

Laboratory of Animal Toxins

School of Pharmaceutical Sciences of Ribeirão Preto – FCFRP – USP. São Paulo, Brazil



Expression and purification of rTs7, a recombinant toxin from *Tityus serrulatus* scorpion venom

Jacob, B.C.S¹; Cordeiro, F.A¹; Wiezel, G.A.¹; Cardoso, I.A.¹; Arantes, E.C.¹

¹ Department of BioMolecular Sciences, School of Pharmaceutical Sciences of Ribeirão Preto, University of São Paulo, Ribeirão Preto, SP, Brazil

*e-mail: beatrizjacobcs@gmail.com

INTRODUCTION

Tityus serrulatus venom is composed of several substances,

including the neurotoxins that interact with ion channels. These channels are involved in many diseases, such as

RESULTS



Figure 1: SDS gel with the screening of positive 1: Molecular weight marker; 2: negative control; 3: positive control, **4-9**:

arrhythmia, autoimmune diseases, hypertension and immune response to infections making T. serrulatus venom a source of biological tools to study them. The Ts7, acts selectively on potassium channels, and can contribute to the treatment of Kv1.3 channel-related diseases, such as autoimmune diseases.

OBJECTIVE

In this work, we present the heterologous expression of

Ts7 in *Pichia pastoris* yeast and its purification.

METHODS

The gene was synthetized by GenScript[®] with TEV (tobacco

Figure 2: Laboratorial-scale expression and purification of rTs7. (A) The expression fractions at 0-144 h. 1: ultra low molecular mass pattern; 2: negative control; 3-9: times 0h-144h. (B) Fractions from IMAC. 1: ultra low molecular mass pattern; 2: VOID; 3: Washed; 4-10: fractions from IMAC (10 mM to 250 mM of imidazole).





etch virus) protease cleavage site before N-terminal sequence and cloned into pPICZaA vector. The *P. pastoris* cells (KM71H strain) were transformed with the linearized plasmid rTs7 pPICZaA. Transformation was confirmed by PCR. Positively transformed colonies were submitted to a screening in a 24-wells plate, under standard conditions (pH 6, for 144 h). And after, the best of them was subjected to laboratorial-scale expression, monitored by SDS-PAGE. The expressed protein was purified by reversed-phase chromatography, on a C-18 column. Three fractions were observed and, analyzed using mass spectrometry.

CONCLUSIONS



Figure 3: Reversed phase chromatography of IMAC fractions. Monitored by the FPLC Äkta UPC 900 system at 214 nm. The fraction was applied to the analytical C18 column (250 x 4.6 mm, with 5 µm particles, Jupiter, Phenomenex) previously equilibrated with 0.1% TFA.



The rTs7 was successfully expressed and purified, with a

high yield of the recombinant toxin, which showed

similarity with the native toxin, its and

immunosuppressive activity in multiple sclerosis model

will be further investigated.

SUPPORT



Figure 4: (A to C). Mass spectra of 1, 2 and 3 fractions from reversed phase in MALDI-TOF positive linear mode.