



Effects of Alkalinity-Induced Iron Deficiency on Physiological and Growth Variables of some Upland Rice Cultivars under Laboratory Condition ⁺

Olayinka Oyedeji 12,3,*, Olalekan Sakariyawo 4, Kehinde Adeboye 2,3,5, Mamadou Fofana 3 and Oludayo Daniel 4

- ¹ Department of Agriculture, Food and Nutritional Science, University of Alberta, Edmonton, AB, T6G 2J8Canada
- ² Centre of Excellence in Agricultural Development and Sustainable Environment, Federal University of Agriculture, Abeokuta, P.M.B. 2240, Alabata, Ogun State. Nigeria; <u>kennyomaak@gmail.com</u>
- ³ Africa Rice Center, International Institute of Tropical Agriculture, Ibadan sub-station, Oyo State, Nigeria; yinkaoyedeji21@gmail.com
- ⁴ College of Plant Sciences and Crop Production, Federal University of Agriculture, Abeokuta, P.M.B 2240, Alabata, Ogun State, Nigeria; <u>adetanwa@yahoo.com</u> (O.S.); <u>drdayodaniel@yahoo.co.uk</u> (O.D.)
- ⁵ Department of Agricultural Technology, Ekiti State College of Agriculture and Technology, P.M.B 394, Isan-Ekiti, Nigeria
- * Correspondence: yinkaogutuga@gmail.com
- + Presented at the 2nd International Electronic Conference on Plant Sciences 10th Anniversary of Journal Plants, 1–15 December 2021; Available online: https://iecps2021.sciforum.net/.

19 Abstract: Prevalence of iron deficiency in upland rice under alkalinity stress is capable of constraining its production. This investigation aimed to explicate the physiological basis of iron 20 deficiency tolerance in some upland rice genotypes. Eighty upland rice genotypes were 21 characterized for iron deficiency tolerance at seedling growth stage in a sand-culture hydroponics 22 with varying NaHCO₃ concentrations (0, 15 and 25 mM). The treatments were arranged in 23 completely randomised design with three replicates. Significant decrease was observed on leaf iron 24 concentration, SPAD meter readings, leaf photosynthetic efficiency, quantum yield and growth 25 variables with increasing concentration of NaHCO3. Iron tolerance index was further estimated 26 27 based on these parameters and used for ranking the genotypes. Based on iron tolerance index, genotypes were divided into 6 groups with Caipo and NERICA 7 identified as the most and least 28 29 tolerant to iron deficiency respectively. The basis of iron deficiency tolerance is discussed on relation to the stability of photosynthetic apparatus and plan growth under alkalinity stress. 30

Keywords: Quantum yield; Photosynthetic apparatus; NERICA; Sand-culture hydroponics; iron 31 tolerance index 32

33

34

1. Introduction

The competing need for freshwater in the cultivation of lowland rice production 35 system necessitated the need to explore other ecologies. Upland rice cultivation is another 36 option in rice production. In the tropics, it accounts for 8% of cultivated land while 37 lowland rice was reported to be 92% [1]. According to [2], the cultivation pattern of upland 38 rice in the tropics is associated with high rainfall, reduced sufficiency for rice and Gross 39 National Income level; however, this had been a subject of debate by other stakeholders. 40

Iron deficiency is one of the production constraints in the establishment of upland 41 rice in the tropics. It is closely associated with the bicarbonate content in the soil with 42 increased soil and plant alkalinity [3]. The increasing soil water content could lead to 43 availability of excess bicarbonates in the soil as CO₂ content increases with reduced soil 44 porosity. Furthermore, increased soil pH had been reported to induce iron deficiency 45

Citation: Oyedeji O.; Sakariyawo O.; Adeboye K.; Fofana M.; Daniel O. Title. *Biol. Life Sci. Forum* **2021**, *1*, x. https://doi.org/10.3390/xxxxx

Academic Editor: Feibo Wu

Published: 7 December 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/).



