



Universidade do Minho
Escola de Engenharia



CENTRO DE CIÊNCIA E
TECNOLOGIA TÊXTIL

Wet-spun cellulose acetate/polycaprolactone fibers modified with essential oils for infection control

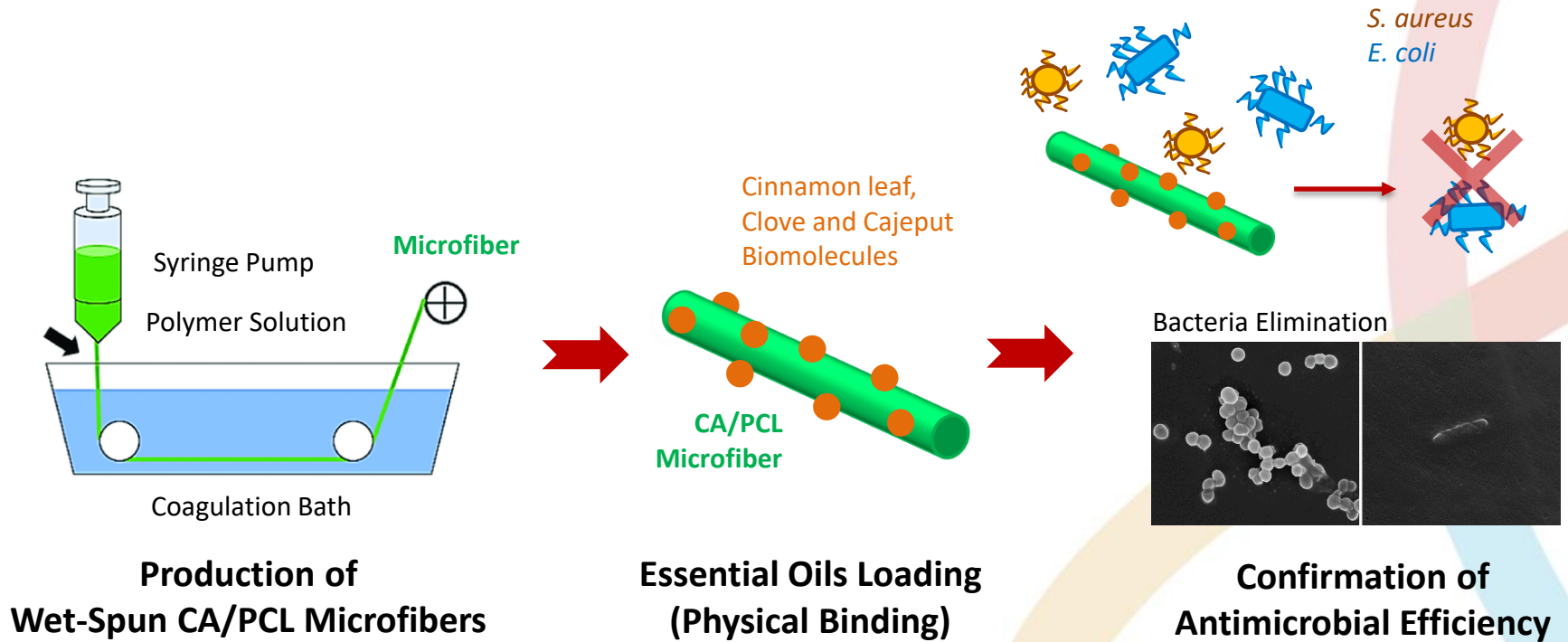
Helena P. Felgueiras*, Natália C. Homem, Ana R. M. Ribeiro, Marta A. Teixeira, Marta O. Teixeira, Joana C. Antunes, M. Teresa P. Amorim

