

# Multi-Omics-Driven Biomarkers for Precision Medicine in Cutaneous Melanoma<sup>†</sup>

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**Abstract:** Cutaneous melanoma (CM) is a public health issue and a major challenge for scientists. At the dawn of the "omics era", we witnessed groundbreaking advances in CM molecular stratification and therapeutic management assisted by genomic profiling and sequencing technologies. However, melanomagenesis is a complex and multifactorial process that cannot be restricted only to genomic aspects, requiring investigation from a multi-omics perspective. The Cancer Genome Atlas (TCGA) is an important repository of multiple types of omics data, and in particular, for CM, it has been shown that multi-integration can lead to prognosis models with improved prediction performance. We present herein how droplet digital PCR (ddPCR), a relatively young omics technology can complement existing approaches in the field to detect multiple types of alterations in both body fluids as well as formalin-fixed tissues harvested from CM patients and how these findings may broaden our vision on CM research, diagnosis, prognosis and therapy in the context of precision medicine.

**Keywords:** cutaneous melanoma; ddPCR; omics; biomarkers; ctDNA; liquid biopsy; targeted therapy; immunotherapy

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## 1. Multi-omics-based biomarkers: the roadmap towards personalized care in CM

Skin melanoma is one of the most heterogeneous and metastatic malignancies, with an increasing incidence in fair-skinned populations [1]. Genomic interrogation, assisted by high-throughput sequencing technologies and microarrays, has guided the rudimentary stratification of these tumors and the therapeutic decisions [2]. However, it has been shown that the pathogenesis of CM is more complex than previously thought, involving dramatic transformations at almost all tumor levels: genomic, epigenomic, transcriptomic, proteomic, metabolomic, and so forth [3,4]. Therefore, to better capture and characterize biological events associated with prognosis or response to therapy in cancers, scientific research has undergone a remarkably swift transition from single-level tumor interrogation to multidimensional omics research. The recent advancements in technology and bioinformatics equipped us with data on multiple types of omics measurements, such as mRNA-gene expression, DNA methylation, microRNAs (miRNAs), copy-number variations (CNVs), and so on. These types of omics activities may be independent or overlap, reflecting distinctive patterns of the disease [5].

Recent studies have revealed the importance of multi-integration for precision medicine in CM. Several research groups have focused on the prognosis potential of multi-omics data. For instance, it has been proved that the integration of gene expression, DNA methylation, and CNVs data may reveal new dysregulated signaling pathways in CM, with important implications for prognosis [3].

Similarly, an integrative study analyzing stage III patients confirmed the prognostic capabilities of gene expression, proteins, and microRNAs in correlation with clinical, pathological, and mutational data [6]. Other studies have focused on the predictive value of multi-omics analysis in CM. For example, in the HOPE project, whole-exome sequencing (WES) and gene expression profiling data revealed that complete remission under the clinically approved anti-PD1 antibody (e.g. Nivolumab, Opdivo) is associated with elevated levels of PDL-1 protein and an increased number of single nucleotide variants (SNVs) [7]. Therefore, in the future, evaluation of this disease from a multi-omics perspective is expected to provide a more specific molecular classification of CM but also to give clues about the mechanisms that drive tumorigenesis, metastasis, and resistance to therapy in these tumors [8].