1

2

3

4

5

6 7

8

9

10

11

12 13

14

15

16 17

15

16

chlorosis and reduce the solubility of iron in the soil. Iron is a component of the electron 1 transport chain in mitochondria and chloroplast. Thus, the disruption of electron transfer 2 would elicit reactive oxygen radicals, causing cellular oxidative damage and reduced 3 growth [4]. As a component of cellular anti-oxidant systems [5], deficiency of iron will 4 also affect redox homeostasis in the cell. Increased soil alkalinity resulting from high 5 bicarbonate concentrations may negatively affect mineral composition, especially 6 micronutrients such as Zn, Mn and Mg [6].

Genotypic variation in response to iron availability in the soil has been reported in 8 the literature [4,7–8] and may provide a basis for breeding-based crop improvement 9 strategies. However, understanding the physiological basis of crop performance may 10 complement crop improvement efforts for developing iron deficiency tolerant rice. The 11 present work is therefore aimed at investigating the physiological characteristics of some 12 selected upland rice genotypes under varying concentration of bicarbonate in the growth 13 medium. 14

2. Materials and Methods

2.1. Plant Materials and Experimental Setup

Eighty upland rice genotypes of a broad range of morphological diversity and origins 17 were used in the screen house experiment. The experiment was conducted at the 18 International Institute of Tropical Agriculture, Ibadan sub-Station, Nigeria, with 19 day/night temperature range of 30/22 °C and relative humidity of at least 50% during the 20 day. The screen house was disease-free and lit by natural lighting. The treatments 21 consisted of eighty genotypes and concentrations of NaHCO₃ (0 mM, 15 mM and 25 mM). 22 The genotypes were planted in a completely randomised design with three replications. 23

River sand was collected and washed thoroughly under water, air dried, autoclaved 24 at 70 °C for 72 h and sieved at 25 mm particle size. The experimental plants were raised in 25 3-litre dark-coloured pots to minimise light penetration into the culture solution thereby, 26 reducing algal growth. The surfaces of the 3-litre pots were wiped with 70% ethanol and 27 washed thrice with 5% HNO₃ to remove any contamination. Each plant pot was labelled 28 and filled with an equal amount of sand to hold the plants in place during growth. A 29 modified basal nutrient solution suggested by [9] for normal growth of seedlings was 30 utilised. Rice seeds were directly sown into the pots and irrigated with the optimum 31 nutrient solution till 15 Days After Sowing (DAS). 32

2.1. Sampling and Data Collection

Four rice seedlings were maintained per pot. SPAD meter readings of seedlings were 34 measured using the SPAD-502 meter (Minolta Ltd., Tokyo, Japan) and leaf colours were 35 visually scored thrice during the screening period i.e., 15, 25 and 35 DAS (5 = normal 36 growth, no chlorosis; 4 = slight yellowing of upper leaves and the leaf veins; 3 = interveinal 37 chlorosis in the upper leaves; 2 = interveinal chlorosis of the upper leaves along with some 38 apparent stunting of growth; 1 = severe chlorosis, stunted growth and necrosis in the 39 youngest leaves [10].

The pH of the nutrient solution in all pots was monitored with a pH meter (Hanna 41 42 Instruments, Italy). Plant height (PHT) was measured with a metre rule at 15, 25 and 35 DAS. At the 35 DAS, biomass index was estimated by weighing oven-dried shoots and 43 roots of two seedlings per pot. Leaf Fe concentrations were also determined and calculated 44 following the [11] protocol. At 35 DAS at 9.00 a.m quantum yield (Fv/Fm) was measured 45 with a pocket PEA Chlorophyll fluorimeter (Hansatec Instruments Ltd., Norfolk, UK). 46 Photosynthetic rates were measured using the LI-6400XT portable photosynthesis system 47 (LI-COR Biosciences, Lincoln, USA). 48

