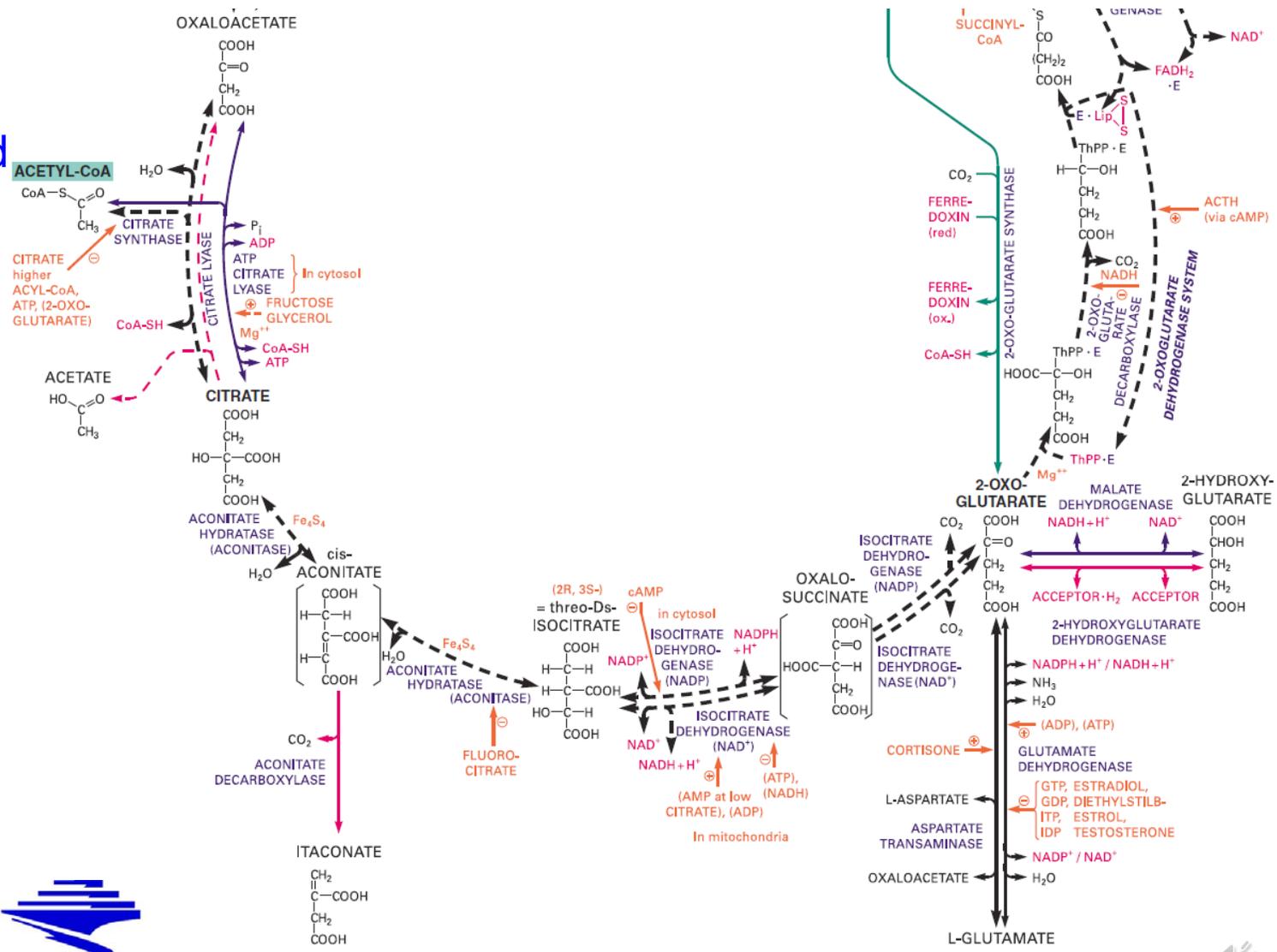


Tracing compartment-specific redox pathways using stable isotopes and mass spectrometry



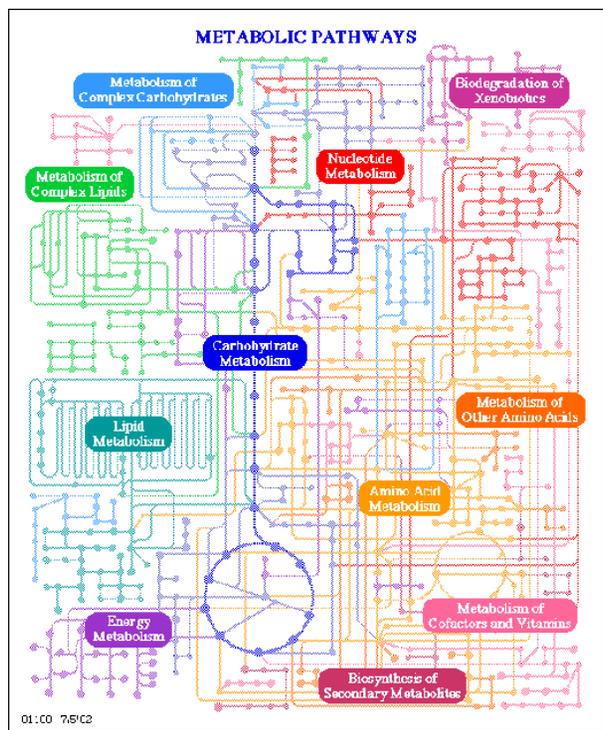
Christian Metallo
IECM 2017

Department of Bioengineering
Moore's Cancer Center
UCSD Diabetes Research Center

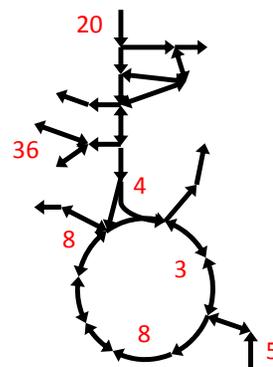


Michal and Schomburg Biochemical Pathways 2nd Ed. 2012

The challenge for biologists, biochemists, and engineers: Translate biochemistry to **metabolic fluxes**



<http://www.genome.jp/kegg/>



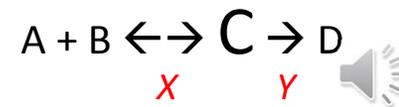
$$Flux = \frac{mol}{time \cdot cell/prot}$$

$$v = \frac{V_{max}[S]}{K_M + [S]}$$

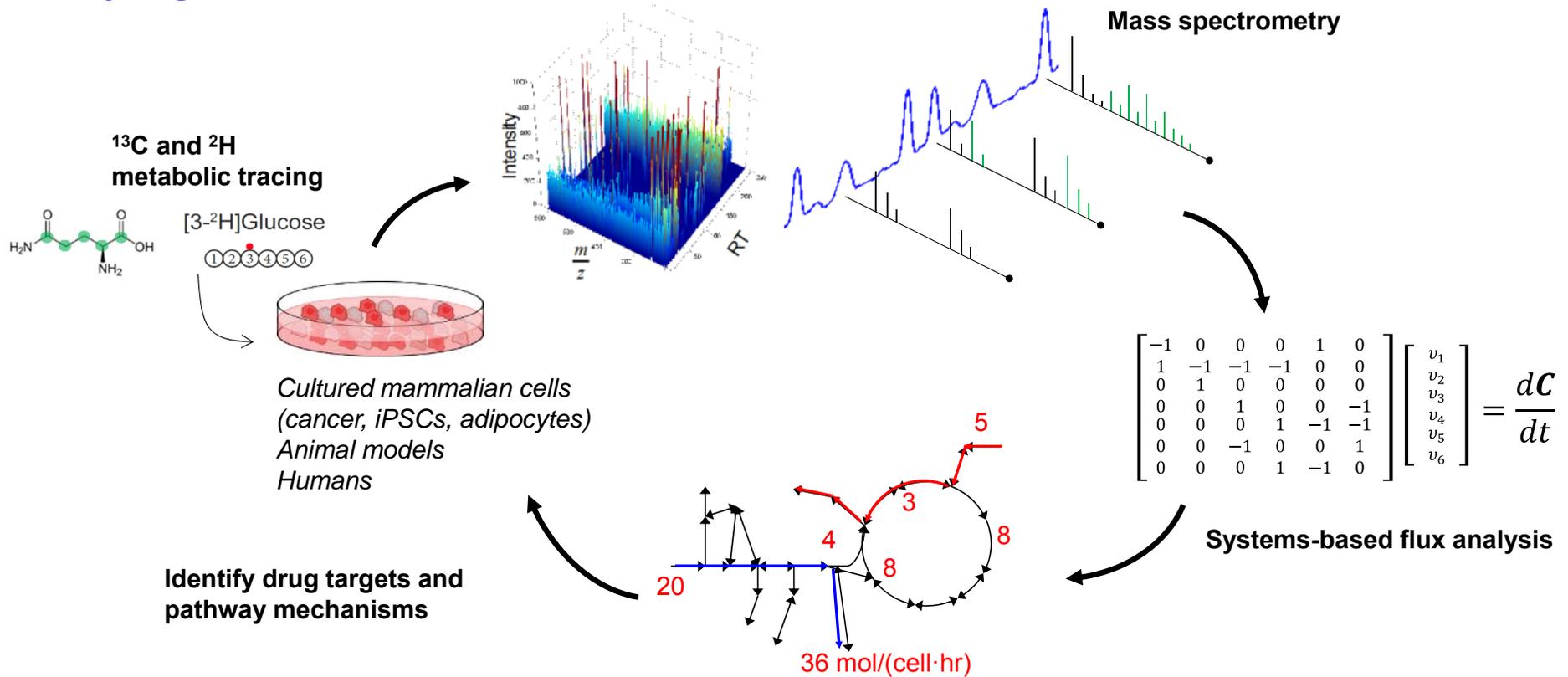
- **Fluxes** describe the ultimate function of metabolic enzymes
- This is where **metabolomics/analytical chemistry** meets **cell biology**
- Metabolite level measurements only get you so far

