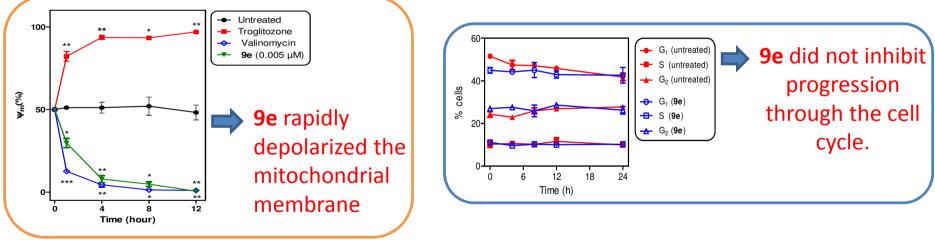
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5. Mechanism of action studies

Effect of **9e** on **oxygen consumption**, **mitochondrial membrane potential** and **cell cycle** in *T. b. brucei* WT





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Conclusions

We have successfully developed a class of potent and selective new hits active against human (*T. brucei* spp.) and veterinary (*T. congolense*) African trypanosomes, and established their probable mode of action via TAO inhibition. This was accomplished by efficiently targeting the compounds to the trypanosome's mitochondrion, thereby increasing the potency of the original small molecule inhibitors against *T. brucei* by up to 3 orders of magnitude.

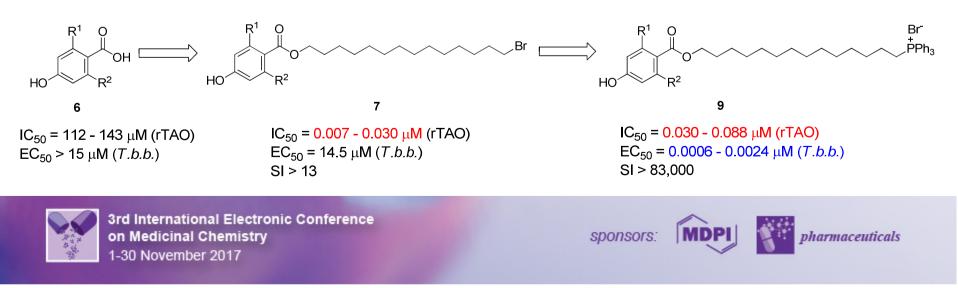
□ Attaching a Lipophilic Cation to a TAO inhibitor using a 14-methylene linker:

- ✓ nanomolar trypanocidal activity
- \checkmark not detrimental to inhibition of TAO

□ The 2,4-DHBA-TPP conjugates are the most potent and selective against *T. brucei*

 \Box Metabolic stability in serum depends on R¹ and R² \rightarrow candidates for in vivo studies have

been selected.



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