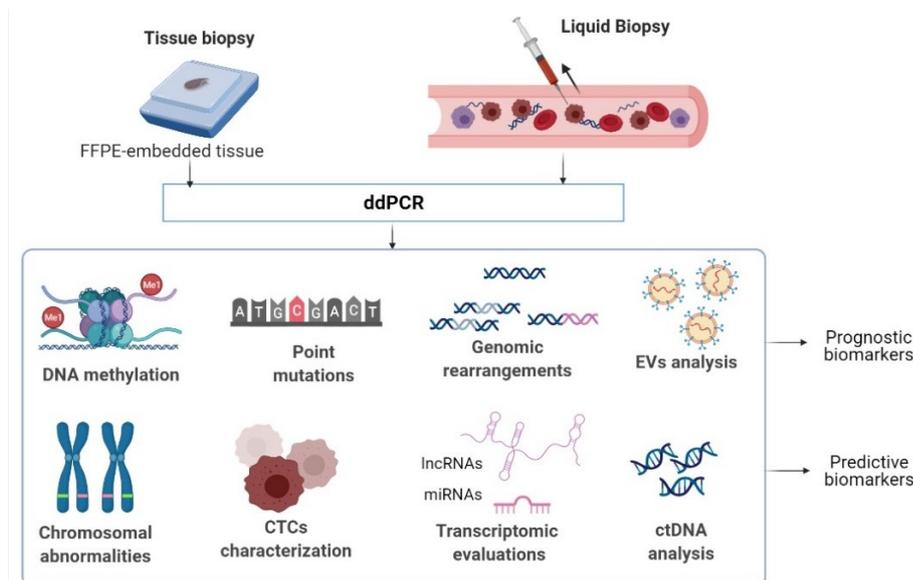
Droplet digital PCR (ddPCR), an accurate and relatively inexpensive omics technology, is arousing considerable scientific interest in the field of biomarker research [9]. This technology enables the investigation and validation of several types of omics alterations detected by whole-genome screening (WGS) techniques, such as Next-Generation Sequencing (NGS). NGS provides a comprehensive picture of the mutational inventory of a tumor; however, the technology is not suitable for tracking mutations over time, as it is extremely expensive and meticulous [9]. Yet, combining NGS with ddPCR can broaden the applications and increase the benefits of both techniques. For instance, once a set of mutations was identified by NGS in ctDNA, researchers can exploit ddPCR, which is less expensive and laborious, to interrogate that set of biomarkers and obtain relevant information about tumor progression and response to therapy [10].

## 2. Droplet digital PCR: a versatile omics technology in oncology

ddPCR, a highly sensitive and specific technology, relies on a water-oil emulsion droplet system that involves the partitioning of nucleic acid samples into 20,000 nanoliter-sized droplets that serve as independent test tubes or reactions [11]. By employing oil, water, and a chemical stabilizer emulsion, each sample is diluted into thousands of individual partitions, some of which do not contain template DNA and others that contain one or more target sequences [12]. A PCR reaction takes place in each tube. Each partition is then examined for amplified target DNA by fluorescence so that the number of positive and negative droplets can be counted, facilitating the quantification of target molecules under the assumption of Poisson distribution [13]. The limit of detection (LoD) is about 0.005%, lower than that of RT-PCR (1%), pyrosequencing (5%), melting curve analysis (10%), and Sanger sequencing (20%) [14]. Through the partitioning process, ddPCR brings multiple advantages over traditional PCR techniques, that include: absolute quantification of DNA copies in the input samples without the need for external calibration curves used in qPCR; low susceptibility to PCR inhibitors; increased accuracy, especially when working with low concentrations of samples or degraded samples, as well as reproducibility and increased sensitivity of the experiments [12]. Applications of this technology may include absolute allele quantification, CNVs analysis, rare mutations, DNA methylation detection, transcriptomic evaluations (mRNA, miRNA), and genetic rearrangements in various types of biological samples [12,15–18]. However, the use of ddPCR requires the identification of specific genetic alterations, which is why it almost always accompanies whole-genome profiling technologies such as NGS [19,20].

ddPCR is useful both for the analysis of archived tumor tissues, which are degraded and have a limited concentration of DNA, but also for the analysis of biological fluids [21]. In recent years, liquid biopsy (LB), based on the analysis of circulating components derived from tumors - circulating tumor DNA (ctDNA), RNA, extracellular vesicles (EVs), or tumor cells (CTCs) has gained tremendous attention because of its potential to provide in real-time an accurate description of the genetic landscape of a tumor [22]. LB has proven to be more informative compared to tissue biopsy, which is spatially limited and ineloquent for tumor evolution [22]. Thus, only by analyzing the biological material from patients' body fluids, researchers and clinicians can obtain valuable information on the dynamics of the genetic profile of the tumor, which can be integrated and used to guide the therapeutic management for each patient [20]. Due to its versatility and ability to operate with small amounts of biological material, ddPCR is an ideal methodology for analyzing LBs (Figure 1) [21].

Several blood-based biomarkers interrogated by ddPCR have also found diagnostic, predictive, and monitoring applications in certain malignancies [21]. Novel approaches use some other body fluids such as cerebrospinal fluid or urine to screen for and validate biomarkers in cancer patients [21]. However, in this article, we aim to briefly present the applications of ddPCR for biomarker research in CM.



**Figure 1.** Applications of ddPCR for biomarker research in oncology. FFPE tissue- formalin-fixed paraffin-embedded tissue; CTCs- circulating tumor cells; EVs- extracellular vesicles; ctDNA- circulating tumor DNA; miRNAs- microRNAs; lncRNAs- long non-coding RNAs.