Use isotopic tracers

Analyze data as a system → **MODELING!!!**



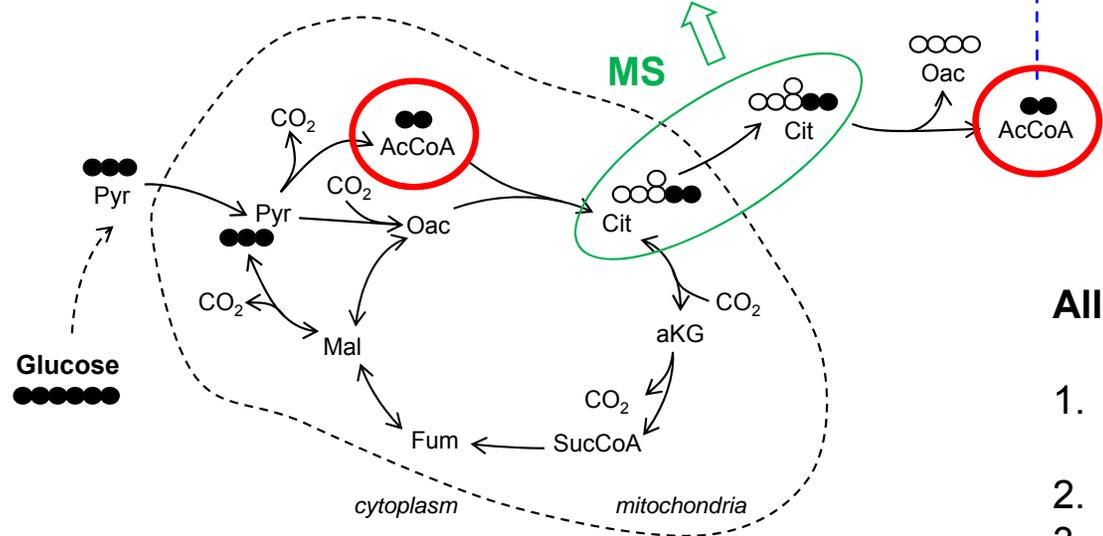
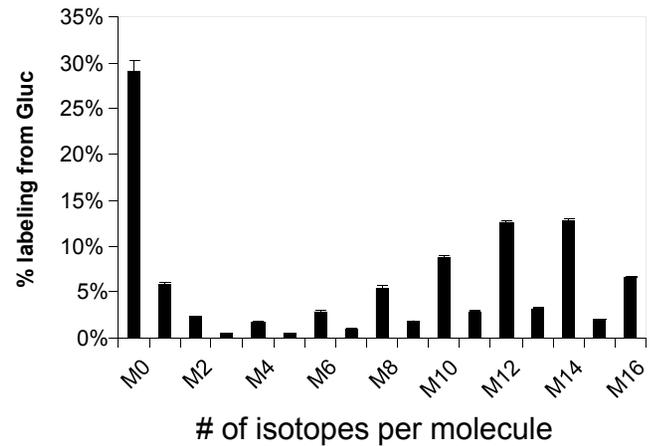
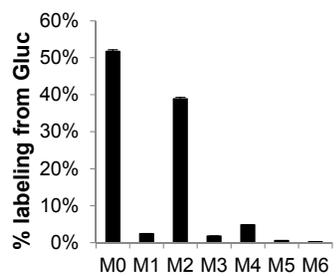
Studying metabolism for flux sake



1. Targeting metabolism in cancer (Grassian et al. *Canc Res* 2014; Svensson et al. *Nat Med* 2016; Parker et al. *Met Eng* 2017)
2. **Cellular compartmentalization and redox metabolism** (Lewis et al. *Mol Cell* 2014; Vacanti et al. *Mol Cell* 2014)
3. Metabolic changes during iPSC growth/differentiation (Badur et al. *Biotech J.* 2015; Zhang et al. *Cell Rep* 2016)
4. Regulation of macrophage metabolism (Cordes et al. *JBC* 2016)
5. Understanding adipose tissue metabolism and physiology in the context of T2DM (Green et al. *Nat Chem Bio* 2016)



Our approach to study cell physiology and metabolism



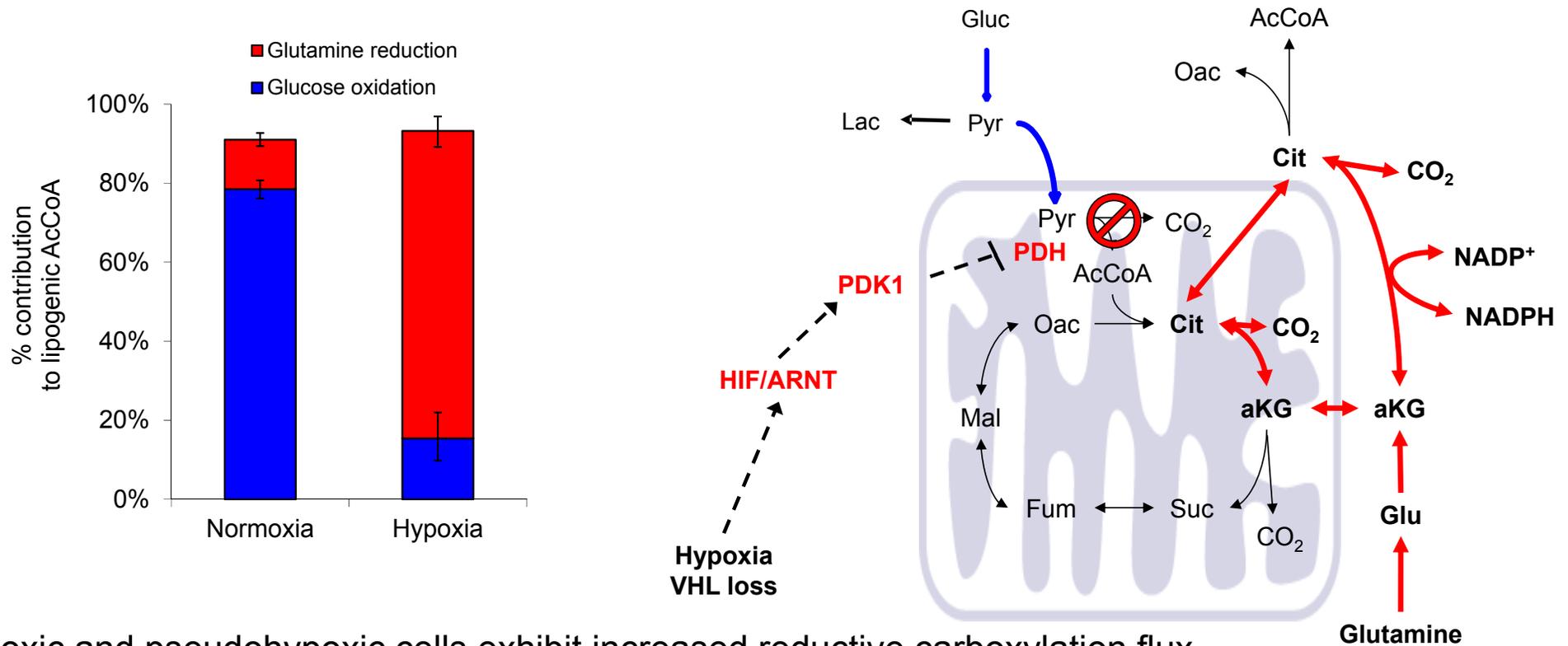
○ = Carbon atom
● = ¹³C atom

Allows quantitation of:

1. Contribution of different substrates to AcCoA pools and mitochondrial metabolism
2. Fatty acid synthesis/de novo lipogenesis rates
3. Directionality of TCA metabolism
4. Intracellular fluxes with MFA modeling



Reprogramming of TCA metabolism under hypoxia



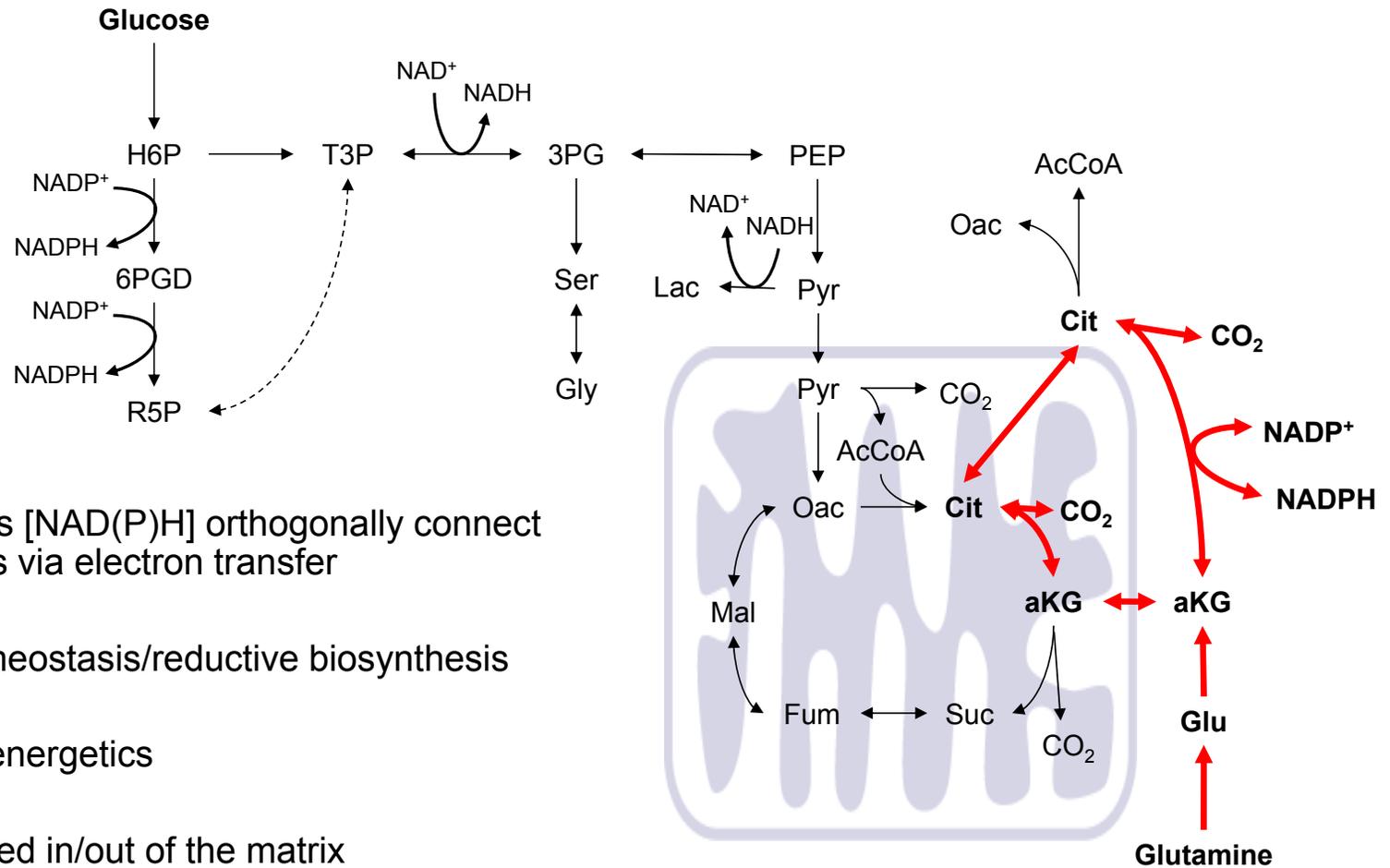
Hypoxic and pseudohypoxic cells exhibit increased reductive carboxylation flux

- Compartmentalization of metabolic processes is critical for cell function (but complicates analysis)
- Redox metabolism is perturbed by hypoxic stresses

Metallo et al. *Nature* (2012), Mullen et al. *Nature* (2012), Scott et al. *JBC* (2011), Wise et al. *PNAS* (2011)



Redox metabolism is highly compartmentalized



Pyridine nucleotides [NAD(P)H] orthogonally connect metabolic pathways via electron transfer

NADPH: redox homeostasis/reductive biosynthesis

NADH: cellular bioenergetics

Neither is transported in/out of the matrix



Eukaryotes are highly compartmentalized

^{13}C tracing and metabolomics **cannot** resolve compartment-specific metabolism

How are NADPH and NADH regenerated in the cytosol and mitochondria?

