## The 1st International Electronic Conference on Forests — Forests for a Better Future: Sustainability, Innovation, Interdisciplinarity 15-30 November, 2020 - ONLINE Histology of Austrocedrus chilensis roots during infection by Phytophthora austrocedri

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Figure 1. Micrographs of sections of non-inoculated root tissue of A. chilensis seedlings (Fast Green staining). A. general view (100X). B. cortical parenchyma (CP) and Phi thickenings (400X). C. Vascular cylinder (CV), tracheids (T) and Phi thickenings (400X). D. Rhizodermis (Rh) and cortical parenchyma (CP) (400X).



Figure 2. Sections of root tissue of A. chilensis seedlings inoculated with P. austrocedri stained w and hypha growing intracellularly. C. Advance zone. Vascular cylinder (CV), cortical parenchyma (CP), hyphae of the pathogen (H), arrows indicate Phi thickenings.

In conclusion this is the first anatomo-histological study done on A. chilensis roots. Some alterations occur in a similar way to what was observed at the stems of inoculated seedlings (Vélez et al. 2012) and at the stems of adult trees naturally or artificially infected (Troncoso 2018). Results evidenced that A. chilensis triggers mechanisms to restrict and resist the infection, but P. austrocedri manages to evade them and finally colonizes and degrades host tissues.

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Phytophthora austrocedri was identified as the primary pathogen causing the "Mal del ciprés" disease in Patagonia, which affects the endemic conifer Austrocedrus chilensis. Trees present root necrosis that may extend to the stem. This study aimed to describe the histological alterations occurring during P. austrocedri infection of roots of A. chilensis seedlings. Plants were inoculated at the roots and histological studies were performed four weeks post-inoculation. Safranin-fast green, phloroglucinol-HCI, toluidine blue, lugol and diaminobenzidine stains were used to describe and compare anatomo-histological features observed in roots of noninoculated versus inoculated seedlings. In healthy tissues, the presence of Phi thickenings in cortical cells is reported for the first time for A. chilensis. In inoculated roots it was observed necrosis of the epidermis and of the cortical parenchyma, and alterations in parenchymal cells (loss of turgor and content, without starch, presence of phenolic compounds). Lignin content remained unaffected by the presence of P. austrocedri. The area occupied by Phi thickenings with was smaller in P. austrocedri-colonized tissues, and these structures showed an accumulation of toluidine blue (400X). A. Non-inoculated healthy tissue. B. Inoculation zone with discolored cortical cells polyphenols, absent in healthy tissue. Parenchymal cells, tracheids, and rays, showed production of hydrogen peroxide. Results evidenced that A. chilensis triggers mechanisms to restrict the infection, but P. austrocedri manages to evade them.







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Figure 3: Sections of root tissue of A. chilensis seedlings inoculated with P. austrocedri stained with phloroglucinol-HCl (400X). A. Non-inoculated healthy tissue. B. Inoculation zone. C. Advance zone. Vascular cylinder (CV), cortical parenchyma (CP), abnormality (A), arrows indicate Phi thickenings.



Figure 4: Sections of root tissue of A. chilensis seedlings inoculated with P. austrocedri stained with lugol (400X). A. Non-inoculated healthy tissue. B. Inoculation zone. C. Advance zone. Vascular cylinder (CV), cortical parenchyma (CP), necrotic zone (ZN), hyphae (H), arrows indicate Phi thickenings