### 3. Interrogating CM mutational landscape via ddPCR

#### 3.1. Screening for biomarkers in tissue biopsies

Although highly invasive and ineloquent for tumor heterogeneity, tissue biopsy remains the gold standard for clinical molecular analyzes in cancer. Particularly for CM, tissue biopsy studies focus on hotspot mutations such as BRAF and KRAS, which are critical for guiding therapeutic decisions in clinical management. Remarkably, ddPCR showed enhanced sensitivity compared to the widely used Cobas® 4800 system based on real-time PCR amplification, Sanger sequencing, and allele-specific PCR (AS-PCR) (35.6% vs. 9.2%, 26.4%, and 26.4%) in the detection of BRAF V600E mutations in FFPE tissues from 87 CM patients diagnosed in different Breslow stages [23]. In a group of 8 patients in the clinical cohort, the BRAF V600E mutation was detectable only by ddPCR; notably, these patients could have benefited from vemurafenib [23]. 5 out of these 8 patients who tested BRAF V600E positive only through ddPCR later developed sentinel lymph node metastases, highlighting that ddPCR should be the primary method for detection and monitoring of BRAF V600E mutant melanomas [23]. In parallel, another study demonstrated the superiority of ddPCR compared to Sanger sequencing and pyrosequencing in the detection of common BRAF, NRAS, and TERT promoter mutations in 40 archived melanoma tissues [14]. Overall, ddPCR was much more sensitive, detecting mutations in 12.5% and 23% of tumors classified as wild-type by pyrosequencing and Sanger sequencing. The sensitivity of ddPCR was also much higher in tumors with <50% tumor cellularity, providing a rationale for the use of ddPCR in the detection and monitoring of human melanomas [14].

Recently, Salgado et al used ddPCR to investigate the molecular mechanisms associated with TERT reactivation in human melanomas. CpG methylation in the TERT promoter (TERTp) was associated with TERT mRNA expression [24]. Hence, two hotspot mutations in TERTp termed C228T and C250T, have been documented to facilitate binding of transcription factor E26 transformation-specific / ternary complex factor (ETS / TCF), and subsequent TERT induction. To elucidate the genetic and epigenetic mechanisms regulating TERT gene expression in CM, Salgado et al.

designated a ddPCR protocol able to assess TERTp methylation fraction (MF), alongside C228T and C250T TERTp mutations in 44 healthy, benign and malignant tumor samples [24]. They observed that TERTp methylation is correlated with chromatin accessibility and TERT expression levels in melanoma cell lines; thus, due to increased TERTp methylation, TERT expression requires an open chromatin state in TERTp-wild type samples or a combination of moderate MF and chromatin accessibility in the presence of C228T/C250T hotspot mutations [24]. Given that TERTp hypermethylation has been proposed as an indicator of a poor patient prognosis in CM and certain other tumors, quantifying TERT methylation by ddPCR may open new perspectives for the prognosis and monitoring of CM patients [24].

### 3.2. Searching for biomarkers in liquid biopsies

In the first instance, ddPCR has proven to be an ideal methodology for the analysis of cell-free ctDNA in CM patients' plasma. ctDNAs are short DNA fragments (134-144 base pairs) derived from tumor cells that have undergone necrosis or apoptosis [25]. These fragments are released into the bloodstream and are actively investigated as surrogate biomarkers for the analysis of genomic and epigenomic alterations in primary tumors and metastatic lesions [26]. By far, ctDNA mutations (mt) are of considerable interest for CM research. It has been reported that BRAF mt may be reliable indicators for response to targeted therapy in CM, being detectable in 70% of patients in the non-responsive group and only in 10% of patients in the responsive group [27]. In line with these observations, Tsao et al confirmed that the screening of BRAF and NRAS mt in ctDNA by ddPCR is an effective method to monitor the response to targeted therapy in stage IV melanoma patients, being even more informative than LDH, a blood-based biomarker correlated with disease relapse [28]. In parallel, other studies have shown that lower baseline ctDNA mt detected through ddPCR, including BRAF V600E and BRAF V600K mt may associate with higher response rates and improved survival in CM patients treated with BRAFi [29,30]. Furthermore, it has been suggested that the basal levels of ctDNA, which can also be assessed through ddPCR, may be important predictive biomarkers in CM patients who underwent immunotherapy; briefly, patients with undetectable ctDNA at baseline had superior progression-free survival (PFS) and overall survival (OS) rates than those with detectable ctDNA [31]. Nonetheless, other research groups revealed that besides the baseline ctDNA status, the dynamics of ctDNA may provide valuable information on the patient's clinical outcome following immunotherapy [32].