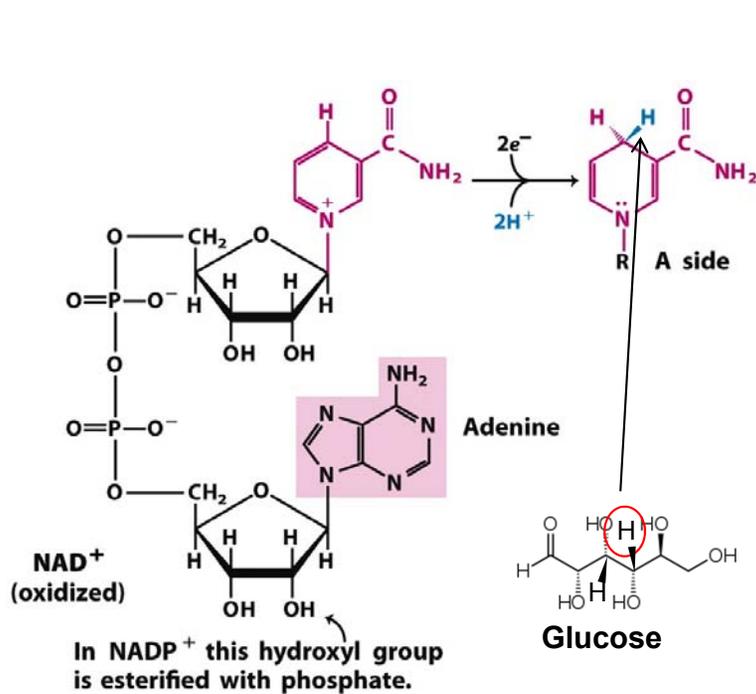
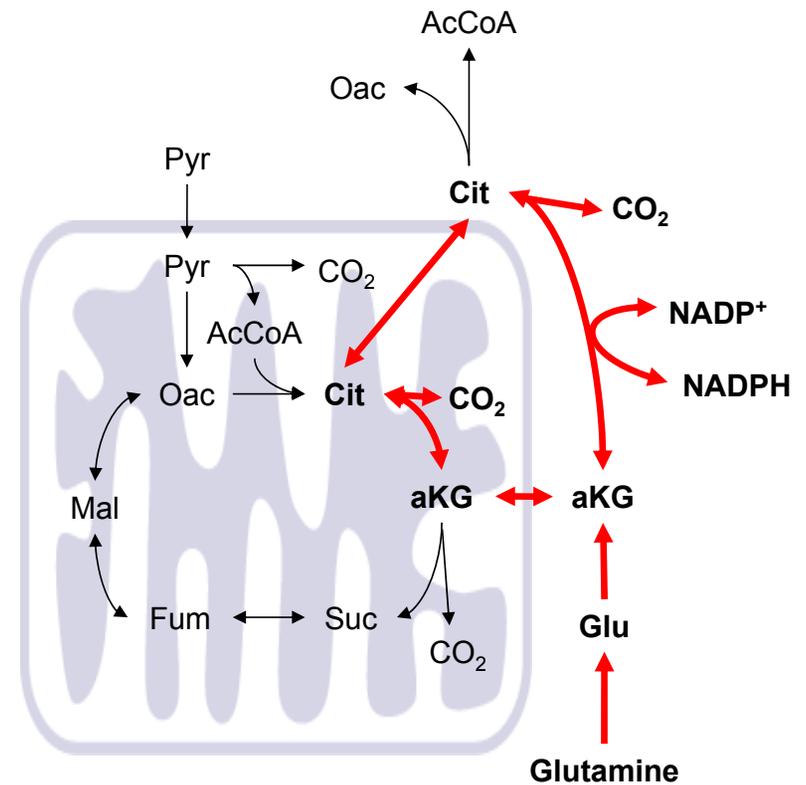
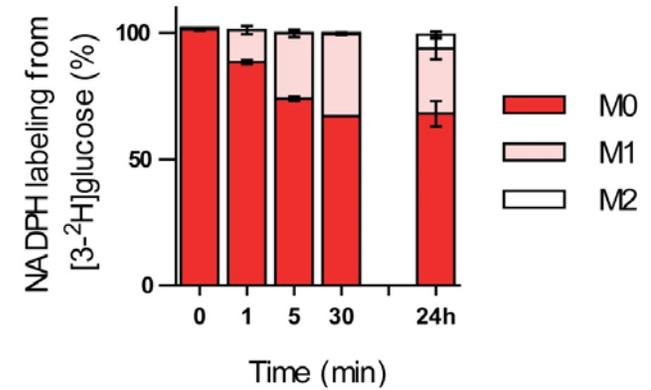
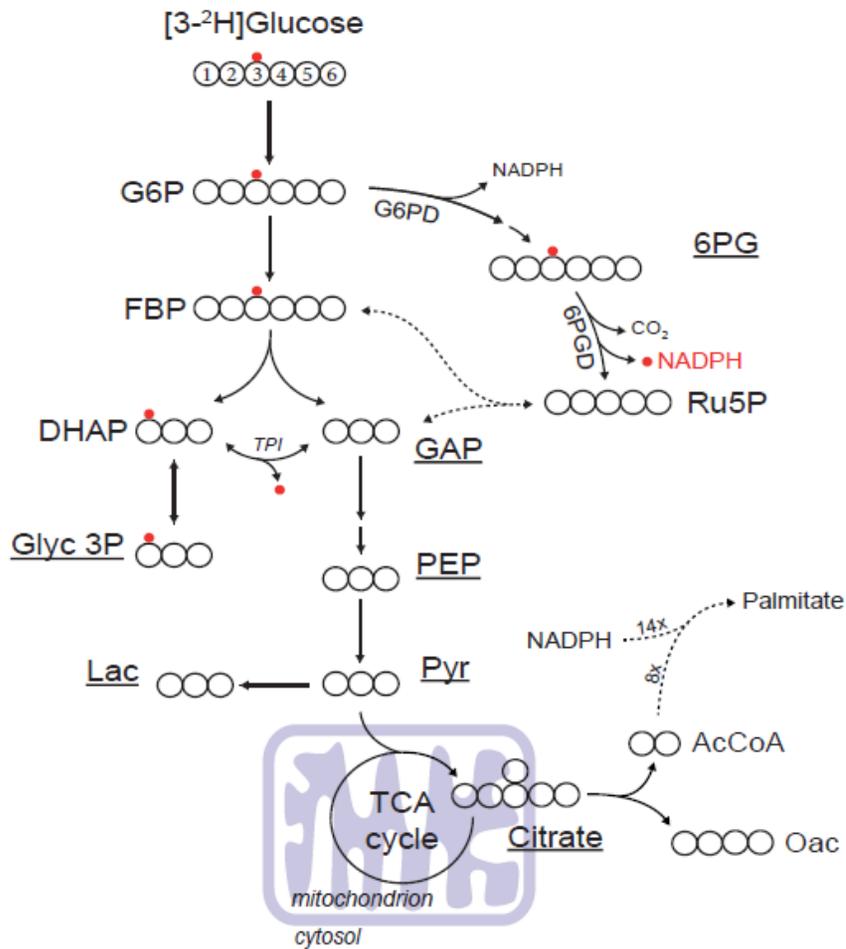


Figure 13-24a
Lehninger Principles of Biochemistry, Fifth Edition
© 2008 W.H. Freeman and Company

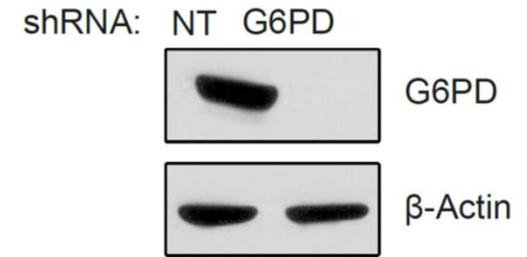
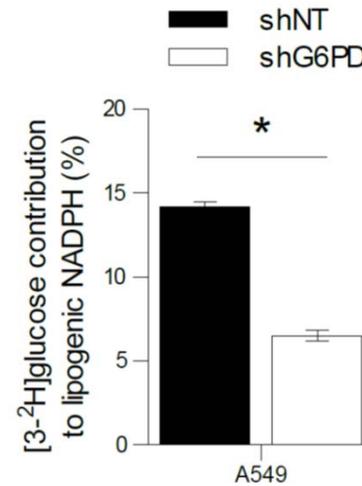
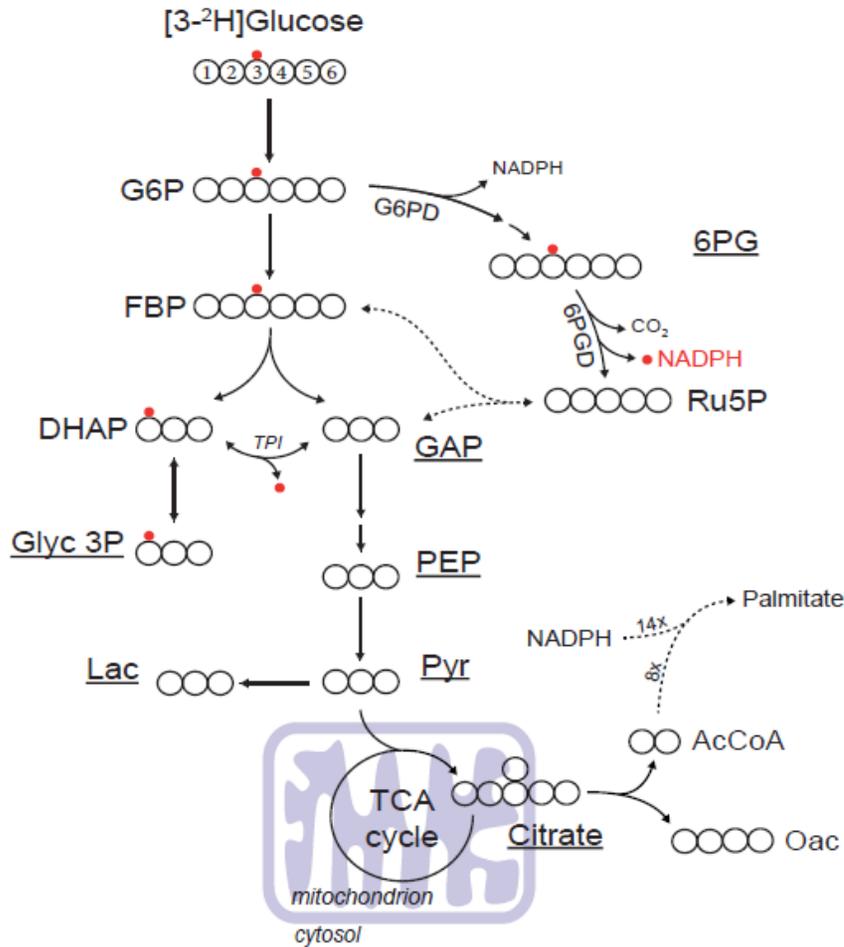