Centre for Textile Science and Technology, University of Minho, Portugal

helena.felgueiras@2c2t.uminho.pt



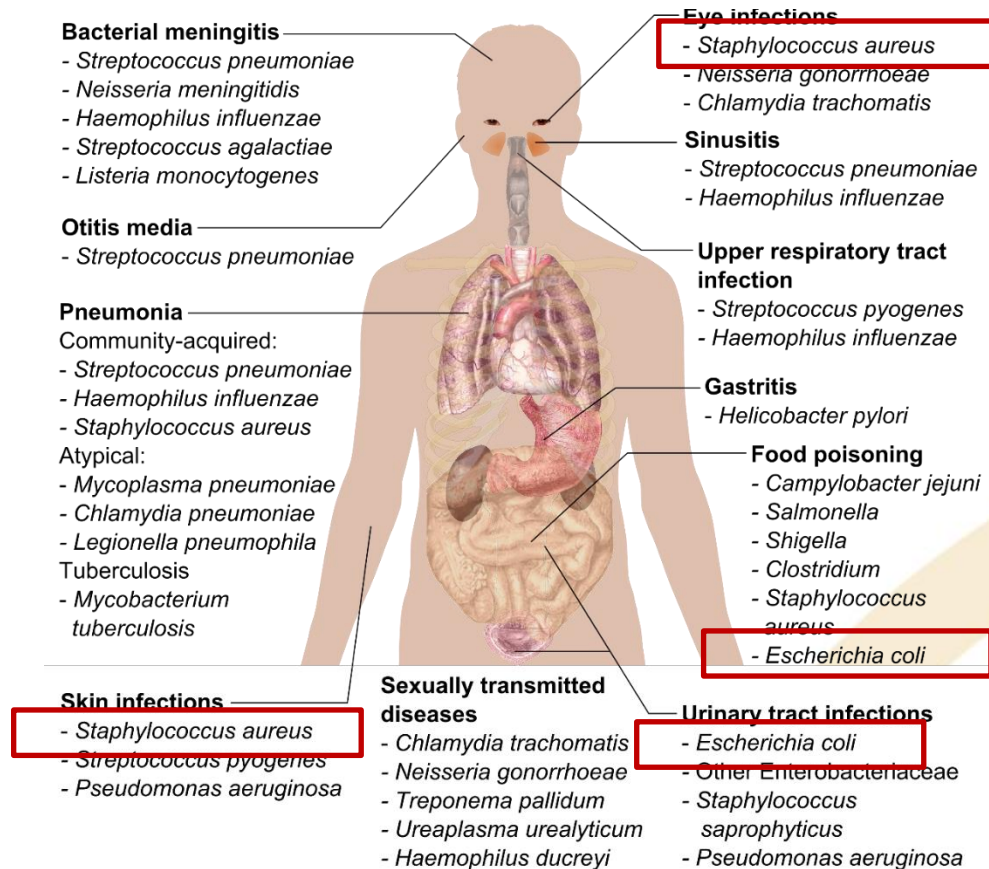
Graphical Abstract



(Representation not to scale)



Bacterial Infections



Recent projections indicate that bacterial infections may be the cause of approximately **10 million annual deaths worldwide by 2050**



Introduction

Conventional Treatments vs EOs

Antibiotics/Antiseptic Agents



Target bacterial functions, growth processes or the bacterial cell wall - **bactericidal** activities.



Rising of antibiotic resistant pathogens



Inefficient

Essential Oils (EOs)



EOs are produced by more than **17,500** species of plants



Volatile biomolecules endowed with **antimicrobial and regenerative potential**



Alternative



Introduction

EOs Drawbacks

- cytotoxic at increased concentrations, which prevents systemic delivery;
- present low resistance to degradation by external factors (e.g. temperature, light, moisture);
- highly volatile in their free, unloaded form.



Goal

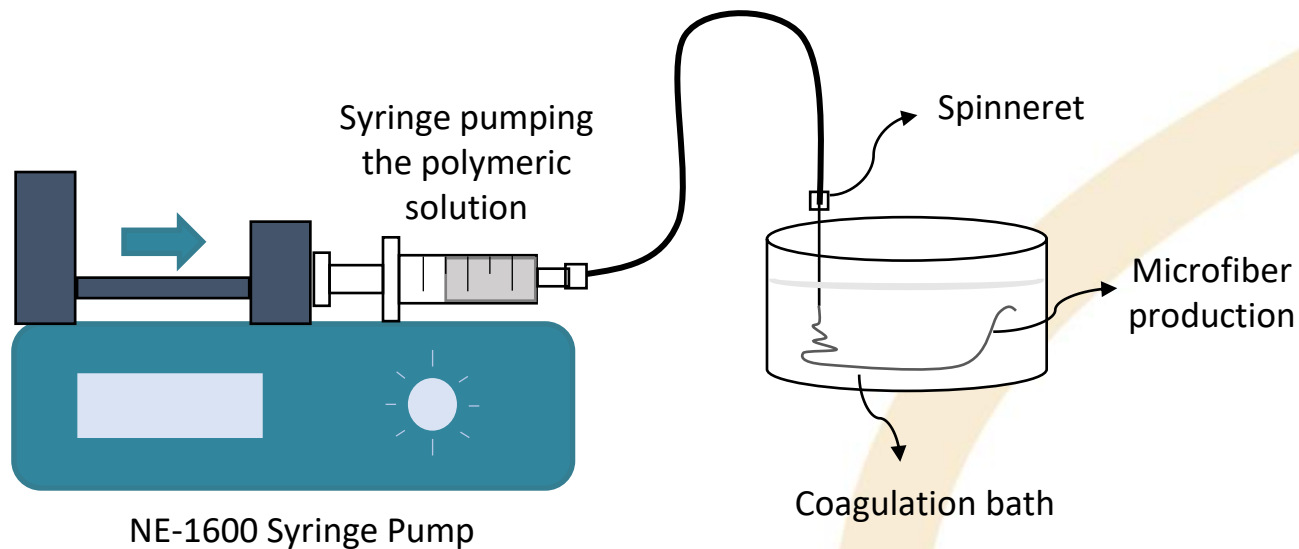
Engineer a biodegradable microfibrous target-delivery platform for EOs, that overcomes these biomolecules limitations for applications in infection control.



