

Extended Abstract

# Customized Screen-Printed Electrodes Based on Ag-Nanoseeds for Enhanced Electroanalytical Response Towards Cd(II), Pb(II) and As(V) in Aqueous Samples <sup>+</sup>

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Abstract: Electrochemical analysis based on screen-printed electrodes (SPEs) represents a great al-22 ternative to conventional analytical methods such as ICP-MS or LC-MS due to their portability, sen-23 sitivity, selectivity, and cost-effectiveness. In addition, the functionalization of SPEs with nano-24 materials has been reported to provide an enhanced analytical performance. In this regard, silver 25 nanoparticles (AgNPs) were synthesized and appropriately characterized, showing spherical silver 26 nanoseeds (Ag-NS) with a diameter of  $12.20 \pm 0.04$  nm. Using drop casting methodology, the syn-27 thesized AgNPs were used to modify screen-printed carbon nanofiber electrodes (SPCNFEs). Ag-28 NS deposition onto the electrode surface was confirmed by scanning electron microscopy (SEM). 29 Furthermore, the analytical response of the modified electrodes (Ag-NS-SPCNFE) was evaluated 30 for the determination of trace Pb(II), Cd(II), and As(V) using differential pulse anodic stripping 31 voltammetry (DPASV), obtaining detection limits of 3.3, 3.7, and 2.6 µg L<sup>-1</sup>, for Pb(II), Cd(II) and 32 As(V), respectively. Finally, Ag-NS-SPCNFE was tested towards the determination of As(V) in a 33 spiked tap water sample, showing a good agreement with concentrations determined by ICP-MS. 34

**Keywords:** screen-printed electrodes; Ag nanoparticles; anodic stripping voltammetry; lead determination; cadmium determination; arsenic determination 36

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# 1. Introduction

Water contamination caused by heavy metal ions (HMIs) is a concerning issue due to their high toxicity, non-biodegradability, bioaccumulation, and adverse health effects in humans [1]. In particular, for As, Cd, and Pb, the World Health Organization (WHO) has established the maximum allowed concentration in drinking water as  $10 \ \mu g \ L^{-1}$ ,  $3 \ \mu g \ L^{-1}$ , and  $10 \ \mu g \ L^{-1}$ , respectively [2]. The determination of these low concentration levels requires very sensitive analytical techniques such as flameless atomic adsorption spectroscopy (FAAS) [3], inductively coupled plasma mass spectrometry (ICP-MS) [4,5], and

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hydride generation atomic fluorescence spectrometry (HG-AFS) [6]. However, these ana-46lytical techniques require expensive equipment and highly-trained personnel, long analy-47sis time, and high operating costs.48

In contrast, electrochemical techniques and anodic stripping voltammetry (ASV) al-49 low the relatively fast determination of trace HMIs with easy-handling and low-cost 50 equipment [7,8]. Stripping techniques present low detection limits and high sensitivity, 51 which can be further improved by modifying the working electrode with nanomaterials 52 [9–11]. In par-ticular, metallic nanoparticles (MNPs) can reduce the electron transfer re-53 sistance at the electrode surface, decreasing the electron transfer limited process and con-54 sequently catalyzing the electrode's response at low analyte concentrations [12–14]. In-55 deed, MNPs-modified sensors have been reported to allow the detection of arsenic [15,16], 56 lead [17,18], and cadmium [19,20] at the level of a few  $\mu$ g L<sup>-1</sup>, fulfilling the WHO guide-57 lines for drinking-water quality [2]. These sensors were based on screen-printing technol-58 ogy, offering significant advantages over conventional voltammetric sensors such as low-59 character, portability, and commercial availability [1,21]. Thus, in cost, disposable 60 this work, the voltammetric determination of HMIs based on the use of carbon-nanofiber-61 based screen-printed electrodes (SPCNFEs) modified with silver nanoparticles (Ag-NPs) 62 is proposed. Ag-NPs were synthesized in the shape of silver nanoseeds (Ag-NS), and the 63 resulting modified electrodes were microscopically and analytically characterized for the 64 determination of As(V), Pb(II), and Cd(II) by means of differential pulse anodic stripping 65 voltammetry (DPASV). In addition, the applicability to real sample analysis was demon-66 strated through the direct determination of As(V) in spiked tap water samples. 67