Tracing the oxidative PPP with [2H]glucose

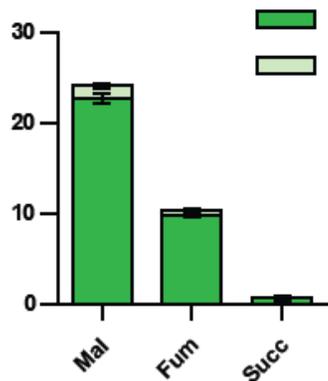
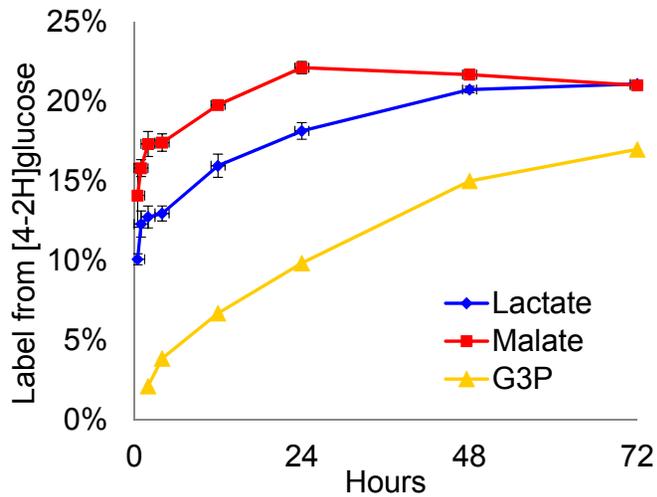


w/ Matt Vander Heiden (MIT)
Lewis et al. *Molecular Cell* 2014

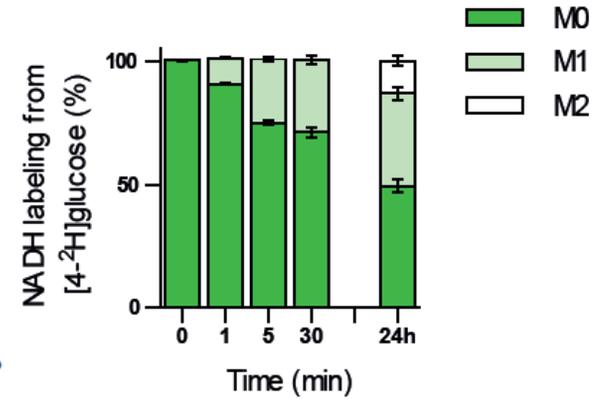
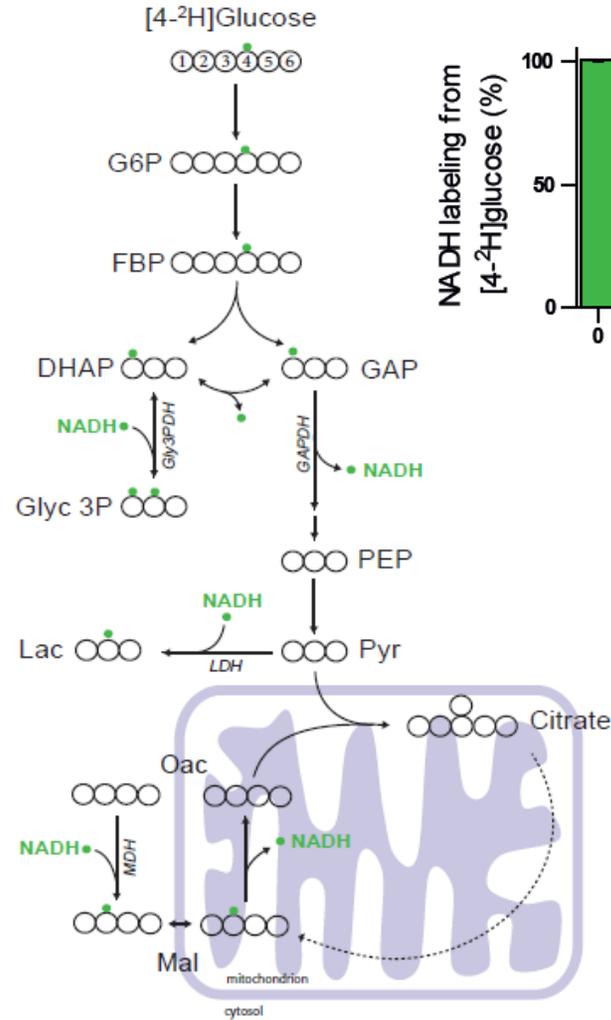
Contribution of the oxidative PPP to NADPH pools



NADH shuttles and mitochondrial metabolism regenerate NAD^+ for glycolysis



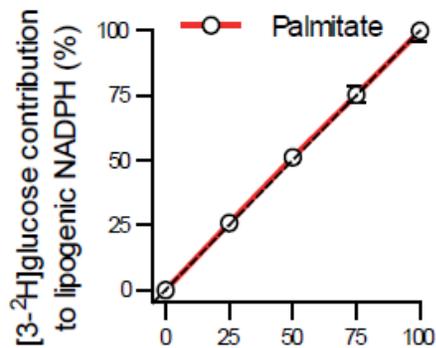
2H label enters TCA cycle via malate-aspartate shuttle



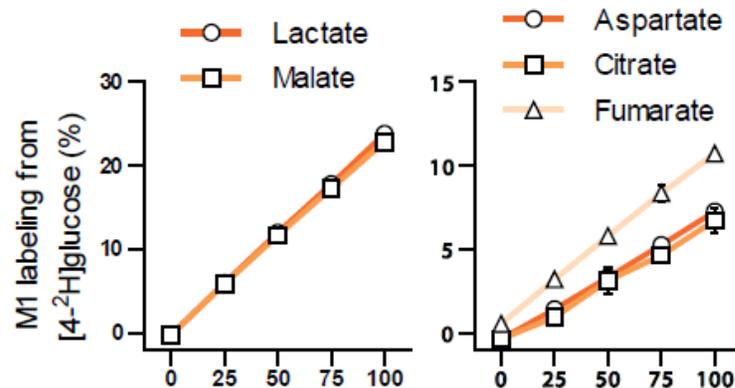
Do kinetic isotope effects affect results?

- Deuterium lowers rates in enzyme reactions (*in vitro*)
- Is this relevant to tracing through metabolic networks?
 - Allow “H” and “D” to compete by diluting
 - Compare labeling

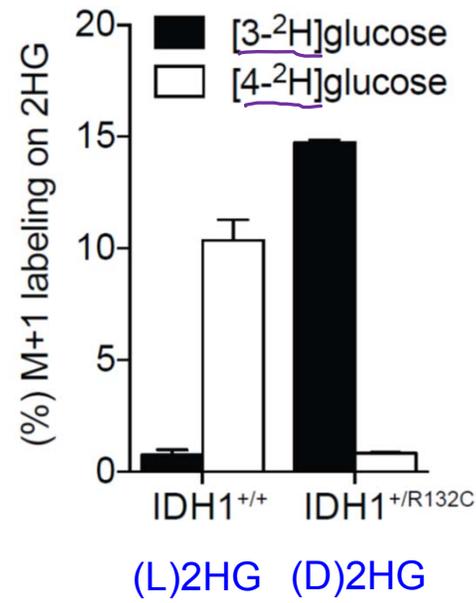
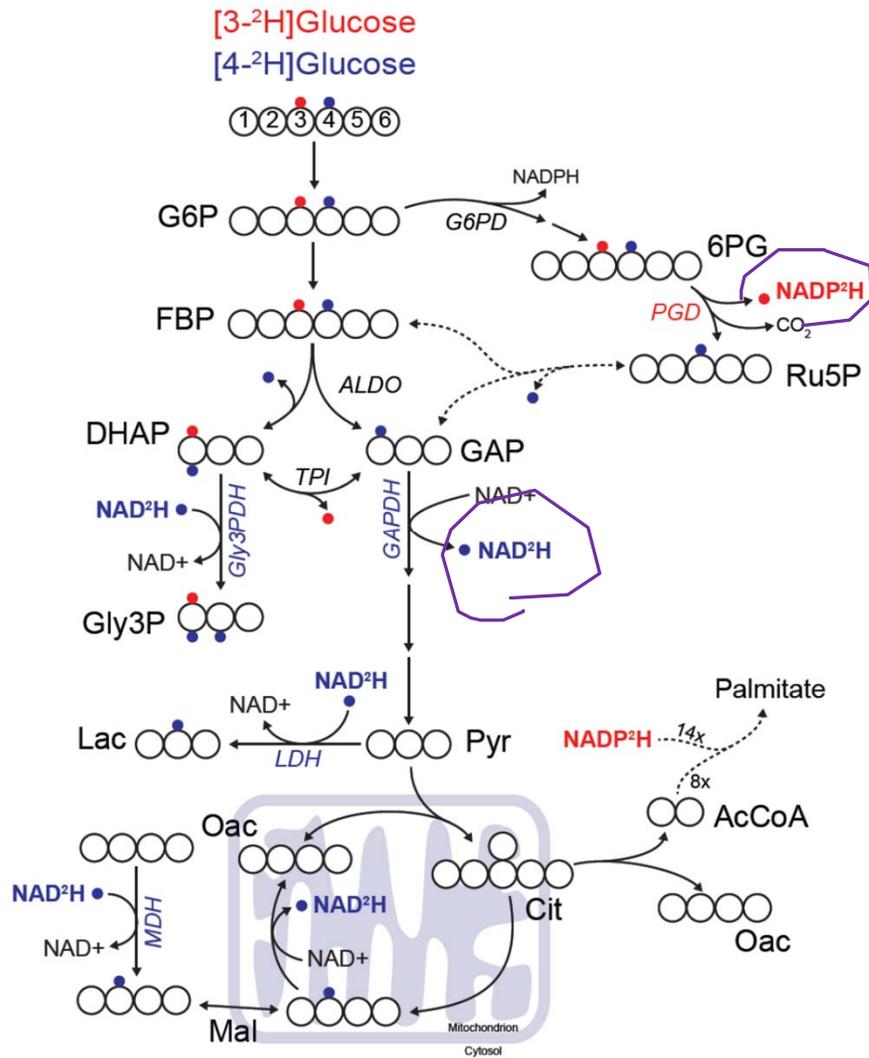
Cytosolic NADPH pathways



NADH metabolism



(L)2HG and (D)2HG have different origins and are labeled distinctly via 2H tracers