Methods

Wet-Spinning

Non-solvent induced phase inversion approach that allows the production of continuous polymeric **microfibers, with a uniform morphology**, by injecting a polymer solution into a non-solvent coagulation bath that prompts the solidification of the extruded material.

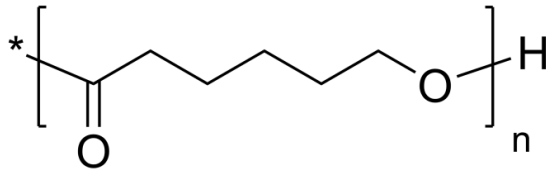


Methods

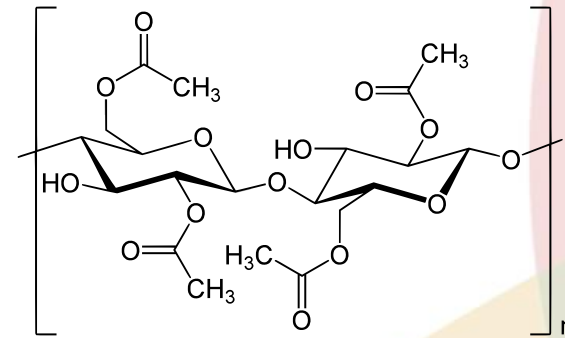
Microfiber Production

Biodegradable Polymers:

Polycaprolactone (PCL)



Cellulose Acetate (CA)



Polymeric solution preparation:

Solvents – acetic acid and acetone

Polymer ratio – 3:1 CA/PCL (10/14 wt%)

Solubilization conditions – 1 h at 75 °C and 200 rpm

Wet-spinning processing conditions:

Flow Rate – 0.5 mL/h

Needle Gauge – 18

Coagulation bath – Ethanol

Temperature of extrusion – 21 to 22 °C



Results & Discussion

EOs Minimum Inhibitory Concentration

20 EOs with antimicrobial potential were examined for their minimum inhibitory concentrations (MICs) against the **Gram-positive *Staphylococcus aureus*** and the **Gram-negative *Escherichia coli*** bacteria, at initial concentration of 1×10^7 CFUs/mL

Most effective:

Cinnamon leaf oil – CLO

Clove oil – CO

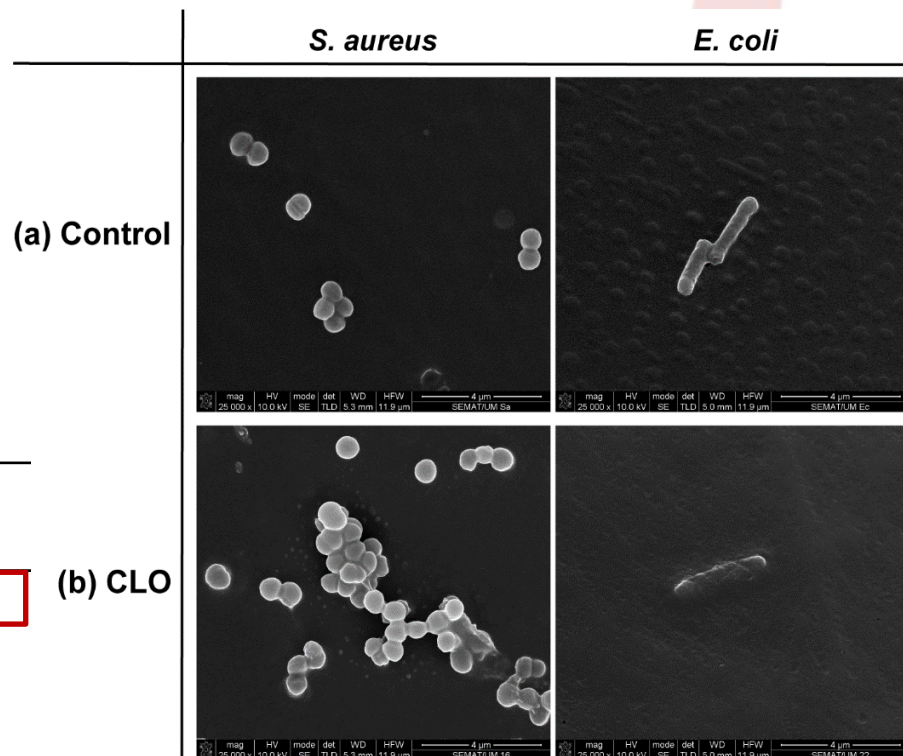
Cajeput oil – CJO

Control Antibiotic:

Ampicillin - A

Antimicrobial Agents	MICs (mg/mL)	
	<i>S. aureus</i>	<i>E. coli</i>
CLO	0.82	0.82
CO	0.83	0.83
CJO	22.38	11.19
A	0.03	0.03

*SD < ± 0.5 mg/mL



Results & Discussion

Loading Efficiency

EOs Incorporation:

Substrate – CA/PCL microfibers

Solvent – Ethanol

EOs concentration – 2 x MIC value

Conditions – 72 h at RT and 200 rpm, protected from light

Loading Efficiency:

(mapped by UV-vis spectroscopy at 280 nm)

Antimicrobial Agents	Loading (MIC %, SD < \pm 3.0%)	Concentration (mg/mL)
CLO	14.42	0.12
CO	66.08	0.55
CJO	76.48	17.12
A*	106.37	0.03

*Ampicillin was used as control to determine the maximum period for immobilization.



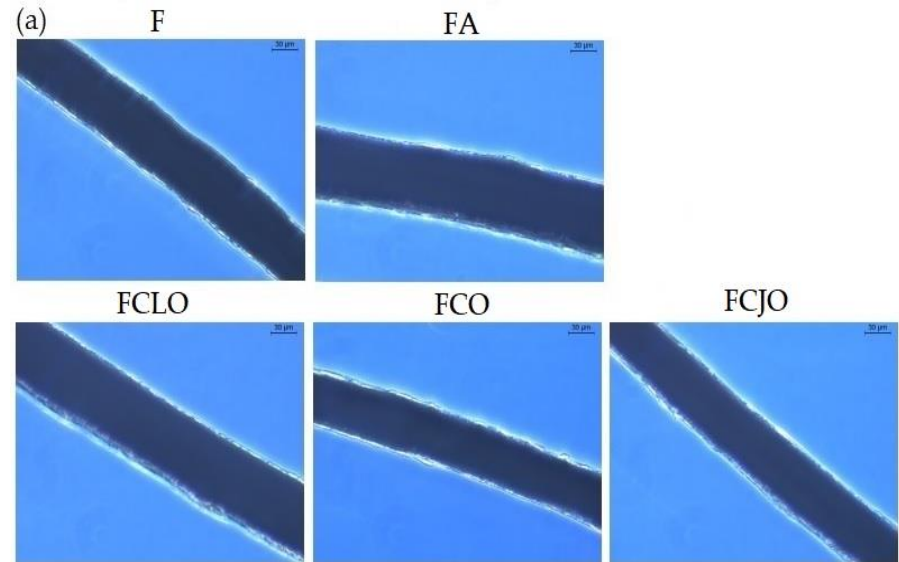
Results & Discussion

Fiber Morphology

Microfibers Observation:

Brightfield microscopy
40x Magnification (30 μm scale bar)

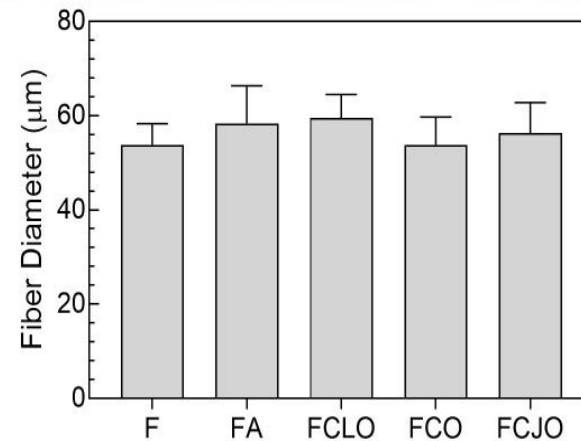
Fibers presented a **uniform, homogeneous morphology**, free from defects.