ddPCR can also be used to assess the methylation status of ctDNA. DNA methylation patterns are extremely dynamic during tumor progression, so DNA methylation analysis can provide important information about disease evolution and response to treatment [26]. Although several methylation-based ctDNA LB assays have been validated for human cancers (liver, lung, and colorectal cancer), currently for CM there is no specific methylation panel that includes biomarkers associated with tumor progression or resistance to therapy [33]. However, detection of certain hypermethylated tumor suppressor genes, such as PTEN, CDKN2A, RASSF1A, and MGMT in plasma has been reported to have diagnostic value in CM patients [34]. Additionally, it was found that the methylation level of transposable element LINE-1 may be associated with poor OS in stage III CM patients [34]. Given this information, it would be interesting to verify these putative blood biomarkers through ddPCR, in order to obtain more accurate clinical information on the prognosis and disease evolution in CM patients.

Additionally, ddPCR may also be a promising methodology for evaluating miRNA expression in CM patients' body fluids [26]. miRNAs are single-stranded RNA molecules (18-22 nucleotides in length) that can regulate the expression of their target genes by binding the complementary mRNA sequences at the 3' untranslated region (3' UTR)[26]. Circulating miRNAs, released from tumor cells into the bloodstream are stable and highly accessible molecules and may be exploited as promising biomarkers in CM. Certain molecules identified through conventional methods (RT-qPCR) in the sera of CM patients, such as miR-221, miR-199a-5p, miR-33a, miR-424, and miR-206 have been reported to play valuable prognosis and diagnosis roles in CM clinical management [35]. At the moment, there are no published studies evaluating CM-related miRNAs in patients' plasma by ddPCR; there were

just a few studies with melanoma cell lines [36]. However, ddPCR has been recently applied to assess miRNA-34b/c methylation status in cfDNA in malignant pleural mesothelioma [37]. Therefore, although a nascent domain, we believe that ddPCR evaluation of CM-associated miRNAs in patients' blood may open new perspectives in the clinical management of CM tumors.

Furthermore, ddPCR can be extremely useful in the analysis and characterization of CTCs. Dissociated either from the primary tumor or metastatic compartments into the blood, these cancer cells may offer valuable information on the tissue of origin, but also on the prognosis and clinical outcome of patients [26]. The first prospective application of ddPCR in the analysis of melanoma CTCs consists in the quantification of specific melanoma-associated antigens (MAAs), such as MAGE-A3, PAX3, and MART-1, which are strong predictive biomarkers [26]. The second prospective application of this technique focuses on digital quantification of CTC-derived transcripts, which will help in predicting the response to targeted therapies and immunotherapies in CM [38]. A recent study, led by Hong et al. highlighted that ddPCR evaluation of 19 melanoma CTC-derived transcripts facilitates non-invasive monitoring of tumor burden in CM patients, supporting the rational application of immunotherapy in these subjects [38]. Furthermore, they also reported that a decline in CTC score at 7 weeks is positively associated with improved OS, whereas a rise in CTC score led to poor survival in 53% of patients. Given that there is virtually no blood-based biomarker for tumor burden and the neural crest origin of melanoma cells provides unique RNA transcripts that help in distinguishing CTCs from normal blood cells, this ddPCR protocol offers new hopes in the fight against drug resistance in CM patients [38].