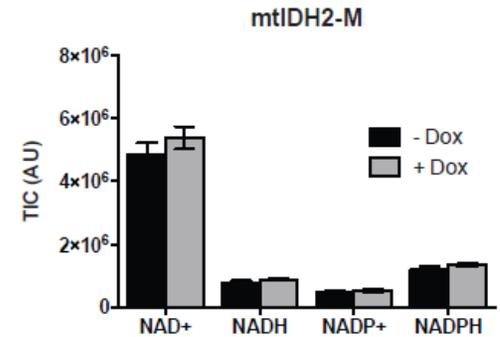
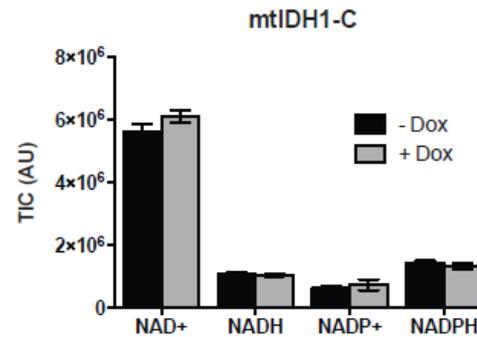
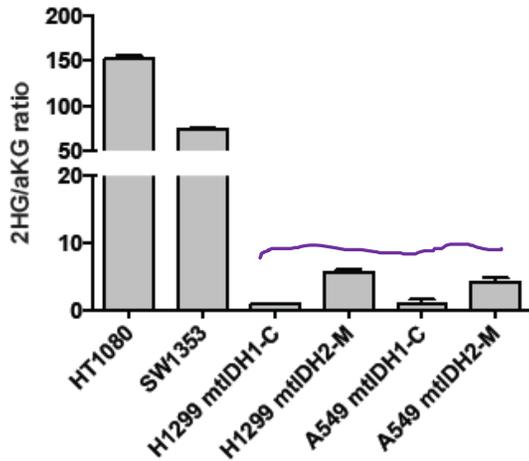
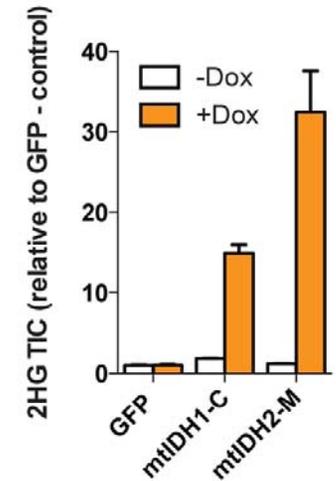
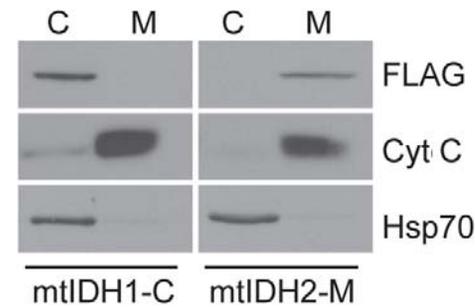
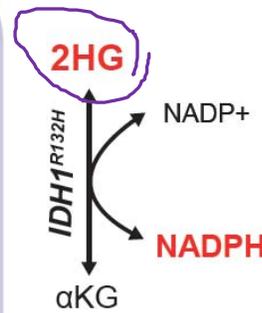
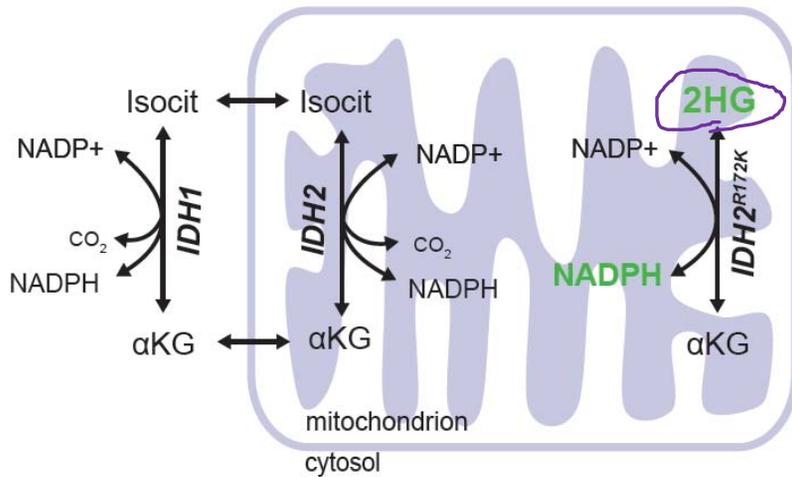
MDH and LDH generate (L)2HG from NADH

Oncogenic IDH1 generates 2HG from cytosolic NADPH

2HG is distinctly labeled by these tracers



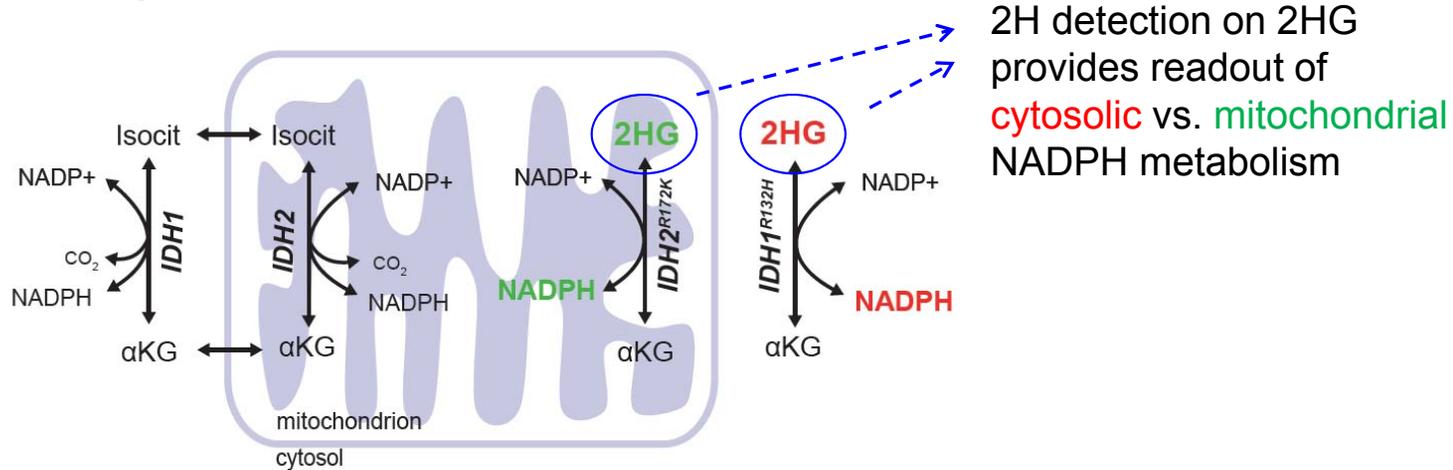
Using 2-HG production as a reporter of compartment-specific NADPH pools



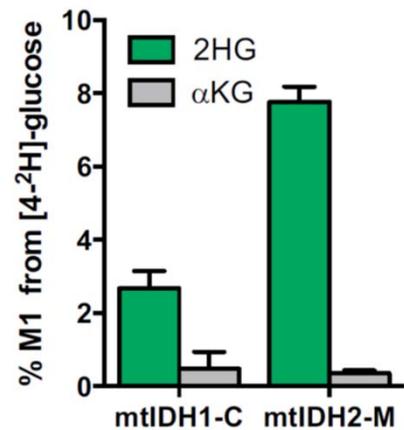
Minimal effect on central carbon metabolism
Grassian et al. *Cancer Research* (2014)



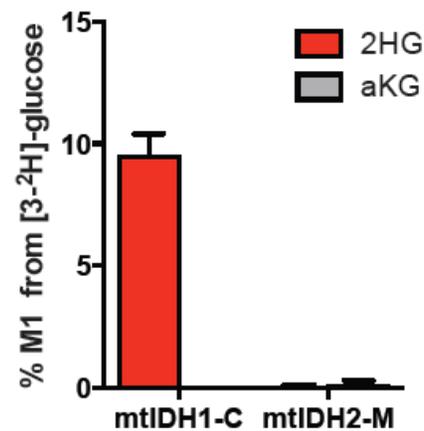
Using 2-HG production as a reporter of compartment-specific NADPH pools



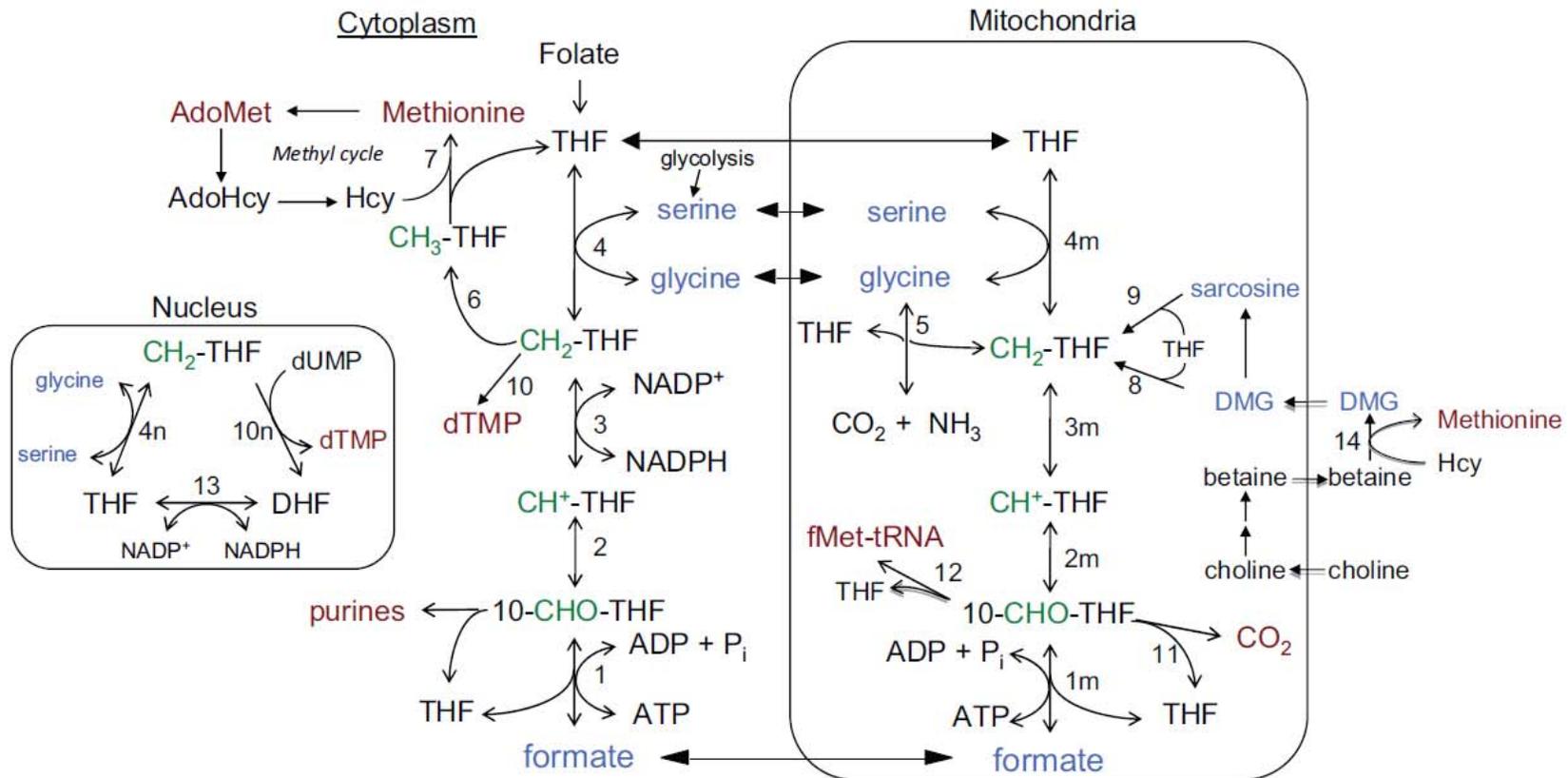
NADH trace (via glycolysis)