Fibers Diameters:

Averaged from 40 measurements

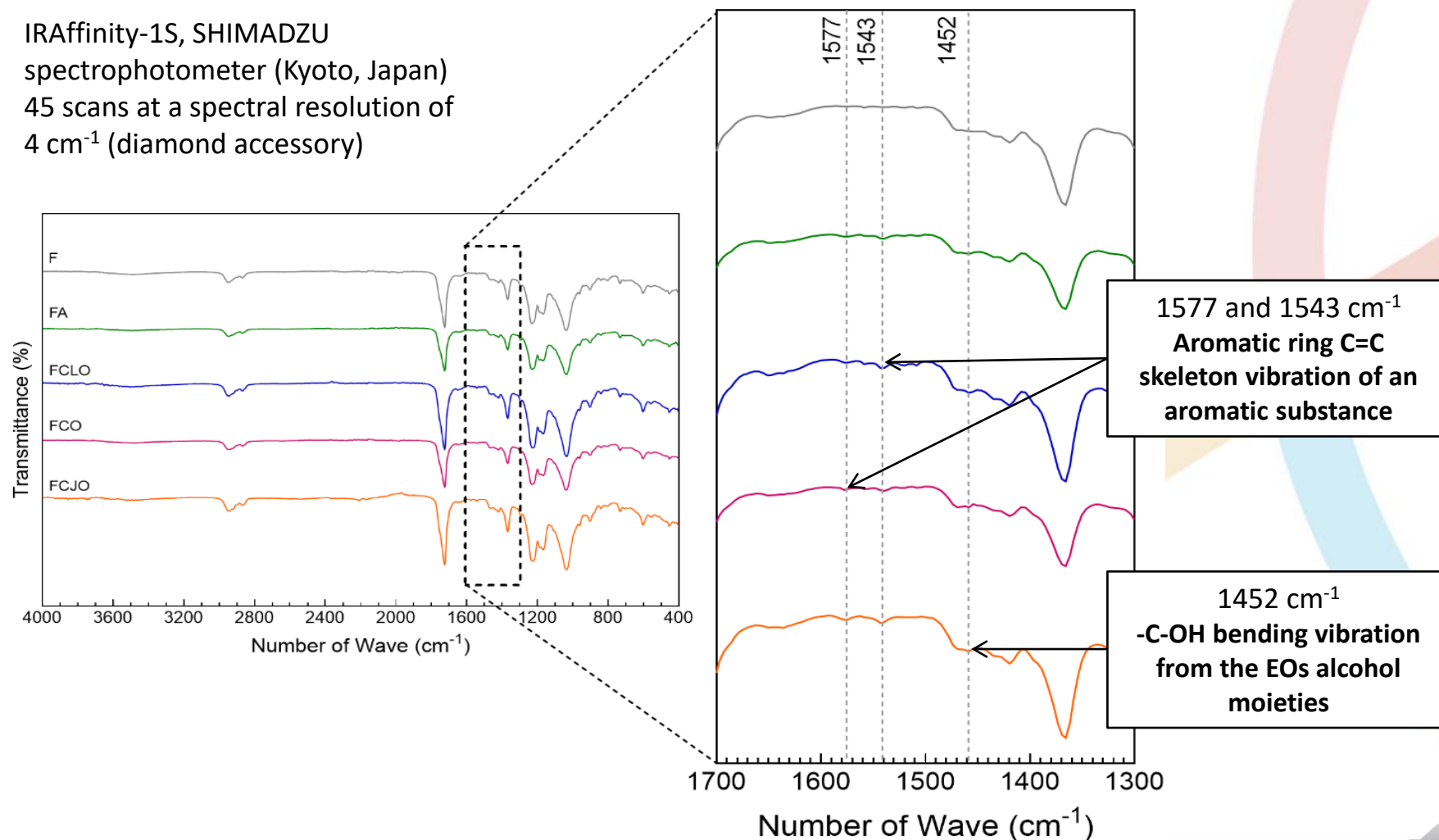
Diameters ranged between **54-59 μm** .