### 2. Materials and Methods

# 2.1. Apparatus

DPASV measurements were carried out with either a Multi Autolab/M204 Modular 70 Multi Potentiostat/Galvanostat or an Autolab PGSTAT204, attached to a Metrohm 663 VA 71 Stand, as well as a personal computer with NOVA 2.1 software package to control the 72 potentiostat and perform the required data treatment. All the electrochemical instrumentations and software were acquired from Metrohm (Herisau, Switzerland). 74

A Crison Basic 20 pH meter (Hach Lange Spain, L'Hospitalet de Llobregat, Spain) 75 was used for pH measurements. 76

Ag-NS and SPCNFE modified with Ag-NS electrodes were characterized using a77JEM-2010 transmission electron microscope (TEM) from JEOL (Tokyo, Japan) and a Gem-78ini scanning electron microscope (SEM) from ZEISS® (Jena, Germany), respectively. Size79distribution histograms were calculated using the Image-J version 1.51m software by the80National Institute of Health (NIH, Bethesda, MD, USA).81

ICP-MS measurements were carried out by means of inductively coupled plasma mass spectrometer model 7800 by Agilent Technologies (Santa Clara, CA, USA).

Commercial SPCNFEs, including working (4 mm disk), counter, and reference electrodes, were purchased from Dropsens (Llanera, Spain).

#### 2.2. Preparation of Working Electrodes

The working electrode (WE) was a SPCNFE modified with silver nanoseeds (Ag-NS-SPCNFE). Ag-NS were first synthesized following a seed-mediated methodology asdescribed elsewhere [11,22]. Then, SPCNFEs were modified by drop casting, dropping 40  $\mu$ L of Ag-NS onto the working electrode, and evaporating the solvent at 50 °C for 30 min. 90

#### 2.3. Electrochemical Measurements

DPASV measurements of Pb(II) and Cd(II) were carried out at a deposition potential 92 (Ed) of -1.4 V, applied under stirring conditions during a deposition time (td) of 180 s in 93 0.1 mol L<sup>-1</sup> acetate buffer (pH 4.5) and scanning the potential from -1.4 to 0.0 V. For As(V) 94

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determination, the experimental conditions used were Ed of -1.3 V and td of 120 s in 0.01 95 mol  $L^{-1}$  HCl pH 2 with a potential scan from -1.3 to -0.65 V. 96

A step potential of 5 mV, a pulse time of 50 ms, and a pulse amplitude of 50 mV were 97 employed in all cases. All experiments were performed at room temperature ( $22 \pm 1$ 98 •C) and without oxygen removal. 99

For real sample analysis, tap water samples were collected from the local water dis-100 tribution network in Barcelona (Spain) and spiked with 20 µg L<sup>-1</sup> of As(V). Prior to elec-101 trochemical analysis, water samples were diluted and acidified with 0.01 mol L<sup>-1</sup> of HCl 102 (pH 2.0), resulting in a final concentration of 10  $\mu$ g L<sup>-1</sup> of As(V). Sample analysis was car-103 ried out by means of the standard addition method, performing four successive As(V) 104 additions from a standard solution of 1 mg L<sup>-1</sup>. DPASV measurements were recorded un-105 der the above-mentioned electrochemical conditions. 106