Another methodology for validating biomarkers in CM refers to the identification of CM-associated mutations in EVs through ddPCR. Released from both cancer and stromal cells, EVs are cup-shaped nanovesicles containing (mi)RNAs, DNAs, and proteins, that have key roles in metastatic niche preparation [39]. Zocco et al used ddPCR technology to assess the BRAF V600E prognostic power in EV-derived DNA [40]. Similar to other reports, the authors found that BRAF V600E copy levels above 50 copies/ml of plasma are suggestive of a poor prognosis and that the dynamics of BRAF V600E copy numbers may be relevant for monitoring the response to BRAF inhibitors (BRAFi) in CM patients [40]. Briefly, BRAF V600E copy levels have been found to become almost undetectable after exposure to BRAFi, but grow rapidly once the tumors become refractory to therapy and the disease progresses [40]. Additionally, Clark et al developed a ddPCR protocol for the detection of BRAF splicing variants p61, p55, p48, and p41, in cell-free RNA (cfRNA) derived from CM patients plasma [41]. Notably, 24 of 38 patients enrolled in the study and treated with BRAF / MEK inhibitor showed an increase in ctDNA levels as a sign of disease progression after treatment initiation. BRAF splicing variants were detected in 3 of these 38 patients: 2 patients harbored the BRAF p61 variant, while one presented the p55 variant. Remarkably, RNA isolation and analysis of EVs from CM resistant cell lines and patient plasma showed that BRAF splicing variants are specific to EVs, suggesting that besides ctDNA, RNA encapsulated in EVs may provide specific information about the tumor [41]. In parallel, another study reported that certain patients may present mutations in EVs that were previously undetectable in tissue, suggesting that screening of EVs-derived nucleic acids by ddPCR may provide clues on the occurrence of BRAF / MEK inhibitor therapy resistance before radiological evaluation of the tumor [39].

#### 4. Conclusions

The clinical management of CM has evolved in recent years towards a more personalized approach that requires an accurate assessment of the molecular alterations associated with tumor growth and evolution, as well as resistance to therapy [41]. Suitable for both archived and LB samples, ddPCR can be used for numerous omics evaluations, including absolute allele quantification, CNVs analysis, rare mutations, DNA methylation detection, transcriptomic evaluations (mRNA, miRNA), and genomic rearrangements, being of great interest to CM research [21]. Given that multi-omics integration is essential for improving the performance of prognostic and predictive models in CM, the validation of novel multi-omics biomarkers through ddPCR can play an important role in this

direction [6]. As ddPCR offers outstanding opportunities for CM biomarker research, it is expected that this technology to open new avenues for precision medicine in this difficult to treat skin tumor.

## References

1. Grzywa, T.M.; Paskal, W.; Włodarski, P.K. Intratumor and Intertumor Heterogeneity in Melanoma. *Transl. Oncol.* **2017**, *10*, 956–975, doi:10.1016/j.tranon.2017.09.007.
