



Proceedings

# Introgression of Bacterial Blight Resistance Genes (*Xa*21, *xa*13 and *xa*5) into CB 174 R, an Elite Restorer Line in Rice <sup>+</sup>

# Govintharaj Ponnaiah \*, Manonmani Swaminathan, Karthika Gunasekaran and Robin Sabariappan

Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

- \* Correspondence: govinth.tnau@gmail.com; Tel.: +91-994-226-0287
- + Presented at the 1st International Electronic Conference on Plant Science, 1–15 December 2020; Available online: https://iecps2020.sciforum.net/.

Published: 1 December 2020

**Abstract:** Bacterial blight caused by *Xanthomonas oryzae* pv. *oryzae* (Xoo) is one of the major disease caused severe yield reduction in the rice growing regions. One dominant (*Xa*21) and two recessive genes (*xa*13 and *xa*5) were introgressed into CB 174 R through marker assisted breeding. The present study found three (*Xa*21 + *xa*13 + *xa*5) and two (*Xa*21 + *xa*13 or *Xa*21 + *xa*5 or *xa*5 + *xa*13) genes introgressed combinations in the early segregated materials through foreground selection. The identified homozygous/heterozygous individuals forwarded to next cycles of breeding to fix homozygous conditions for all three genes with an improved agronomic performance background, and thus could be used as a donor source for future rice breeding programme.

Keywords: marker assisted selection; gene specific marker; gene pyramiding

# 1. Introduction

Rice (*Oryza sativa* L.) is an important staple cereal food crop for half of the world populations. Bacterial blight (BB) caused by *Xanthomonas oryzae* pv. *oryzae* (*Xoo*), which is significantly reduces the yield up to 80% in rice [1]. Till date, a total of 40 BB have been identified from wild, cultivated and mutant population of rice [2]. Of these, BB resistance genes *Xa3*, *Xa4*, *Xa7*, and *Xa21*, have been extensively utilized by the breeders in their breeding programs [3–7]. Breeding strategies like stacking of multiple resistance genes into the elite genetic background [8]. In this context, marker assisted selection (MAS) is an efficient and cost effective approach along with precise phenotyping for disease free cultivars development which has been proved by several rice researchers in the past [9,10]. The CORH 04 is a medium duration grain quality hybrid popular among farmers which was released by Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India. However, hybrid CORH 04 was susceptible to BB disease in the rice grown areas resulted significant yield reduction. The CB 174 R is an elite restorer line have been involved as parent in several released high yielding rice hybrids. An improvement of parental lines of the hybrid would be the best option to develop resistance against BB disease through MAS [11]. Therefore, the present study was aimed to introgress the BB resistance genes to parental lines of the released rice hybrid through MAS.

The 1st International Electronic Conference on Plant Science, 1–15 December 2020 2. Experiments

The experimental material CB 174 R, is an elite restorer line of popular released rice hybrid CORH 4, used as a recurrent parent. The parent IRBB 60 with two recessive genes (*xa*5 and *xa*13) and one dominant gene (*Xa*21) is used as a donor. Two functional markers (xa 5 and xa 13) [12,13] and one SSR (simple sequence repeats) (RM 21 for *Xa*21 gene) [14] markers used to tag traits of interest. The hybrid F<sub>1</sub> generated by crossing CB 174 R and IRBB 60. The BB resistance genes confirmed F<sub>1</sub> plant along with phenotypically desirable plant tagged and advanced to F<sub>2</sub> through self fertilization breeding. Hundred and ten F<sub>2</sub> individual plants screened for BB genes by employing gene-specific and SSR markers, and phenotyped for BB isolate. Fifty four out of 110 F<sub>2</sub> individual plant were possessed all three or two BB resistance genes tagged with foreground selection and forwarded to F<sub>2.3</sub> through self fertilization. All these field experiments were conducted at Department of Rice (11° N, 77° E, and 427 m above mean sea level), Paddy Breeding Station (PBS), Tamil Nadu Agricultural University, Coimbatore, India.