Results & Discussion

Chemical Characterization

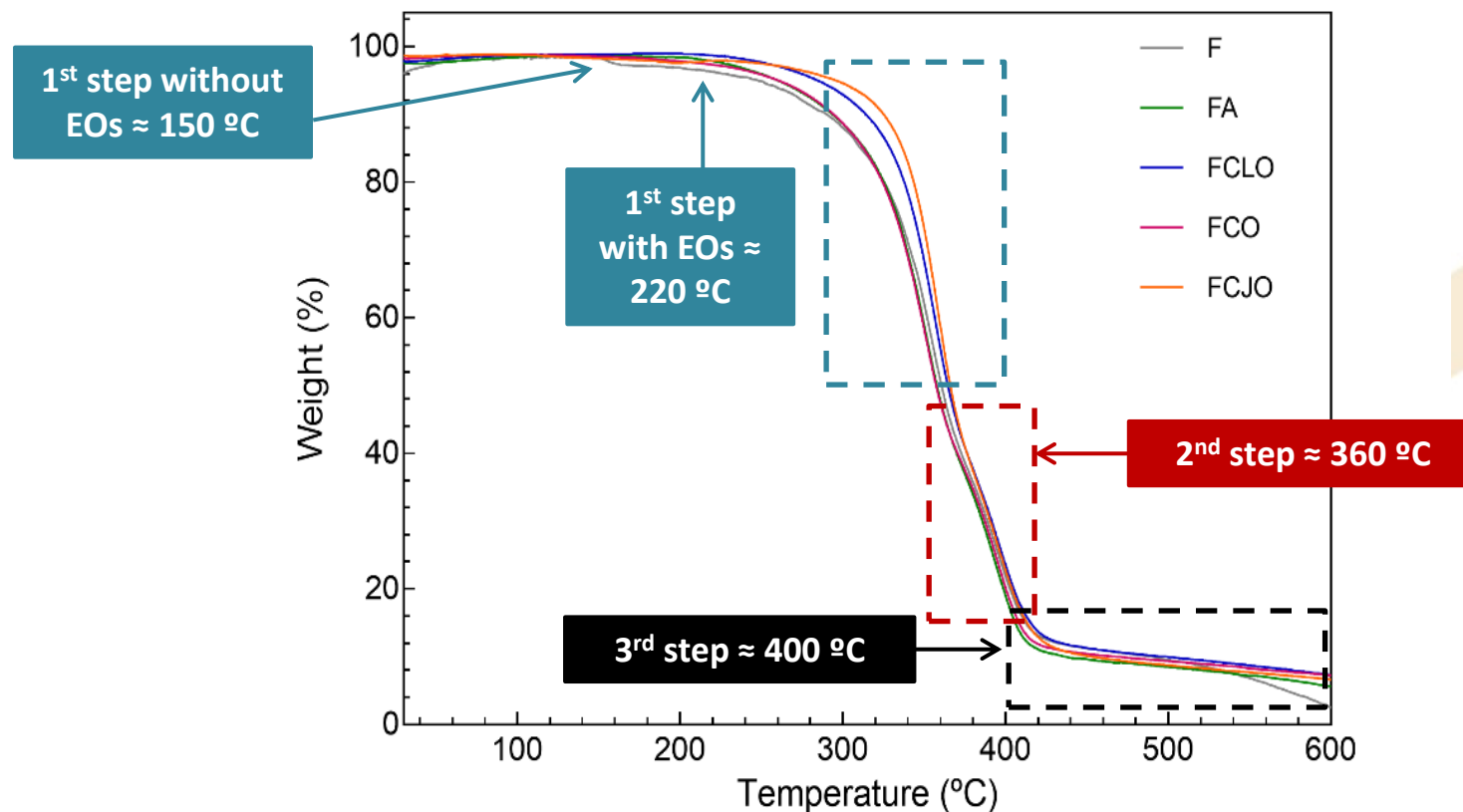
IRAffinity-1S, SHIMADZU
spectrophotometer (Kyoto, Japan)
45 scans at a spectral resolution of
 4 cm^{-1} (diamond accessory)



Results & Discussion

Thermal Stability

STA 7200 Hitachi® (Fukuoka, Japan) with platinum pan
N₂ atmosphere, flow rate of 200 mL/min and T rise of 20°C/min