2. Wilson, M.A.; Nathanson, K.L. Molecular testing in melanoma. *Cancer J.* **2012**, *18*, 117–123, doi:10.1097/PPO.0b013e31824f11bf.
3. Jiang, Y.; Shi, X.; Zhao, Q.; Krauthammer, M.; Rothberg, B.E.G.; Ma, S. Integrated analysis of multidimensional omics data on cutaneous melanoma prognosis. *Genomics* **2016**, *107*, 223–230, doi:10.1016/j.ygeno.2016.04.005.
4. Olivier, M.; Asmis, R.; Hawkins, G.A.; Howard, T.D.; Cox, L.A. The Need for Multi-Omics Biomarker Signatures in Precision Medicine. *Int. J. Mol. Sci.* **2019**, *20*, doi:10.3390/ijms20194781.
5. Zhao, Q.; Shi, X.; Xie, Y.; Huang, J.; Shia, B.; Ma, S. Combining multidimensional genomic measurements for predicting cancer prognosis: observations from TCGA. *Brief. Bioinform.* **2015**, *16*, 291–303, doi:10.1093/bib/bbu003.
6. Jayawardana, K.; Schramm, S.-J.; Haydu, L.; Thompson, J.F.; Scolyer, R.A.; Mann, G.J.; Müller, S.; Yang, J.Y.H. Determination of prognosis in metastatic melanoma through integration of clinico-pathologic, mutation, mRNA, microRNA, and protein information. *Int. J. cancer* **2015**, *136*, 863–874, doi:10.1002/ijc.29047.
7. Yoshikawa, S.; Kiyohara, Y.; Otsuka, M.; Kondou, R.; Nonomura, C.; Miyata, H.; Iizuka, A.; Ohshima, K.; Urakami, K.; Nagashima, T.; et al. Multi-omics Profiling of Patients with Melanoma Treated with Nivolumab in Project HOPE. *Anticancer Res.* **2017**, *37*, 1321–1328, doi:10.21873/anticancer.11450.
8. Dumitru, C.; Constantin, C.; Popp, C.; Cioplea, M.; Zurac, S.; Vassu, T.; Neagu, M. Innovative array-based assay for omics pattern in melanoma. *J. Immunoassay Immunochem.* **2017**, *38*, 343–354, doi:10.1080/15321819.2017.1340898.
9. Huerta, M.; Roselló, S.; Sabater, L.; Ferrer, A.; Tarazona, N.; Roda, D.; Gambardella, V.; Alfaro-Cervelló, C.; Garcés-Albir, M.; Cervantes, A.; et al. Circulating Tumor DNA Detection by Digital-Droplet PCR in Pancreatic Ductal Adenocarcinoma: A Systematic Review. *Cancers (Basel)*. **2021**, *13*, doi:10.3390/cancers13050994.
10. Ding, P.N.; Becker, T.; Bray, V.; Chua, W.; Ma, Y.; Xu, B.; Lynch, D.; de Souza, P.; Roberts, T. Plasma next generation sequencing and droplet digital PCR-based detection of epidermal growth factor receptor (EGFR) mutations in patients with advanced lung cancer treated with subsequent-line osimertinib. *Thorac. Cancer* **2019**, *10*, 1879–1884, doi:10.1111/1759-7714.13154.
11. Mao, X.; Liu, C.; Tong, H.; Chen, Y.; Liu, K. Principles of digital PCR and its applications in current obstetrical and gynecological diseases. *Am. J. Transl. Res.* **2019**, *11*, 7209–7222.
12. Manoj, P. Droplet digital PCR technology promises new applications and research areas. *Mitochondrial DNA* **2016**, *27*, 742–746, doi:10.3109/19401736.2014.913168.
13. Tong, Y.; Shen, S.; Jiang, H.; Chen, Z. Application of Digital PCR in Detecting Human Diseases Associated Gene Mutation. *Cell. Physiol. Biochem. Int. J. Exp. Cell. Physiol. Biochem. Pharmacol.* **2017**, *43*, 1718–1730, doi:10.1159/000484035.
14. McEvoy, A.C.; Wood, B.A.; Ardakani, N.M.; Pereira, M.R.; Pearce, R.; Cowell, L.; Robinson, C.; Grieco-Iacopetta, F.; Spicer, A.J.; Amanuel, B.; et al. Droplet Digital PCR for Mutation Detection in Formalin-Fixed, Paraffin-Embedded Melanoma Tissues: A Comparison with Sanger Sequencing and