Two grams of fresh leaf bits collected from 18 days old seedlings of 110 F<sub>2</sub> individuals used for genomic DNA (Deoxyrhibonucleic Acid) using CTAB (Cetyltrimethylammonium bromide) method as described by Doyle and Doyle [15]. The PCR (Polymerase chain reaction) performed for two functional markers and one SSRs with an initial denaturation at 94 °C for 5 min 35 cycles of 1 min denaturation at 94 °C for 1 min annealing (for xa5–56 °C for xa13–59 °C RM 21–55 °C) and 1.30 min for primer extension at 72 °C, and the final extension at 72 °C for 7 min. The 5  $\mu$ L PCR product subjected to gel electrophoresis and then bands visualized using UV trans-illumination after ethidium bromide staining. For, functional marker *xa5*, the PCR product was digested with *Bsr* 1 and bands were visualized as same like other markers.

One isolate of *Xoo*, prevalent in major rice growing areas was isolated, and multiplied on peptone sucrose agar plates and incubated for 48 h at 28 °C and then inoculum of the isolate into suspension by adding 10 mL of distilled water per slant to give a concentration of bacterial cells of about  $10^8$  to  $10^9$  colony forming units (CFU)/mL. Hundred and ten F<sub>2</sub> individuals and their parents were artificially inoculated the Xoo isolate when plants reached maximum of tillering as described earlier by Kauffman et al. [16]. Disease reaction scoring was done 14 days after inoculation based on standard evaluation system in 2011–2012 (SES 2011–2012) (Table 1).

S.No.	Score	<b>Description (Affected Lesion Area)</b>		
1	0–1 (Resistance)	1–5% of leaf area affected		
2	1–3 (Moderately Resistance)	6–12% of leaf area affected		
3	3–5 (Moderately Susceptible)	13–25% of leaf area affected		
4	5–7 (Susceptible)	26–50% of leaf area affected		
5	7–9 (Highly Susceptible)	51–100% of leaf area affected		

Table 1. SES scale for bacterial leaf blight (2011-2012).

#### 3. Results

The 110 F<sub>2</sub> individuals using foreground selection molecular markers led identify combinations of genes introgressed individuals (Figure 1). None of the F<sub>2</sub> individuals showed homozygous resistance loci for all three genes. Though, more number of F<sub>2</sub> individuals in CB 174R × IRBB 60 identified having homozygous resistance for two loci (*Xa*21*Xa*21and *xa*13*xa*13) and also three F<sub>2</sub> individuals identified having two heterozygous resistance loci (*Xa*5*xa*5 and *Xa*21*xa*21). Furthermore, 5 F<sub>2</sub> individuals were identified in heterozygous state in all three genes (*Xa*5*xa*5, *Xa*13*xa*13 and *Xa*21*Xa*21) and also two individuals were identified having heterozygous resistance for two loci (*Xa*5*xa*5 and *Xa*13*xa*13) and homozygous for other locus (*Xa*21*Xa*21). Based on phenotypic score and resistance gene's combination, 54 F<sub>2</sub> individuals were selected, selfed and advanced to next generation (F<sub>2:3</sub>). Nine gene introgressed F<sub>2:3</sub> individuals showed maximized yield potential than parents with recurrent background features (Table 2).

The 1st International Electronic Conference on Plant Science, 1-15 December 2020



(b)

**Figure 1.** Gene tagging of  $F_2$  population derived from cross between CB 174 R × IRBB 60 for functional marker (**a**) *xa*13, and (**b**) *Xa*21 linked bacterial blight resistance genes.