Cytosolic NADPH trace (via oxidative PPP)



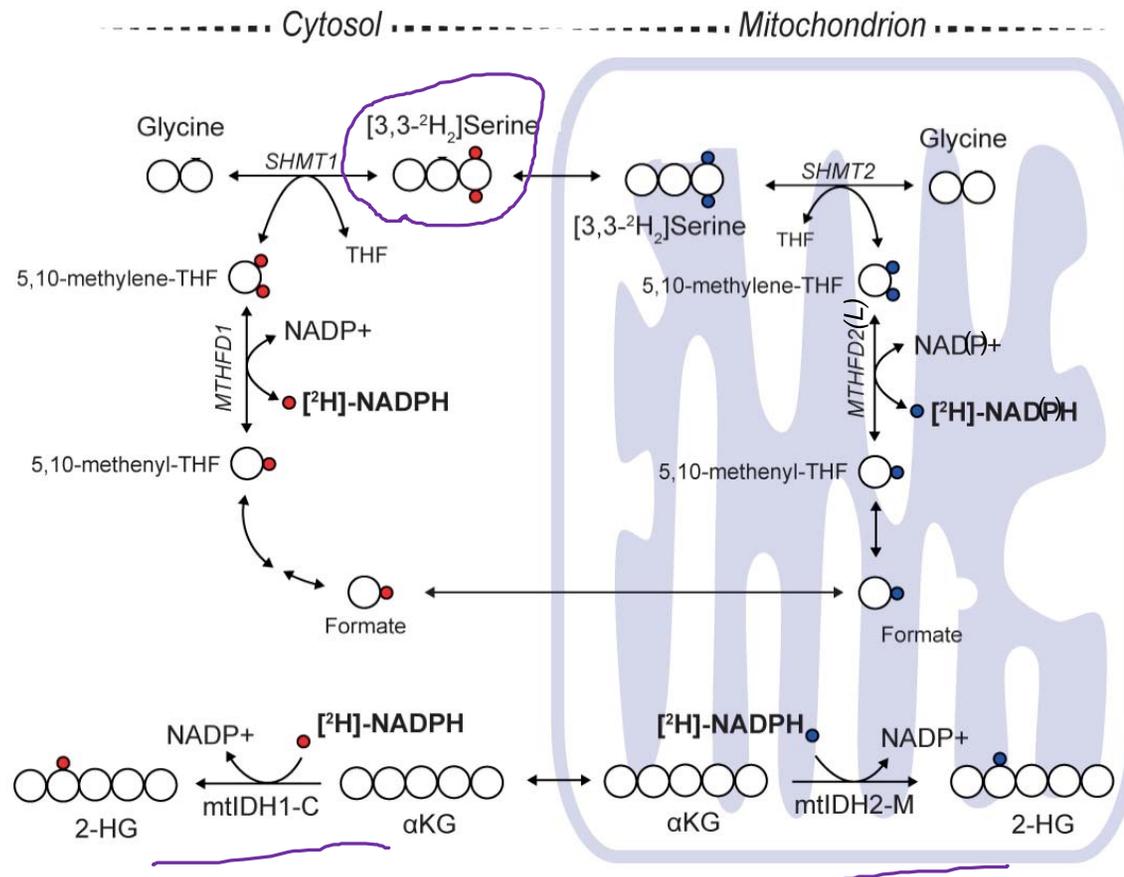
Can we use this reporter to annotate compartment-specific metabolic pathways? Folate-mediated one carbon metabolism



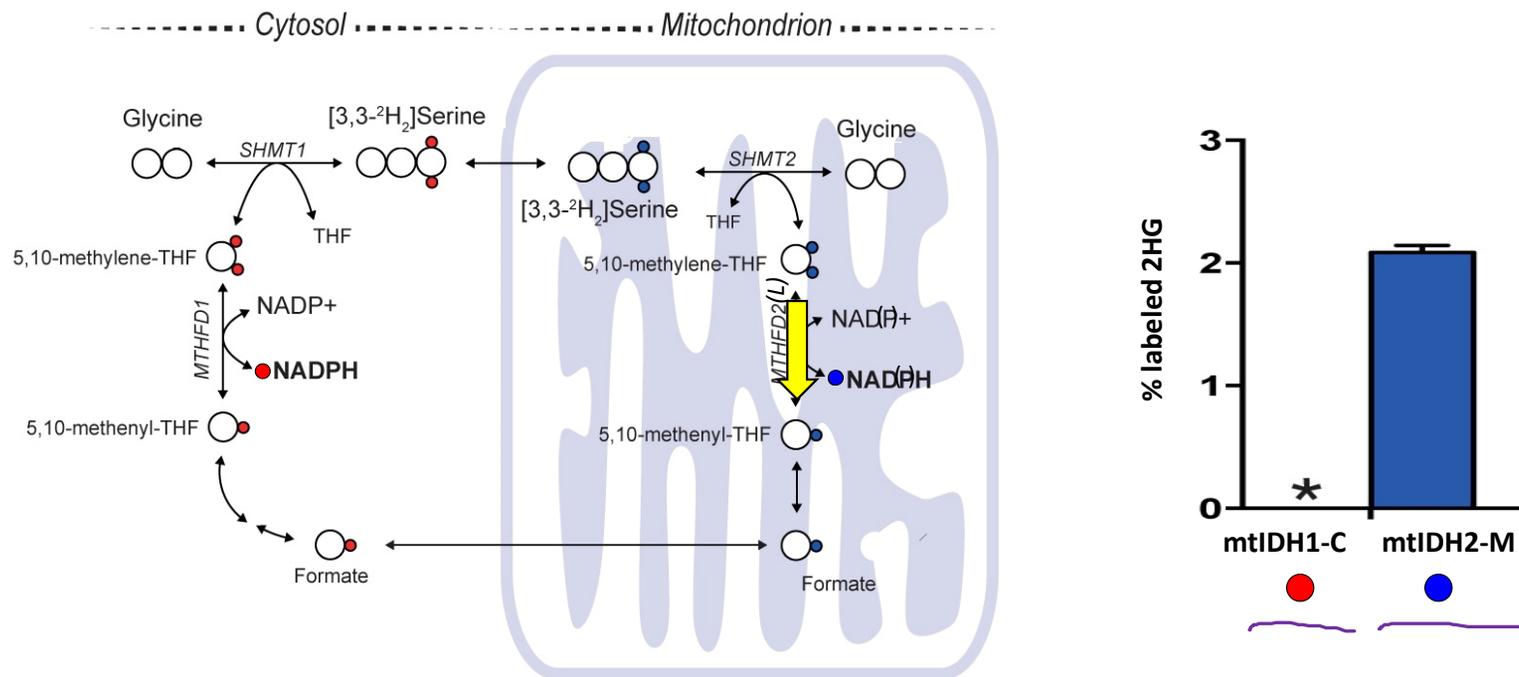
Tibbetts and Appling, *Ann. Rev. Nutr.* 2010



Can we use this reporter to annotate compartment-specific metabolic pathways? Folate-mediated one carbon metabolism



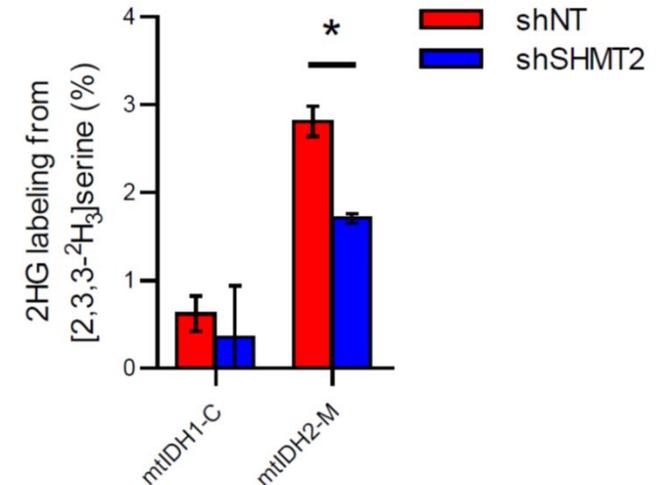
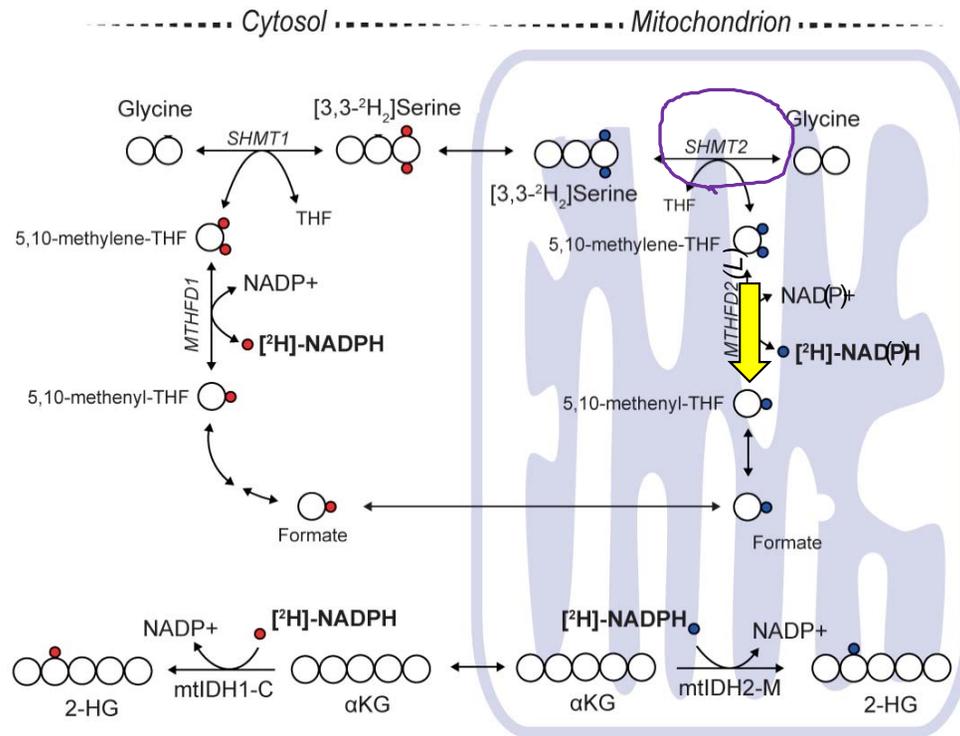
Discerning compartment-specific serine metabolism using cofactor tracing