- Pyrosequencing. *J. Mol. Diagnostics* **2018**, *20*, 240–252, doi:10.1016/j.jmoldx.2017.11.009.
15. Cirillo, P.D.R.; Margiotti, K.; Mesoraca, A.; Giorlandino, C. Quantification of circulating microRNAs by droplet digital PCR for cancer detection. *BMC Res. Notes* **2020**, *13*, 351, doi:10.1186/s13104-020-05190-3.
  16. Demaree, B.; Weisgerber, D.; Dolatmoradi, A.; Hatori, M.; Abate, A.R. Direct quantification of EGFR variant allele frequency in cell-free DNA using a microfluidic-free digital droplet PCR assay. *Methods Cell Biol.* **2018**, *148*, 119–131, doi:10.1016/bs.mcb.2018.10.002.
  17. Preobrazhenskaya, E. V; Bizin, I. V; Kuligina, E.S.; Shleykina, A.Y.; Suspitsin, E.N.; Zaytseva, O.A.; Anisimova, E.I.; Laptiev, S.A.; Gorodnova, T. V; Belyaev, A.M.; et al. Detection of BRCA1 gross rearrangements by droplet digital PCR. *Breast Cancer Res. Treat.* **2017**, *165*, 765–770, doi:10.1007/s10549-017-4357-7.
  18. Diefenbach, R.; Lee, J.; Chandler, D.; Wang, Y.; Pflueger, C.; Long, G.; Scolyer, R.; Carlino, M.; Menzies, A.; Kefford, R.; et al. Hypermethylation of Circulating Free DNA in Cutaneous Melanoma. *Appl. Sci.* **2019**, *9*, 5074, doi:10.3390/app9235074.
  19. Reinert, T.; Schøler, L. V; Thomsen, R.; Tobiasen, H.; Vang, S.; Nordentoft, I.; Lamy, P.; Kannerup, A.-S.; Mortensen, F. V; Stribolt, K.; et al. Analysis of circulating tumour DNA to monitor disease burden following colorectal cancer surgery. *Gut* **2016**, *65*, 625–634, doi:10.1136/gutjnl-2014-308859.
  20. Busser, B.; Lupo, J.; Sancey, L.; Mouret, S.; Faure, P.; Plumas, J.; Chaperot, L.; Leccia, M.T.; Coll, J.L.; Hurbin, A.; et al. Plasma Circulating Tumor DNA Levels for the Monitoring of Melanoma Patients: Landscape of Available Technologies and Clinical Applications. *Biomed Res. Int.* **2017**, *2017*, 5986129, doi:10.1155/2017/5986129.
  21. Olmedillas-López, S.; García-Arranz, M.; García-Olmo, D. Current and Emerging Applications of Droplet Digital PCR in Oncology. *Mol. Diagn. Ther.* **2017**, *21*, 493–510, doi:10.1007/s40291-017-0278-8.
  22. Yu, W.; Hurley, J.; Roberts, D.; Chakraborty, S.K.; Enderle, D.; Noerholm, M.; Breakefield, X.O.; Skog, J.K. Exosome-based liquid biopsies in cancer: opportunities and challenges. *Ann. Oncol.* **2021**, *32*, 466–477.
  23. Malicherova, B.; Burjanivova, T.; Grendar, M.; Minarikova, E.; Bobrovskaya, M.; Vanova, B.; Jasek, K.; Jezkova, E.; Kapinova, A.; Antosova, M.; et al. Droplet digital PCR for detection of BRAF V600E mutation in formalin-fixed, paraffin-embedded melanoma tissues: a comparison with Cobas® 4800, Sanger sequencing, and allele-specific PCR. *Am. J. Transl. Res.* **2018**, *10*, 3773–3781.
  24. Salgado, C.; Roelse, C.; Nell, R.; Gruis, N.; van Doorn, R.; van der Velden, P. Interplay between TERT promoter mutations and methylation culminates in chromatin accessibility and TERT expression. *PLoS One* **2020**, *15*, e0231418, doi:10.1371/journal.pone.0231418.
  25. Burjanivova, T.; Malicherova, B.; Grendar, M.; Minarikova, E.; Dusenka, R.; Vanova, B.; Bobrovskaya, M.; Pecova, T.; Homola, I.; Lasabova, Z.; et al. Detection of BRAFV600E Mutation in Melanoma Patients by Digital PCR of Circulating DNA. *Genet. Test. Mol. Biomarkers* **2019**, *23*, 241–245, doi:10.1089/gtmb.2018.0193.
  26. Huang, S.K.; Hoon, D.S.B. Liquid biopsy utility for the surveillance of cutaneous malignant melanoma patients. *Mol. Oncol.* **2016**, *10*, 450–463.
  27. Shinozaki, M.; O'Day, S.J.; Kitago, M.; Amersi, F.; Kuo, C.; Kim, J.; Wang, H.-J.; Hoon, D.S.B. Utility of circulating B-RAF DNA mutation in serum for monitoring melanoma patients receiving biochemotherapy. *Clin. cancer Res. an Off. J. Am. Assoc. Cancer Res.* **2007**, *13*, 2068–2074, doi:10.1158/1078-0432.CCR-06-2120.
  28. Tsao, S.C.-H.; Weiss, J.; Hudson, C.; Christophi, C.; Cebon, J.; Behren, A.; Dobrovic, A. Monitoring