Plant. No	PH (cm)	NPT	PL (cm)	NG	TGW (g)	SPY (g)
1	115	16	25	102	21.90	35.5
2	105	14	23	192	24.05	34.5
3	116	15	27	198	18.52	41.0
4	119	12	26	141	23.62	33.0
5	115	17	20	137	21.07	37.0
6	105	19	21	129	23.4	32.5
7	94	22	20	167	22.18	32.0
8	115	17	14	95	22.09	31.5
9	100	17	21	253	24.70	48.5
CB 174 R	146.67	13	31.67	270.67	16.35	28.14
IRBB 60	88	11	27	124	13.60	22.90

Table 2. Morphological characteristics of selected progenies of F2:3 population of CB 174 R × IRBB60.

Note: PH: Plant height, NPT: Number of productive tillers, PL: Panicle length, NG: Number of grains per panicle, TGW: Thousand grain weight, SPY: Single plant yield.

# 4. Discussion

Fifty four out of 110 F<sub>2</sub> individuals identified as having three/two genes combination in this study. Of these, 42 individuals having fertility restoration gene *Rf*4 characterized earlier in CB 174R × IRBB 60 by Govintharaj et al. [17]. We further focused 5 F<sub>2</sub> individuals were in heterozygous state for all three genes (Xa5xa5, Xa13xa13 and Xa21Xa21), and also two individuals had heterozygous resistance for two loci (Xa5xa5 and Xa13xa13) and homozygous for one locus (Xa21Xa21), along with fertility genes which were characterized earlier. Presence of Xa21 in homozygote or heterozygote

The 1st International Electronic Conference on Plant Science, 1-15 December 2020

state in combinations with other genes found to have higher level of resistance. Also, two recessive genes shown higher level of resistance when they were in homozygote (xa5xa5 and xa13xa13) than heterozygote (Xa5xa5 and Xa13xa13) condition. Similar to this study, Perumalsamy et al. [18] who have pyramided three BB resistance genes (xa5, xa13 and Xa21) using functional markers in rice. More than 80% of the F<sub>2</sub> individuals possessed Xa21 + xa13 genes combination in this study showed higher level of resistance. Several studies have been showed same level of resistance (Xa21 + xa13) with gene-pyramided lines like in Samba Mahsuri, PR106, Pusa Basmati 1 and IR 24 could provide long-lasting resistance in India [19–21]. It has been stated that broad spectrum of resistance observed when multiple genes introgressed into a elite line rather than single gene against BB resistance [22]. The identified different combinations of homozygous/heterozygous resistance plants F<sub>2</sub> with fertility restoration genes, and the subsequent F<sub>2.3</sub> families showed an improved agronomic performance would be used as a donor parent for future rice breeding programme.

## 5. Conclusions

BB resistance genes identified in heterozygous and/or homozgygous with superior agronomic performances of the studied breeding materials led to use as a donor parent in the BB resistance genes introgression breeding.

**Author Contributions:** S.M., G.P. and R.S. conceived and designed the experiments; G.P. performed the experiments; G.P. and S.M. analyzed the data; S.M. contributed reagents/materials/analysis tools; G.P., S.M. and K.G. wrote the paper. All authors have read and agreed to the published version of the manuscript.

**Acknowledgments:** This research work was carried out as part of a M.Sc. thesis of Govintharaj, submitted to Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu.

Conflicts of Interest: The authors declare no conflict of interest.

#### Abbreviations

- BB Bacterial blight
- MAS Marker assisted selection
- SSR Simple sequence repeats
- DNA Deoxyrhibo Nucleic Acid
- CTAB Cetyltrimethylammonium bromide
- PCR Polymerase chain reaction
- CFU Colony forming unit
- SES Standard evaluation system

