

Evaluation of the inhibitory activities of acetylcholinesterase, α -glucosidase, pancreatic lipase and tyrosinase by phenolic extracts of honey.



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Abstract

Enzyme inhibition is one of the targeted therapeutic strategies in the treatment of Alzheimer's disease, diabetes, obesity and hyperpigmentation. The objective of this work is to evaluate the inhibitory effect of phenolic extracts of honey on key enzymes that are involved in these diseases (acetylcholinesterase, α-glucosidase, pancreatic lipase and tyrosinase). The color of the honey samples was determined spectrophotometrically and the polyphenols of monofloral and polyfloral honeys were recovered by liquid-solid extraction using amberlite XAD-4 resin. The inhibitory activities of phenolic extracts on the four enzymes were tested with six concentrations and the results obtained showed that the color of the tested honeys varied from extra light to dark amber. All the phenolic extracts tested exerted enzymatic inhibition. The phenolic fractions of *Myrtaceae* and polyfloral honeys showed the best anti-acetylcholinesterase, anti-lipase and anti-tyrosinase activities, respectively. The inhibitory effect of these honeys was comparable to that obtained with the reference inhibitors (galanthamine, acarbose, orlistat and α-kojic acid). The correlation matrix revealed highly significant correlations between color and inhibitory activity of the honeys. Since acetylcholinesterase, α-glucosidase, pancreatic lipase, and tyrosinase are closely associated with Alzheimer's, hyperglycemia, obesity, and hyperpigmentation, the use of honey polyphenols may have a beneficial effect in treating these diseases.

Introduction

Today, neurodegenerative diseases, such as Alzheimer's disease, are a huge health problem as they affect the majority of the elderly and are incurable. Acetylcholinesterase (AChE) inhibition is one of the targets for the treatment of Alzheimer's disease. The inhibitory effect of honey on AChE is attributed mainly to phenolic compounds that allow restoration of deficient cholinergic neurotransmission (Ahrari-Roodi et al., 2018; Muhammad et al., 2015). Diabetes is a disease that is of great concern. The α-glucosidase is the main enzyme responsible for hydrolyzing carbohydrates and increasing blood glucose levels. Phenolic compounds in honey exert inhibition on these digestive enzymes and consequently control hyperglycemia (Lakshmana-Senthil et al., 2019; Zaidi et al., 2019). Pancreatic lipase plays an important role in the hydrolysis of triglycerides into glycerol and fatty acids. Several studies have reported that natural polyphenols can inhibit pancreatic lipase and subsequently influence fat digestion and decrease energy intake (McDougall et al., 2009). Melanin formation in human skin occurs in the presence of a catalyst (tyrosinase) and UV light, which can lead to hyperpigmentation. Therefore, there is a need to find bioactive and harmless compounds from natural sources with tyrosinase inhibitory activity (Arrowitz et al., 2019). In search of natural and sustainable treatments for the mentioned diseases, the inhibitory activities of BSA, acetylcholinesterase, α -glucosidase, pancreatic lipase and tyrosinase of phenolic extracts from honey are investigated.

Results and discussion

 Table 1 : Intensities and color names (Pfund scale) of analyzed honeys

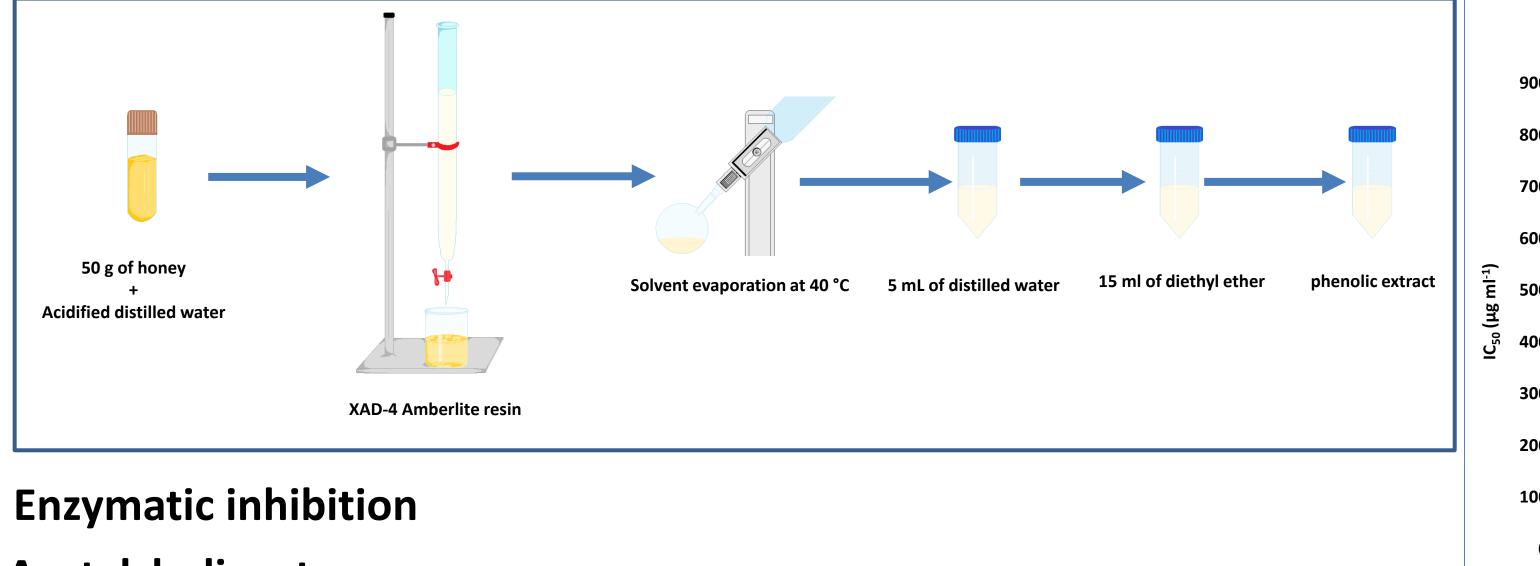
Sample	Color intensity	Name of the	Sample	Color intensity	Name of the	
Sample	mm Pfund	color		mm Pfund	color	