NADPH produced from serine only observed in mitochondria



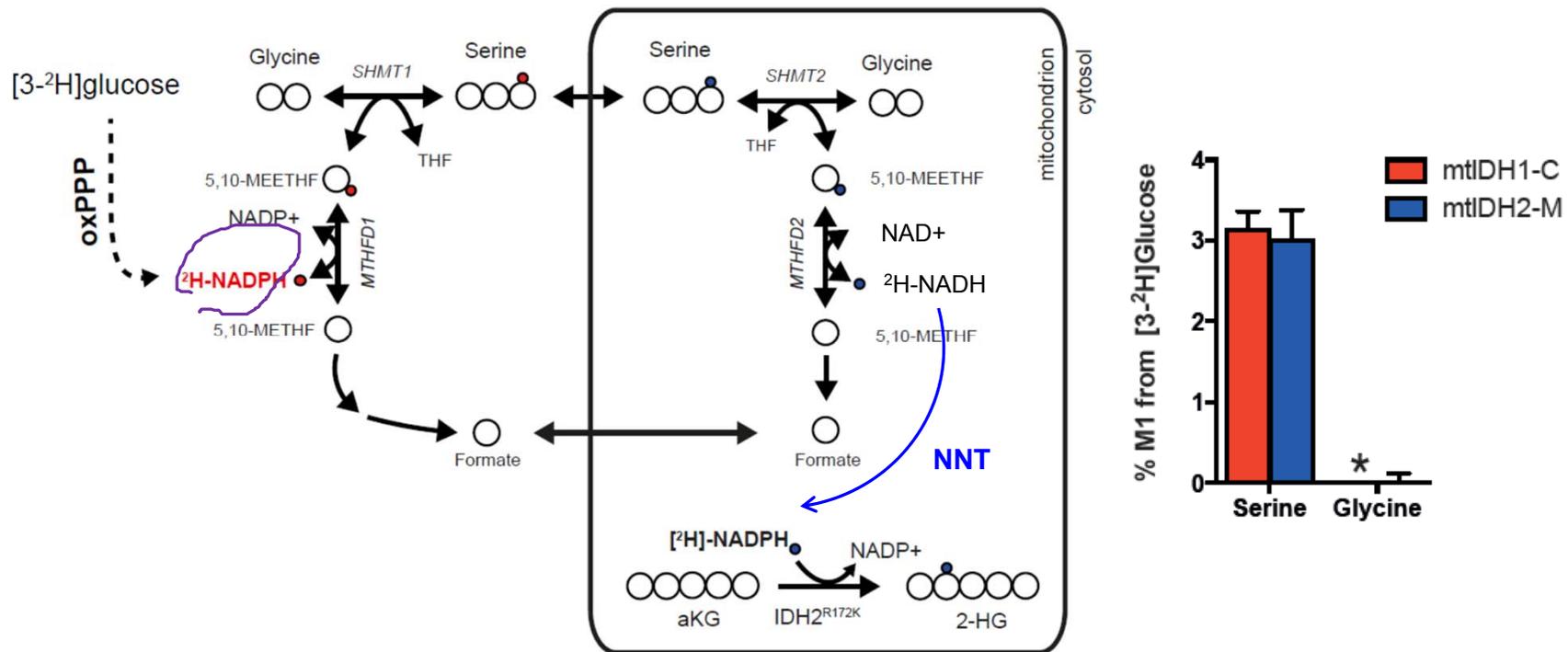
Discerning compartment-specific serine metabolism using cofactor tracing and mIDH reporters



Serine, glycine, and folate-mediated one carbon metabolism generate mitochondrial reducing equivalents



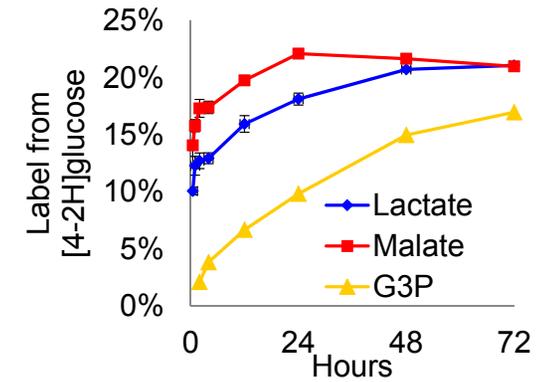
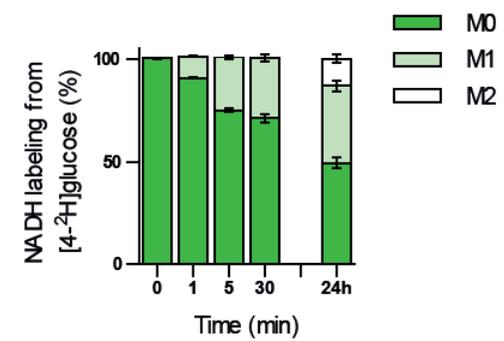
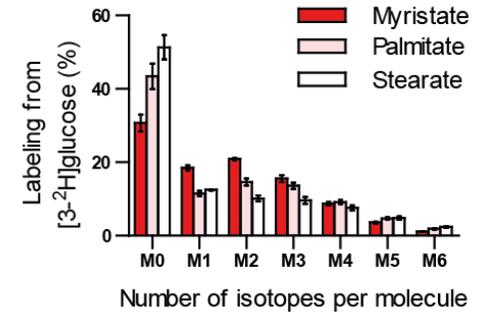
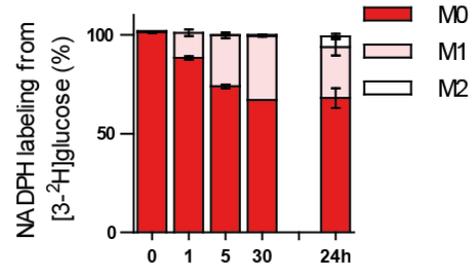
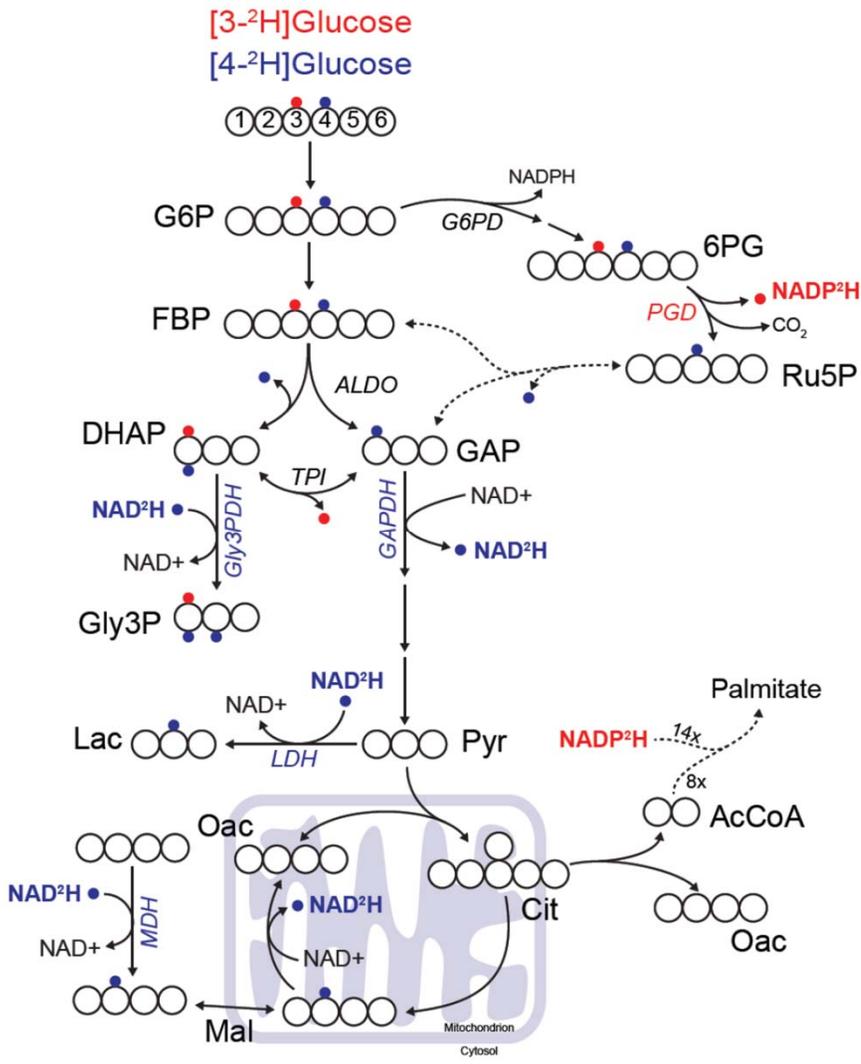
Discerning compartment-specific serine metabolism using cofactor tracing



Cytosolic reactions consume NADPH/produce serine
 NADPH from the oxidative PPP appears on serine

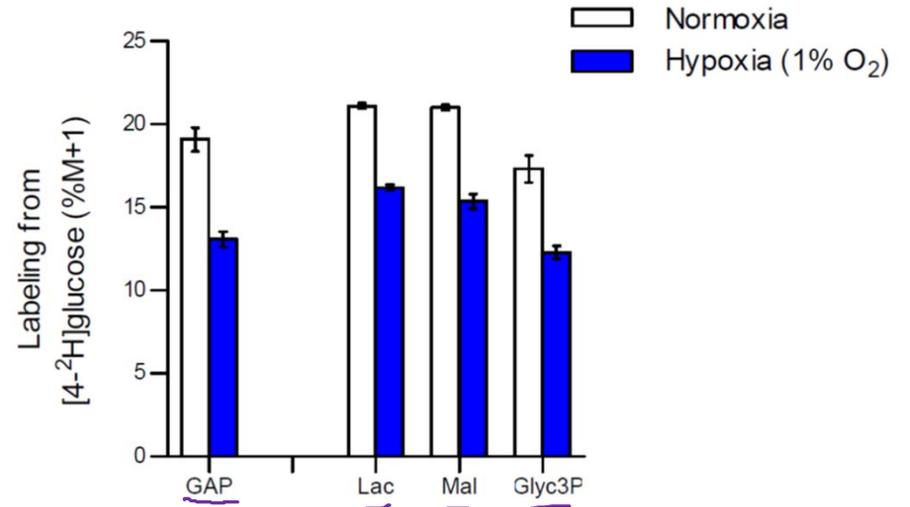
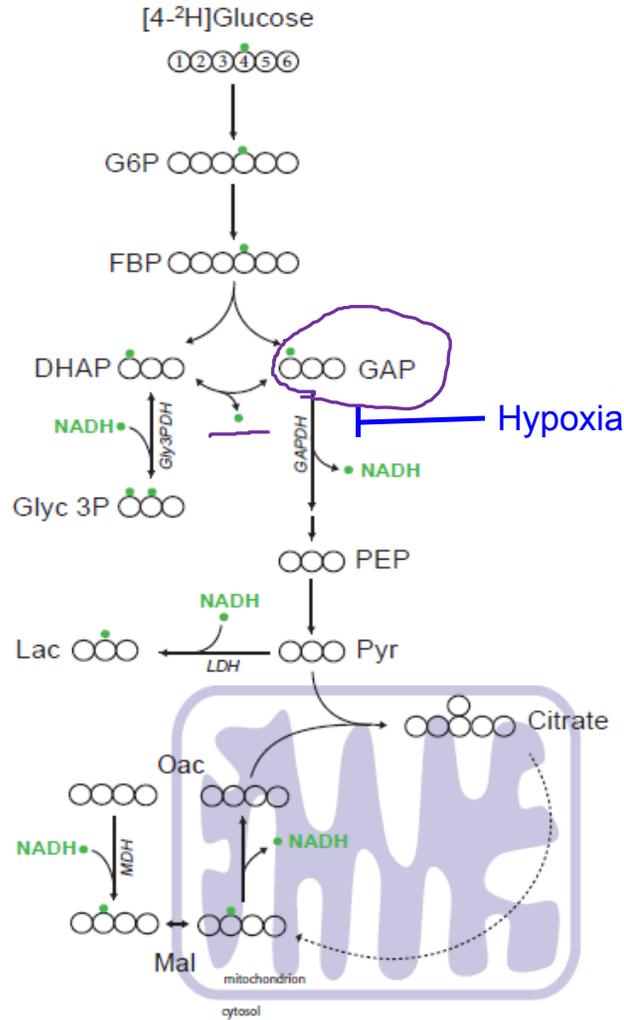


Resolving compartment-specific NADPH metabolism using 2H tracers and mutant IDH



- ^2H tracers allow for quantitation of NAD(P)H metabolism
- Oncogenic IDH1 and IDH2 used as reporters for compartment-specific NADPH labeling

How is NAD(P)H metabolism reprogrammed under hypoxia?