- response to therapy in melanoma by quantifying circulating tumour DNA with droplet digital PCR for BRAF and NRAS mutations. *Sci. Rep.* **2015**, *5*, 11198, doi:10.1038/srep11198.
29. Santiago-Walker, A.; Gagnon, R.; Mazumdar, J.; Casey, M.; Long, G. V.; Schandendorf, D.; Flaherty, K.; Kefford, R.; Hauschild, A.; Hwu, P.; et al. Correlation of BRAF Mutation Status in Circulating-Free DNA and Tumor and Association with Clinical Outcome across Four BRAFi and MEKi Clinical Trials. *Clin. cancer Res. an Off. J. Am. Assoc. Cancer Res.* **2016**, *22*, 567–574, doi:10.1158/1078-0432.CCR-15-0321.
  30. Sanmamed, M.F.; Fernández-Landázuri, S.; Rodríguez, C.; Zárata, R.; Lozano, M.D.; Zubiri, L.; Perez-Gracia, J.L.; Martín-Algarra, S.; González, A. Quantitative cell-free circulating BRAFV600E mutation analysis by use of droplet digital PCR in the follow-up of patients with melanoma being treated with BRAF inhibitors. *Clin. Chem.* **2015**, *61*, 297–304, doi:10.1373/clinchem.2014.230235.
  31. Seremet, T.; Jansen, Y.; Planken, S.; Njimi, H.; Delaunoy, M.; El Housni, H.; Awada, G.; Schwarze, J.K.; Keyaerts, M.; Everaert, H.; et al. Undetectable circulating tumor DNA (ctDNA) levels correlate with favorable outcome in metastatic melanoma patients treated with anti-PD1 therapy. *J. Transl. Med.* **2019**, *17*, 303, doi:10.1186/s12967-019-2051-8.
  32. Lee, J.H.; Long, G. V.; Boyd, S.; Lo, S.; Menzies, A.M.; Tembe, V.; Guminski, A.; Jakrot, V.; Scolyer, R.A.; Mann, G.J.; et al. Circulating tumour DNA predicts response to anti-PD1 antibodies in metastatic melanoma. *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* **2017**, *28*, 1130–1136, doi:10.1093/annonc/mdx026.
  33. Diefenbach, R.J.; Lee, J.H.; Rizos, H. Methylated circulating tumor DNA as a biomarker in cutaneous melanoma. *Melanoma Manag.* **2020**, *7*, MMT46.
  34. Micevic, G.; Theodosakis, N.; Bosenberg, M. Aberrant DNA methylation in melanoma: biomarker and therapeutic opportunities. *Clin. Epigenetics* **2017**, *9*, 34, doi:10.1186/s13148-017-0332-8.
  35. Neagu, M.; Constantin, C.; Cretoiu, S.M.; Zurac, S. miRNAs in the Diagnosis and Prognosis of Skin Cancer. *Front. cell Dev. Biol.* **2020**, *8*, 71, doi:10.3389/fcell.2020.00071.
  36. Poenitzsch Strong, A.M.; Setaluri, V.; Spiegelman, V.S. MicroRNA-340 as a modulator of RAS-RAF-MAPK signaling in melanoma. *Arch. Biochem. Biophys.* **2014**, *563*, 118–124, doi:10.1016/j.abb.2014.07.012.
  37. Sato, H.; Soh, J.; Aoe, K.; Fujimoto, N.; Tanaka, S.; Namba, K.; Torigoe, H.; Shien, K.; Yamamoto, H.; Tomida, S.; et al. Droplet digital PCR as a novel system for the detection of microRNA-34b/c methylation in circulating DNA in malignant pleural mesothelioma. *Int. J. Oncol.* **2019**, *54*, 2139–2148, doi:10.3892/ijo.2019.4768.
  38. Hong, X.; Sullivan, R.J.; Kalinich, M.; Kwan, T.T.; Giobbie-Hurder, A.; Pan, S.; LiCausi, J.A.; Milner, J.D.; Nieman, L.T.; Wittner, B.S.; et al. Molecular signatures of circulating melanoma cells for monitoring early response to immune checkpoint therapy. *Proc. Natl. Acad. Sci. U. S. A.* **2018**, *115*, 2467–2472, doi:10.1073/pnas.1719264115.
  39. Yap, S.A.; Münster-Wandowski, A.; Nonnenmacher, A.; Keilholz, U.; Liebs, S. Analysis of cancer-related mutations in extracellular vesicles RNA by Droplet Digital™ PCR. *Biotechniques* **2020**, *69*, 99–107, doi:10.2144/btn-2020-0028.
  40. Zocco, D.; Bernardi, S.; Novelli, M.; Astrua, C.; Fava, P.; Zarovni, N.; Carpi, F.M.; Bianciardi, L.; Malavenda, O.; Quaglino, P.; et al. Isolation of extracellular vesicles improves the detection of mutant DNA from plasma of metastatic melanoma patients. *Sci. Rep.* **2020**, *10*, 15745, doi:10.1038/s41598-020-72834-6.
  41. Clark, M.E.; Rizos, H.; Pereira, M.R.; McEvoy, A.C.; Marsavela, G.; Calapre, L.; Meehan, K.; Ruhen, O.; Khattak, M.A.; Meniawy, T.M.; et al. Detection of BRAF splicing variants in plasma-derived cell-free nucleic acids and extracellular vesicles of melanoma patients failing targeted therapy therapies.

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