### References

- Srinivasan, B.; Gnanamanickam, S.S. Identification of a new source of resistance in wild rice *Oryza rufipogon* to bacterial blight of rice caused by Indian strains of *Xanthomonas oryzae* pv. *oryzae*. *Curr. Sci.* 2005, *88*, 1229–1231. Available online: https://www.jstor.org/stable/24110290.
- Neelam, K.; Mahajan, R.; Gupta, V.; Bhatia, D.; Gill, B.K.; Komal, R.; Lore, J.S.; Mangat, G.S.; Singh, K. High-resolution genetic mapping of a novel bacterial blight resistance gene *xa*-45(t) identified from *Oryza glaberrima* and transferred to *Oryza sativa*. *Theor. Appl. Genet.* 2020, *133*, 689–705, doi:10.1007/s00122-019-03501-2.
- Chen, S.; Liu, X.; Zeng, L.; Ouyang, D.; Yang, J.; Zhu, X. Genetic analysis and molecular mapping of a novel recessive gene *xa*34 (t) for resistance against *Xanthomonas oryzae* pv. *oryzae*. *Theor. Appl. Genet.* 2011, 122, 1331–1338, doi:10.1007/s00122-011-1534-7.
- 4. Luo, Y.; Ma, T.; Zhang, A.; Ong, K.H.; Li, Z.; Yang, J.; Yin, Z. Marker-assisted breeding of the rice restorer line Wanhui 6725 for disease resistance, submergence tolerance and aromatic fragrance. *Rice* 2016, *9*, 66, doi:10.1186/s12284-016-0139-9.