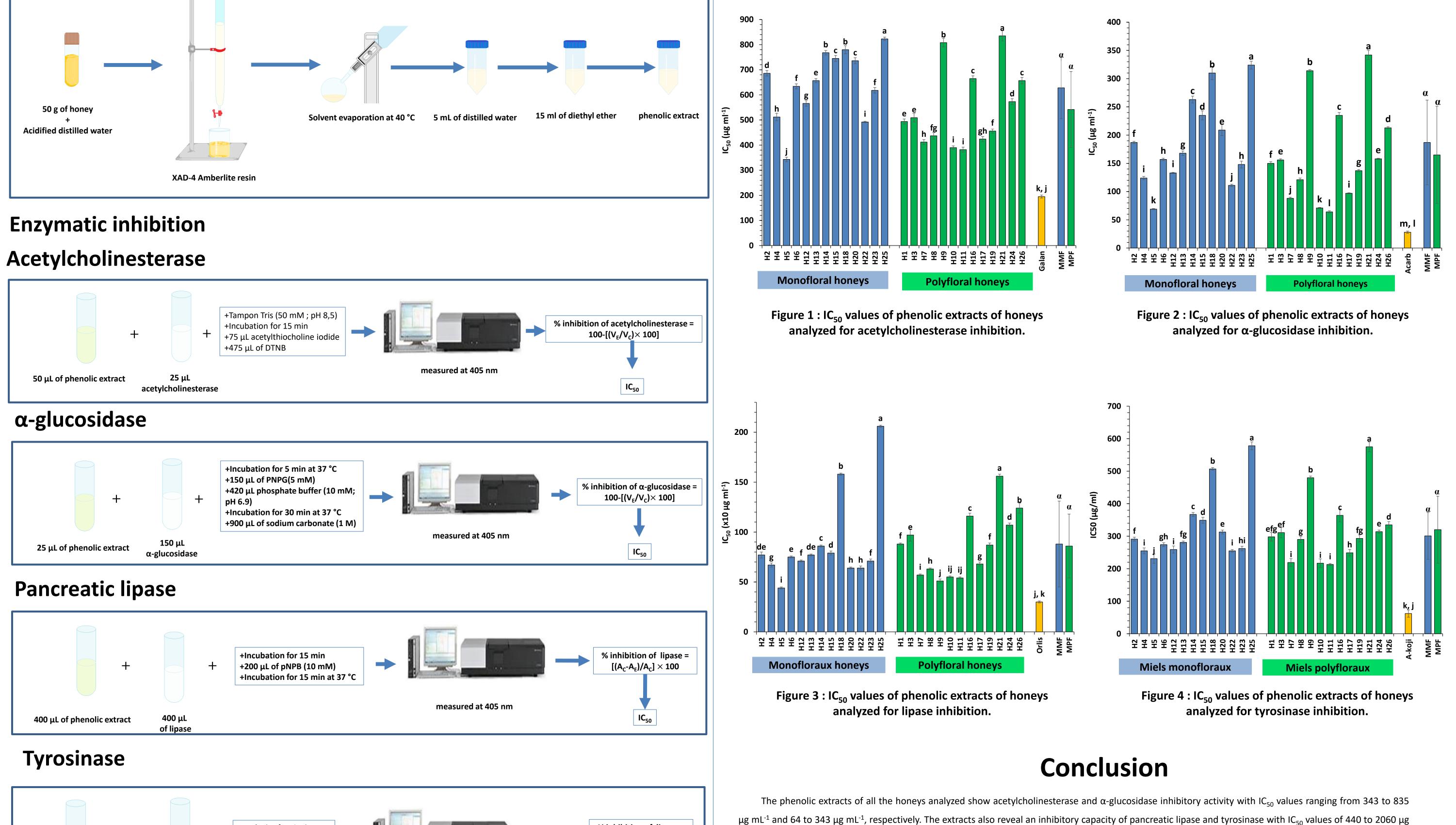
Samples

A total of 26 honeys are provided by beekeepers from 16 wilayas of Algeria from East to West (Skikda, Batna, Jijel, Sétif, Bejaia, Bordj Bouarrerid, Tizi-Ouzou, Boumerdès, Djelfa, Médéa, Aïn Defla, Chlef, El Bayadh, Mostaganem, Naâma and Tlemcen). The samples were stored at room temperature and protected from light and humidity. the color intensity of the studied honeys is determined with the method Pfund.