49

2.3. Data Analysis

Fe tolerance indexes for each parameter and the average Fe tolerance indexes per 2 genotype were calculated as described by [12,13]. The genotypes were grouped into 3 clusters based on average Fe tolerance indexes using the agglomerative clustering 4 method. All data collected except Fe tolerance indexes were subjected to Analysis of 5 Variance (ANOVA) using R statistical package. Means were separated using LSD at 5%, 6 1% and 0.1% probability level. 7

3. Results

9 The average Fe tolerance indexes of the 80 upland genotypes ranged from 34.3 to 109.0. They were categorised into 6 clusters with CAIPO and NERICA 7 as first and last 10 genotype on the table as shown in Table 1. At 35 DAS, there was a significant reduction 11 in the SPAD meter readings in all the genotypes with increasing concentrations of 12 13 NaHCO₃ (Table 2). A similar pattern was observed on the leaf iron concentration, leaf photosynthetic rate, quantum yield, plant height, shoot dry weight and root dry weight 14 in all the upland rice genotypes (Table 2). Similar to all other parameters, there was a 15 significant reduction in visual chlorosis scoring with increasing concentration of NaHCO3 16 with the exception of Caipo that maintained stable visual chlorosis score at 15 and 25 mM 17 NaHCO₃. 18

Table 1. Characteristics of clusters obtained from average iron tolerance indexes based on physiological and growth parameters of upland rice genotypes.

Cluster	Membership	Size	Min	Max	Mean	StdDev
1	Caipo, OFADA3, LAC23	3	91.2	109	97.5	9.98
2	FARO65	1	85.1	85.11	85.11	
	PCT11-1-3-1, NERICA3, DURADO, NERICA15, NERICA16, CIRAD409,					
3	IGBEMORED, Azucena, IAC120, CIRAD403, Palapo, NERICA4, IRAT13,	17	61.4	80.36	67.33	5.53
	EbonyiLocalBest, Wayrem, OS4, NERICA18					
4	NERICA13, NERICA11, NERICA17, ARICA5, NERICA14, CIRAD358,	12	49.7	59.17	54.76	3.13
4	IRAT170, NERICA8, NERICA5, CT13582-15-M, IRAT226, IRAT2	12				5.15
5	CIRAD394, WAB56-50, ITA301, FARO63, IRAT216, NERICA12,	14	37.9	53.42	47.4	5.33
5	Vandana, IRAT212, IRAT257, ARICA4, IAC47, ChinaBest, IRAT364, OS6	14				5.55
	WAB181-18, WABC165, ITA128, IR7267-12-2-3, WAB56-104,					
	MOROBEREKAN, NERICA6, Pamira, NERICA9, IRAT133, WAB99-16,		34.3	51.99	43.89	
(APO, NERICA10, IRAT144, CIRAD488, NERICA1, OFADA4, Palawan,	22				4 77
6	IGUAPECATETO, IRAT112, IRAT109, ART-27-58-7-1-2, OFADA1,	33				4.77
	IRAT362, ART27-190-1-3-3, FARO64, Curinga, SabonDaga, NERICA2,					
	Primavera, OFADA2, WAB638-1, NERICA7					

Table 2. Effect of NaHCO₃ concentrations on some physiological and growth parameters in upland rice genotypes using a representative of the 80 upland rice genotypes.