# 3. Results

# 3.1. Microscopic Ccharacterization

0 nm

Ag-NS synthesis was microscopically confirmed by both SEM (Figure 1a) and TEM 109 (Figure 1b). As it can be deduced from the TEM image, most Ag-NS presented a spherical 110 shape. On the other hand, SEM images were used to calculate the corresponding size dis-111 tribution histogram (Figure 1c), which was computed from 400 Ag-NSs. The obtained re-112 sults show that the synthesized Ag-NS presented an average diameter of  $12.2 \pm 0.4$  nm. 113 These structures are in good agreement with the reported shapes of Ag-NPs [23,24]. 114

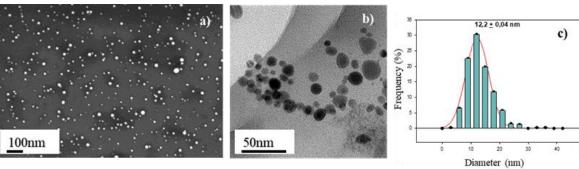
00nm 50nm 20 Diameter (nm)

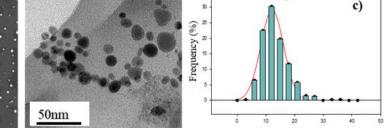
Figure 1. Microscopic characterization of Ag-Nanoseeds (a) SEM micrograph, (b) TEM micrograph and (c) corresponding 116 size distribution histogram [14]. 117

SEM micrographs were also obtained for a bare SPCNFE (Figure 2a) and an Ag-NS-118 SPCNFE (Figure 2b) to assess the modification of SPCNFEs by drop casting. Compared to 119 the non-modified carbon nanofiber surface of the bare electrode, Ag-NS can be spotted as 120 white dots deposited onto the carbon nanofibers in the modified electrode (Ag-NS-121 SPCNFE), thus confirming the successful modification of the working electrode. 122

Figure 2. SEM micrographs for (a) Bare SPCNFE, and (b) Ag-NS-SPCNFE modified using the drop casting methodology [11,14]. 125

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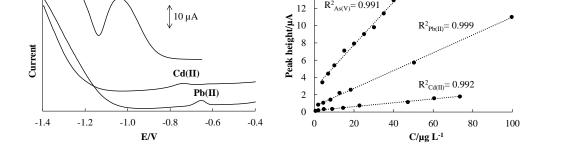
As(V)

DPASV measurements were carried out in solutions containing either Pb(II), Cd(II), 127 or As(V). Well-defined voltammetric peaks were obtained in all cases, with peak poten-128 tials of ca. -0.65 V, -0.75 V, and -1.0 V for Pb(II), Cd(II), and As(V), respectively (see 129 figure 3a). 130

Individual calibration curves of Pb(II), Cd(II), and As(V) were obtained by DPASV by increasing metal ion concentration in the ranges 1.9 to 150.0  $\mu$ g L<sup>-1</sup>, 0.6 to 120.6  $\mu$ g L<sup>-1</sup>, 132 and 1.0 to 50.1  $\mu$ g L<sup>-1</sup>, respectively. The obtained data were used to calculate the corre-133 sponding analytical parameters (i.e., sensitivity, limit of detection (LOD), limit of quanti-134 fycation (LOQ), and linear range), which are displayed in Table 1.

From this data, LODs and LOQs were calculated by using the Miller and Miller pro-136 cedure [25,26]. 137

b)



a)

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Figure 3. (a) DPASV voltammograms of Pb(II), Cd(II), and As(V) at 25 µg L<sup>-1</sup> and (b) their calibration plots at the previously mentioned conditions using Ag-NS-SPCNFE.