Results & Discussion: Antimicrobial Action

Time Kill Kinetics

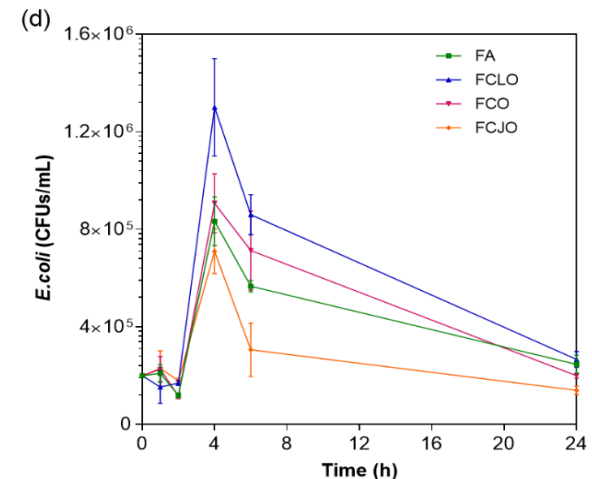
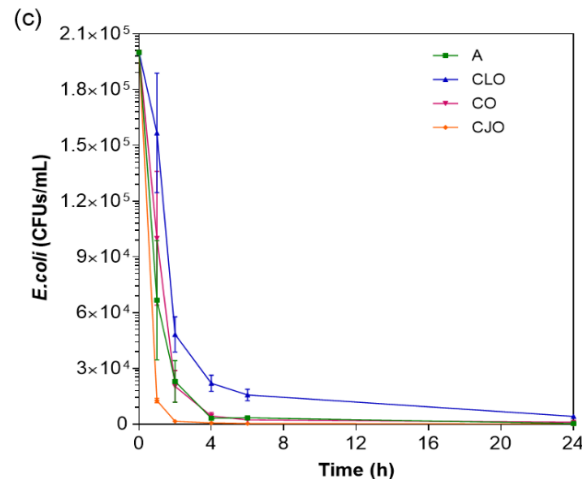
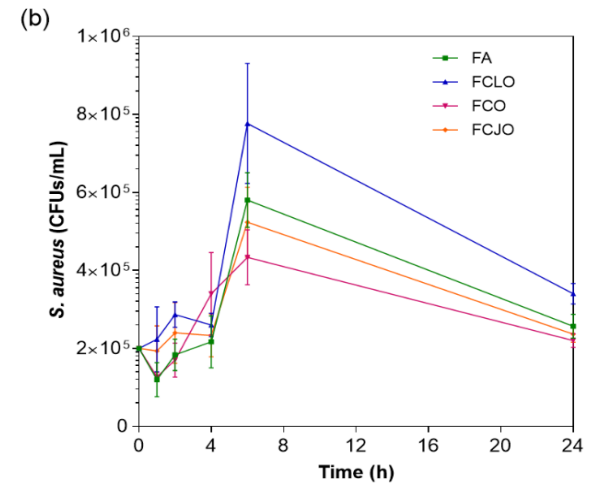
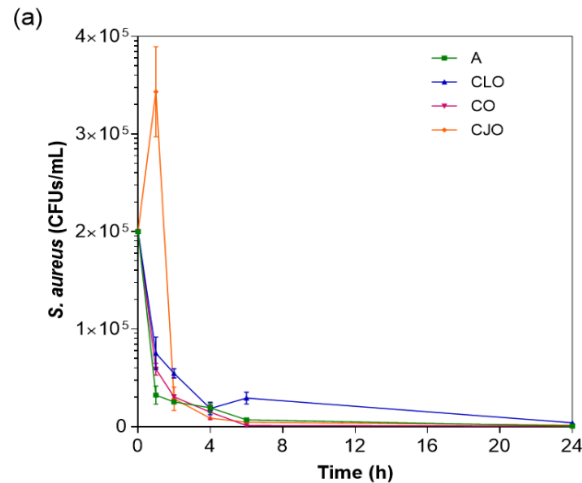
Bacteria Concentration:
1x10⁵ CFUs/mL

Growth Conditions:
37°C and 120 rpm.

Incubation Periods:
0, 1, 2, 4, 6 and 24 h

Bacteria reduction was observed from the first moments of interaction.

Free EOs were more effective than loaded.



Results & Discussion: Antimicrobial Action

Time Kill Kinetics

Bacteria Concentration:

1×10^5 CFUs/mL

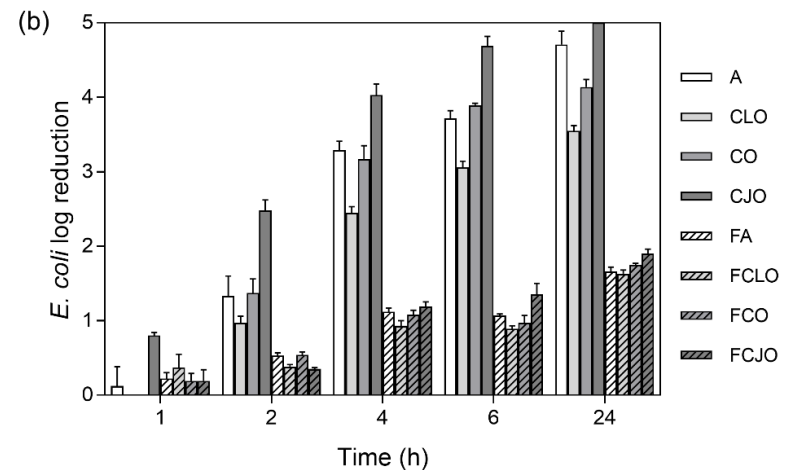
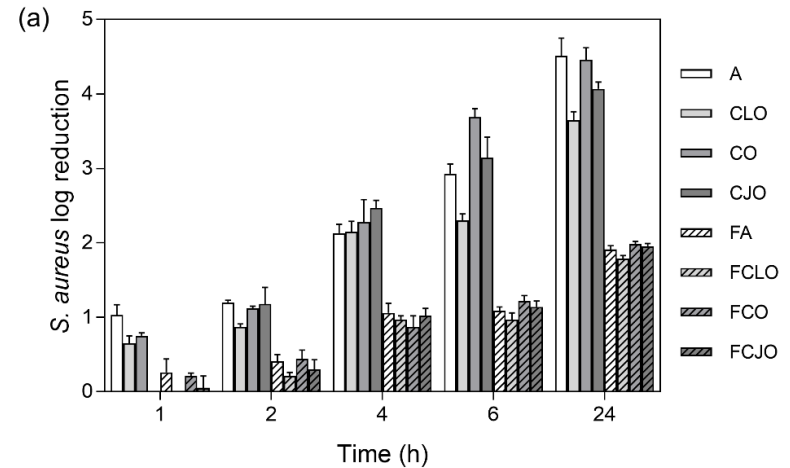
Growth Conditions:

37°C and 120 rpm.

Incubation Periods:

0, 1, 2, 4, 6 and 24 h

Log reduction was most significant after 24 h of culture. At this point, it was evident that *S. aureus* was more susceptible to the prolonged action of the EOs than the *E. coli*, the only exception being the CJO.



Results & Discussion: Antimicrobial Action

Membrane Permeability

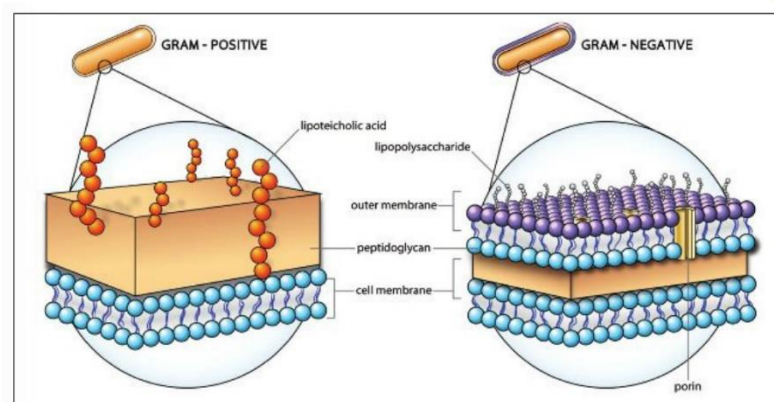
Bacteria Concentration: 1×10^5 CFUs/mL (adjusted in 5% glucose)

Growth Conditions: 37°C and 120 rpm for 6 h

Conductance of bacteria suspensions can be used to examine the cell membrane penetration by antimicrobial agents.

Relative electric conductivities (%)										
Bacteria	C	A	CLO	CO	CJO	F	FA	FCLO	FCO	FCJO
<i>S. aureus</i>	-0.49	19.10	9.81	19.83	20.41	-0.29	1.96	1.10	3.19	8.34
<i>E. coli</i>	-1.01	15.50	6.08	12.69	25.09	-1.32	0.75	0.67	1.69	4.49

Free state
Loaded



<https://www.ddw-online.com/therapeutics/p320363-tackling-multi-drug-resistant-bacteria.html>



Conclusions

EOs were successfully immobilized onto CA/PCL wet-spun fibers;

Microfibers displayed a **uniform and homogeneous** morphology with little variations in diameter;

FTIR and TGA data confirmed the successful incorporation of the EOs within the fibers by detecting characteristic peaks of the EOs and by demonstrating the increased overall **thermal stability** of the polymeric blend, respectively;

Even at small amounts, below MIC value, the **EOs-modified microfibers promoted cell death** compared to the control groups (unloaded and ampicillin-modified fibers), by disrupting and permeabilizing the cell cytoplasmic membrane;

The results demonstrated the potential of CA/PCL wet-spun microfibers loaded with EOs for applications in biomedicine, in which treatment of infections are a main target.

Full paper at: <https://doi.org/10.3390/biom10081129>

Felgueiras, H.P., et al. Biomolecules 2020, 10, 1129



Acknowledgments



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