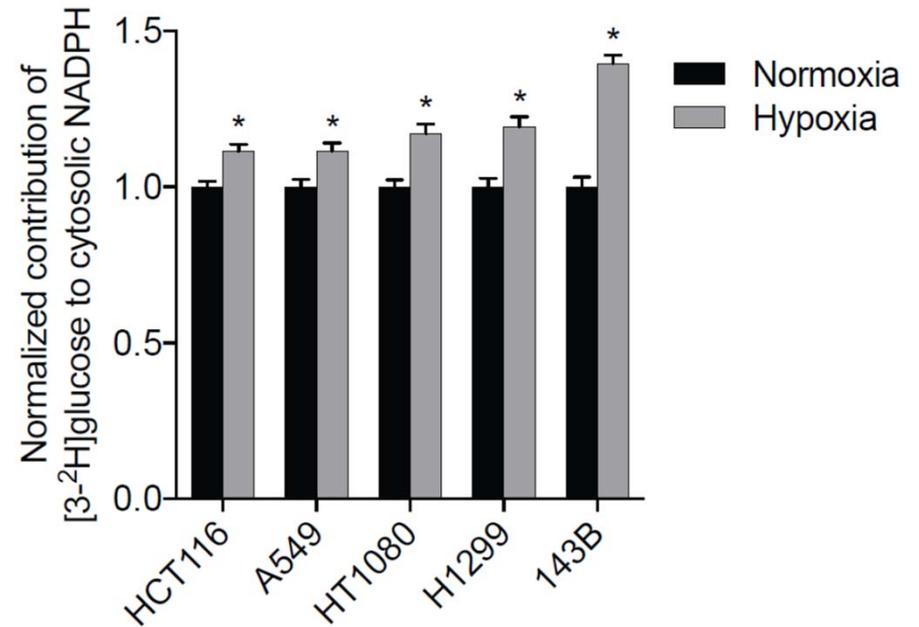
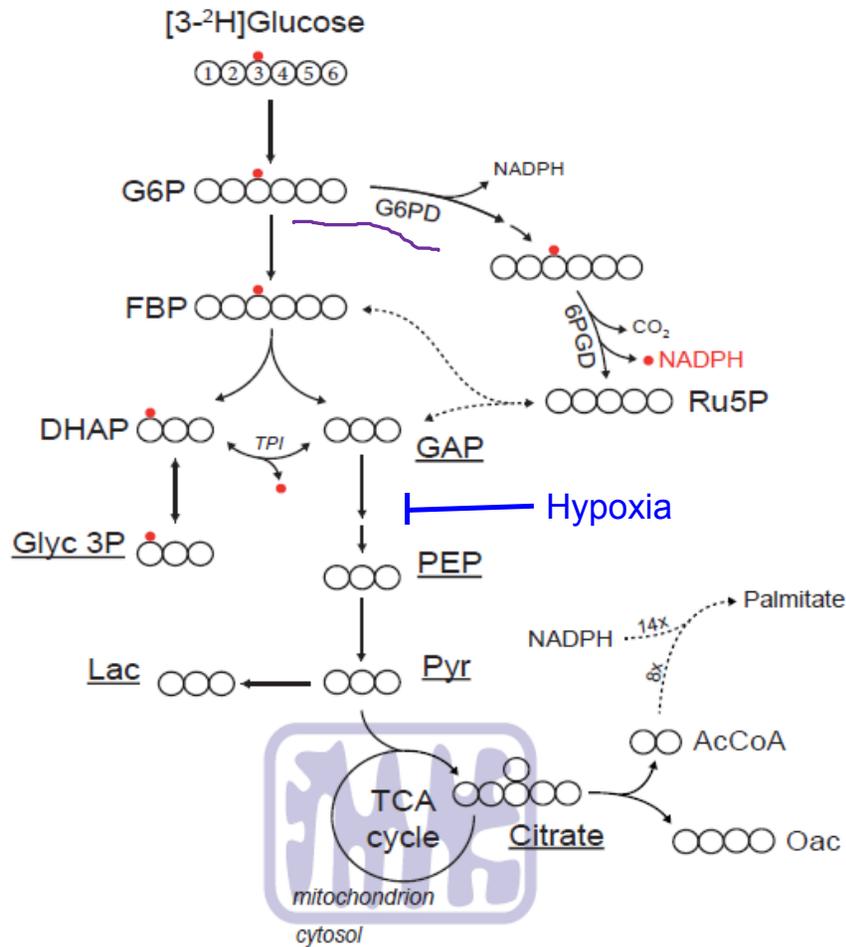


Oxidation of GAPDH under hypoxia leads to increased loss of isotope

Increased exchange flux at TPI/aldolase



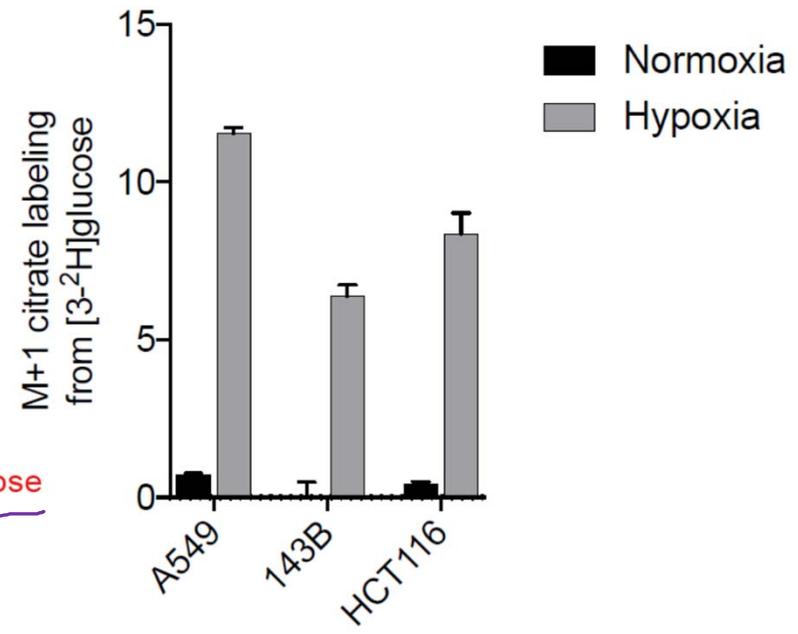
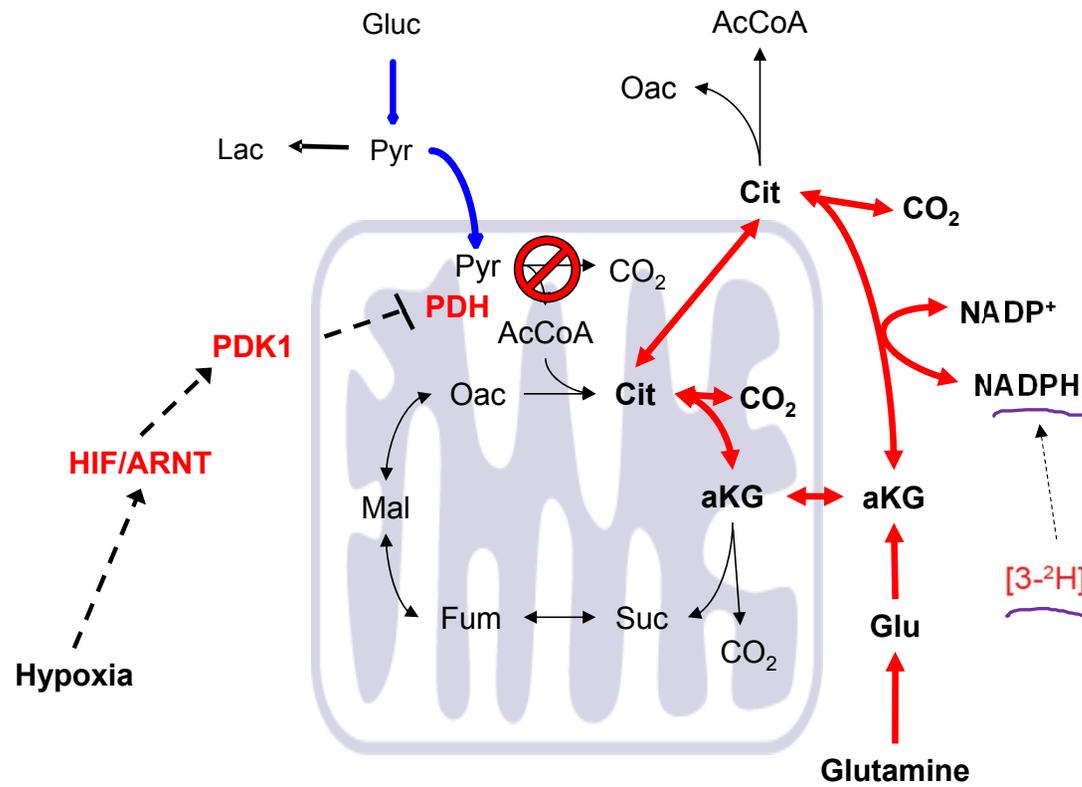
Hypoxia increases flux through the oxidative pentose phosphate pathway



GAPDH oxidation leads to increased (15-40%) oxidative PPP contribution to NADPH pools



Hypoxic induction of reductive carboxylation is mediated by cytosolic oxPPP flux and IDH1



Acknowledgements

Metallo Lab

Martina Wallace
Thekla Cordes
Le You

Mehmet Badur
Courtney Green
Avi Kumar
Austin Lefebvre
Noah Myers

Hui Zhang
Seth Parker
Nate Vacanti

Support

NSF CAREER Award
DOD Lung Cancer Research Program
NIH/NCI
Searle Scholars Program
California Institute of Regenerative Medicine
Hellman Faculty Fellowship
Lowy Medical Research Foundation
Camille and Henry Dreyfus Teacher-Scholar Award

UCSD

Pedro Cabrales
Rohit Loomba
Anne Murphy
Ajit Divakaruni

UCSD/VAMC SD

Ted Ciaraldi
Bob Henry

MIT

Matt Vander Heiden

SBMRI

Jorge Moscat
Maria Diaz-Meco

UPenn

Katy Wellen

Salk Institute

Reuben Shaw

UMass Worcester

Dave Guertin

UC Berkeley

Dan Nomura