Table 1. Calibration data for the individual determination of Pb(II), Cd(II), and As(V) using Ag-NS-SPCNFE and the corresponding buffer and DPASV parameters (see the experimental section 142 for more details). 143

Analyte	LOD (µg L-1)	Linear range (µg L-1)ª	<b>R</b> <sup>2</sup>	Sensitivity (nA µg <sup>-1</sup> L) <sup>b</sup>
Pb(II)	3.3	11.00-99.6	0.999	103 (1)
Cd(II)	3.7	12.2-73.4	0.992	22 (1)
As(V)	2.6	8.9-40.0	0.991	260 (10)

<sup>a</sup>The lowest value of the linear range corresponds to the LOQ. <sup>b</sup>The standard deviations are ex-144 pressed in parentheses 145

As shown in Table 1, good linear response between the peak heights and the concen-146 tration of the different analytes was achieved using the Ag-NS-SPCNFE. LODs were at  $\mu g$ 147 L-1 levels in all cases, and similar or even lower to other LODs reported in the literature. 148 For example, LODs of 3.30 and 4.43 µg L-1 for Pb(II) and Cd(II), respectively, were reported 149 using a graphene/polyaniline/polystyrene (G/PANI/PS) nanoporous fiber modified 150 screen-printed carbon electrode [27]. Also, the obtained LOD for As(V) is considerably 151 lower than that reported using boron-doped diamond electrodes and ASV (12 µg L<sup>-1</sup>) [28]. 152 However, it is important to mention that Ag-NS-SPCNFE for the determination of As(V)153 presented a more restricted linear range in which the highest value is limited to a lower 154concentration value (until 40.0 µg L-1) compared to the one reached by Nagaoka et al. (until 155 100 µg L-1) [28]. 156

In terms of sensitivities (nA  $\mu g^{-1}$  L), which were calculated as the slope of the calibra-157 tion curves, Ag-NS-SPCNFE exhibited higher sensitivity toward As(V) (260 nA  $\mu g^{-1}$  L). In 158 the case of Pb(II) and Cd(II), the sensitivities were significantly lower (103 and 22 nA  $\mu$ g<sup>-1</sup> 159 L, respectively). 160

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The applicability of Ag-NS-SPCNFE for real sample determination was evaluated 162 through the determination of As(V) in a spiked tap water sample. The determination of 163 As(V) was performed in triplicate by the standard addition calibration method. Repre-164 sentative voltammograms are shown in Figure 4. As it can be observed, a well-shaped 165 As(V) peak and a good correlation between peak area and concentration were acquired. 166 Sample concentration calculated by extrapolation was  $10.04 \ \mu g \ L^{-1}$  (SD:0.37  $\ \mu g \ L^{-1}$ ), which 167 is in good agreement with values obtained by ICP-MS (10.7  $\mu$ g L<sup>-1</sup>, SD:0.20  $\mu$ g  $\mu$ L<sup>-1</sup>), as an 168 analytical reference technique. These results confirm the suitability of Ag-NS-SPCNFE for 169 the analysis of real samples. 170

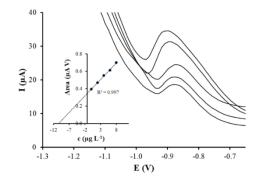


Figure 4. DPASV measurements of As(V) in a spiked tap water sample on Ag-NS-SPCNFE at pH 172 2.0 applying an Ed of -1.30 V and a td of 120 s. Inset: As(V) standard addition plot [11].

#### 4. Conclusions

In this work, a DPASV method for the determination of trace Pb(II), Cd(II), and As(V) based on the modification of SPCNFE with Ag-NS has been proposed. The Ag-NSs were 176 synthesized, microscopically characterized, and used for the modification of SPCNFEs. 177

The analytical performance of the modified electrode was evaluated for the three 178 studied analytes. It was demonstrated that Ag-NS-SPCNFE is suitable for determining 179 Pb(II), Cd(II), and As(V) at low  $\mu$ g L<sup>-1</sup> levels, showing wider linear ranges for Pb(II) and 180 Cd(II) but lower sensitivities as compared to As(V). Regarding previous studies of Pb(II), 181 Cd(II), and As(V) determination, the LODs achieved in this investigation are equal or 182 lower than other LODs previously reported. 183

The suitability of Ag-NS-SPCNFE for real sample analysis was demonstrated for the 184 determination of As(V) in spiked water samples, achieving comparable results to those 185 obtained by ICP-MS measurements with good reproducibility. 186

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