honey	H2	150±3 f	Dark amber	Polyfloral honey	H1	191±0 f	Dark amber
	H4	412 ± 4 b	Dark amber		H3	210±1 e	Dark amber
	H5	402±6 c	Dark amber		H7	115±0 h	Dark amber
	H6	112 ± 3 i	Amber		H8	342±1 c	Dark amber
	H12	255±3 d	Dark amber		H9	61±4 k	Light amber
	H13	195±7 e	Dark amber		H10	520±9 b	Dark amber
	H14	53±3 j	Light amber		H11	619±6 a	Dark amber
nofile	H15	133 ± 4 g	Dark amber		H16	73±2 j	Light amber
Monofloral	H18	46±1 k	Extra light amber		H17	174±2 g	Dark amber
	H20	123 ± 1 h	Dark amber		H19	331±1 d	Dark amber
	H22	462±0 a	Dark amber		H21	62±1 k	Light amber
	H23	130±0 g	Dark amber		H24	108 ± 3 hi	Amber
	H25	33±1 l	White		H26	107 ± 2 i	Amber

Extraction of phenolic compounds





mL⁻¹ and 213 to 578 µg mL⁻¹, respectively.

+ 1 mL of L-DOPA (2.5 mM)	- + 1.8 mL phosphate buffer (100 mM; pH 6.8)	+ 0.1 mL of tyrosinase + 0.1 mL of phenolic extract +Incubation for 25 min	measured at 492 nm	$[(A_{c}-A_{E})/A_{c}] \times 100$ IC_{50}	mL ⁻¹ and 213 to 578 μg mL ⁻¹ , respectively. Bibliogra		
					Ahrari-Roodi, P., Moosavi, Z., Afkhami Goli, A., Azizzadeh, M., & Hosseinzadeh, H. (2018). Histopathological st		
					and liver. Iranian Journal of Toxicology, 12(3), 1–8.		
					Arrowitz, C., Schoelermann, A. M., Mann, T., Jiang, L. I., Weber, T., & Kolbe, L. (2019). Effective tyrosinase		
~ I					Investigative Dermatology, 139(8), 1691-1698.		
Statistical analysis					Lakshmana Senthil, S., Chandrasekaran1, R., Arjun, H.A., & Anantharaman, P. (2019). In vitro and in silico inh		
	•				mellitus. Carbohydrate Polymers, 209, 350–355.		
					McDougall, G. J., Kulkarni, N. N., & Stewart, D. (2009). Berry polyphenols inhibit pancreatic lipase activity in viti		
Student's t-test, ANOVA (analysis of variance with a single classification criterion) and correlation matrix are applied to the data presented and are performed					Muhammad, A., Odunola, O. A., Gbadegesin, M. A., Sallau, A. B., Ndidi, U. S., & Ibrahim, M. A. (2015). Inhibit		
using STATISTICA 7.1 softw	vare.				Alzheimer's Disease, 2015, 1–7.		
ICED of phonolic avtracts from honoy camples to inhibit anyymas (AChE, a glycosidase, linase and tyrosinase) are calculated by regression analysis using Eyeol					Zaidi, H., Ouchemoukh, S., Amessis-Ouchemoukh, N., Debbache, N., Pacheco, R., Serralheiro, M.L., &		

% inhibition of lipase =

IC50 of phenolic extracts from honey samples to inhibit enzymes (AChE, α-glycosidase, lipase and tyrosinase) are calculated by regression analysis using Excel 2013.

+Incubation for 10 min + 0.1 mL of tyrosinase



tudy of protective effects of honey on subacute toxicity of acrylamideinduced tissue lesions in rats brain

inhibition by Thiamidol results in significant improvement of mild to moderate Melasma. Journal of

nibition properties of fucoidan against α -amylase and α -D-glucosidase with relevance to type 2 diabetes

tro. Food Chemistry, 115(1), 193–199.

tory effects of sodium arsenite and Acacia honey on acetylcholinesterase in rats. International Journal of

ujo, M.E. (2019). Biological properties of phenolic compound extracts in selected Algerian honeys-The inhibition of acetylcholinesterase and α -glucosidase activities. European Journal of Integrative Medicine, 25, 77–84.



The 7th International Electronic Conference on Medicinal Chemistry 01–30 NOVEMBER 2021 ONLINE