The 1st International Electronic Conference on Plant Science, 1–15 December 2020

- 5. Shalini, T.; Govintharaj, P.; Ameenal, M.; Manonmani, S.; Robin, S. Development of bacterial blight resistant genetic stocks by stacking three genes (*Xa*21, *xa*13 & *xa*5) through marker assisted recombination breeding. *Bioscan* **2016**, *11*, 2331–2334.
- 6. Govintharaj, P.; Tannidi, S.; Swaminathan, M.; Robin, S. Effectiveness of selection, parent-offspring correlation and regression in bacterial blight resistance genes introgressed rice segregating population. *Ci enc. Rural* **2017**, *47*, 1–6, doi:10.1590/0103-8478cr20160987.
- 7. Govintharaj, P.; Manonmani, S.; Robin, S. Variability and genetic diversity study in an advanced segregating population of rice with bacterial blight resistance genes introgressed. *Cienc. Agrotec.* **2018**, *42*, 291–296, doi:10.1590/1413-70542018423022317.
- 8. Rajpurohit, D.; Kumar, R.; Kumar, M.; Paul, P.; Awasthi, A.; Basha, P.O.; Puri, A.; Jhang, T.; Singh, K.; Dhaliwal, H.S. Pyramiding of two bacterial blight resistance and a semi-dwarfing gene in Type 3 Basmati using marker-assisted selection. *Euphytica* **2011**, *178*, 111–126, doi:10.1007/s10681-010-0279-8.
- Ni, D.; Song, F.; Ni, J.; Zhang, A.; Wang, C.; Zhao, K.; Yang, Y.; Wei, P.; Yang, J.; Li, L. Marker-assisted selection of two-line hybrid rice for disease resistance to rice blast and bacterial blight. *Field Crops Res.* 2015, 184, 1–8, doi:10.1016/j.fcr.2015.07.018.
- Pradhan, S.K.; Nayak, D.K.; Pandit, E.; Barik, S.R.; Mohanty, S.P.; Anandan, A.; Reddy, J.N. Characterization of morpho-quality traits and validation of bacterial blight resistance in pyramided rice genotypes under various hotspots of India. *Aust. J. Crop Sci.* 2015, *9*, 127–134.
- 11. Prabhu, A.S.; Guimarães, E.P.; Filippi, M.C.; Araujo, L.G.; Cutrim, V.A. Expression of resistance in rice hybrids to *Pyricularia grisea*. *Fitopatol*. *Bras*. **2002**, *27*, 454–460, doi:10.1590/S0100-41582002000500003.
- 12. Iyer-Pascuzzi, A.; McCouch, S. Recessive resistance genes and the *Oryza sativa-Xanthomonas oryzae* pv. *oryzae* pathosystem. *Mol. Plant Microbe Interact.* **2007**, *20*, 731–739, doi:10.1094/mpmi-20-7-0731.
- Han, K.D.; Ling-Guang, K.; Yan-Hu, J.; Ming, C.; Zhi-Gang, C.; Xin-Hua, D.; Zhao-Hui, C. Screening and Identification of Susceptibility and Fertility Related Mutants from Transgenic Rice IRBB13 (Pxa13:GFP). *Sci. Agric. Sin.* 2012, 45, 3899–3908, doi:10.3864/j.issn.0578-1752.2012.19.001.
- 14. Cao, J.; Bao-Rong, L.U.; Hui, X.; Jun, R.; Francesco, S.; Alberto, S.; Fabrizio, G. Genetic Diversity and Origin of Weedy Rice (*Oryza sativa f. spontanea*) Populations Found in North-eastern China Revealed by Sequence Repeat (SSR) Markers. *Ann. Bot.* **2006**, *98*, 1241–1252, doi:10.1093%2Faob%2Fmcl210.
- 15. Doyle, J.J.; Doyle, J.L. Isolation of plant DNA from fresh tissue. *Focus* **1990**, *12*, 13–15.
- 16. Kauffman, H.E.; Reddy, A.; Hsieh, S.P.Y.; Merca, S.D. An improved technique for evaluating of varieties to Xanthomonos oryzae pv.oryzae. *Plant Dis. Rep.* **1973**, *57*, 537–541.
- 17. Govintharaj, P.; Manonmani, M.; Robin, S. Screening for fertility restoration genes linked marker in bacterial blight resistance genes introgressed segregating population in rice. *Vegetos* **2018**, *31*, 158–161, doi:10.5958/2229-4473.2018.00021.6.
- Perumalsamy, S.; Bharani, M.; Sudha, M.; Nagarajan, P.; Arul, L.; Saraswathi, R.; Balasubramanian, P.; Ramalingam, J. Functional marker-assisted selection for bacterial leaf blight resistance genes in rice (*Oryza sativa* L.). *Plant Breed.* 2010, *129*, 400–406, doi:10.1111/j.1439-0523.2009.01705.x.
- Rajpurohit, D.; Kumar, R.; Kumar, M.; Paul, P.; Awasthi, A.; Basha, P.O.; Puri, A.; Jhang, T.; Singh, K.; Dhaliwal, H.S. Pyramiding of two bacterial blight resistance and a semidwarfing gene in Type 3 Basmati using marker-assisted selection. *Euphytica* 2011, *178*, 111–126, doi:10.1007/s10681-010-0279-8.
- 20. Singh, S.; Sidhu, J.S.; Huang, N.; Vikal, Y.; Li, Z.; Brar, D.S.; Dhaliwal, H.S.; Khush, G.S. Pyramiding three bacterial blight resistance genes (*xa*5, *xa*13 and *Xa*21) using marker-assisted selection into indica rice cultivar PR106. *Theor. Appl. Genet.* **2001**, *102*, 1011–1013, doi:10.1007/s001220000495.
- 21. Zhang, J.; Li, X.; Jiang, G.; Xu, Y.; He, Y. Pyramiding of *Xa*7 and *Xa*21 for the improvement of disease resistance to bacterial blight in hybrid rice. *Plant Breed.* **2006**, *125*, 600–605, doi:10.1111/j.1439-0523.2006.01281.x.
- 22. Chen, J.M.; Fu, Z.Y.; Quan, B.Q.; Tian, D.G.; Li, G.; Wang, F. Breeding hybrid rice restoring line with double resistance to rice blast and bacterial blight by marker-assisted selection. *Mol. Plant Breed.* **2009**, *7*, 465–470.

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

The 1st International Electronic Conference on Plant Science, 1–15 December 2020



© 2020 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons by Attribution (CC-BY) license (http://creativecommons.org/licenses/by/4.0/).