[A]													
Genotype	SPAD reading			Leaf Fe concentration (mg/kg)			Photosynthetic rate (µmol(CO ₂) m ⁻² s ⁻¹)			Quantum yield (F _v /F _m)			
	0mM	15mM	25mM	0mM	15mM	25mM	0mM	15mM	25mM	0mM	150Mm	25mM	
	Most tolerant												
Caipo	40.54	29.61	27.38	86.64	61.32	49.57	27.44	20.33	8.28	0.787	0.736	0.731	
OFADA3	33.03	24.84	21.95	85.34	62.51	50.76	28.66	21.54	9.49	0.783	0.734	0.727	
LAC23	35.19	25.97	23.26	89.54	62.21	49.96	27.21	20.09	8.05	0.787	0.738	0.731	
FARO65	37.42	27.88	14.09	86.24	38.61	15.05	26.77	11.37	6.49	0.786	0.698	0.599	
PCT111	35.44	23.33	20.33	85.15	55.89	37.14	27.98	20.44	7.98	0.781	0.724	0.698	
Least tolerant													
NERICA2	37.91	12.06	9.79	88.54	36.72	13.08	27.50	12.08	6.45	0.787	0.702	0.601	
PRIMAVER	38.44	23.01	8.77	91.94	40.92	16.48	27.87	12.45	6.80	0.785	0.697	0.594	

1

8

19

20

OFADA2	32.36	12.59	9.31	86.94	35.18	11.48	27.86	12.44	6.81	0.786	0.699	0.600
WAB638-1	31.26	17.56	7.52	89.84	38.82	14.38	26.65	11.23	5.58	0.780	0.690	0.590
NERICA7	41.21	10.59	8.47	88.34	36.58	12.88	26.86	11.44	5.81	0.784	0.694	0.594
Mean	36.28	19.37	16.47	87.85	46.87	27.08	27.483	15.343	7.177	0.785	0.712	0.646
F values (ANOVA)												
G		1389.8**	ŧ		451.52***		2880.9***					
Т	Т 7851.76***					48719.0***				57428.9***		
G x T	G x T 209.84***			453.6***			183.45***			1351.2***		
[B]												

[4]													
Constra	Colour visual score			Plant Height (cm)			ST	DRYWEI	GHT	RTDRYWEIGHT			
Genotype							(g)			(g)			
	0mM	15mM	25mM	0mM	15mM	25mM	0mM	15mM	25Mm	0mM	15mM	25mM	
					Most	tolerant							
Caipo	5.0	4.0	4.0	43.71	36.47	32.27	0.949	0.706	0.629	0.694	0.554	0.489	
OFADA3	5.0	4.0	3.0	42.21	32.84	19.44	0.713	0.423	0.397	0.402	0.311	0.219	
LAC23	5.0	4.0	3.0	41.21	30.52	29.20	0.902	0.607	0.403	0.408	0.314	0.289	
FARO65	5.0	4.0	2.0	58.93	43.02	33.73	1.029	0.576	0.199	0.750	0.282	0.094	
PCT111	5.0	3.5	3.0	44.11	34.69	29.85	0.746	0.346	0.149	0.252	0.174	0.089	
	Least tolerant												
NERICA2	5.0	2.0	1.5	53.54	20.86	10.88	0.814	0.402	0.017	0.409	0.084	0.011	
PRIMAVERA	5.0	2.5	1.0	54.51	33.72	21.16	0.837	0.449	0.092	0.457	0.189	0.002	
OFADA2	5.0	2.0	1.0	56.94	20.18	19.43	1.013	0.795	0.022	0.352	0.036	0.008	
WAB638-1	5.0	2.0	1.0	57.52	23.48	16.24	0.669	0.201	0.043	0.208	0.054	0.005	
NERICA7	5.0	1.5	1.0	65.32	33.29	24.17	0.813	0.402	0.022	0.409	0.084	0.011	
Mean	5.00	2.95	2.05	51.79	30.90	25.12	0.849	0.491	0.197	0.429	0.208	0.122	
F values													
(ANOVA)													
G	41.00***			75.55***		371.04***			1618.75***				
Т	627.25***			2162.414*** 6920.06***				÷	6706.19***				
G x T	19.75***			65.73***			111.69***			234.94***			
		/	0) (45)	<i>c</i> 1 <i>c</i>		LILCON		1	—			J.J. J.J.J.	

G, Genotypes; T, Treatments (0Mm, 15Mm and 25Mm of NaHCO₃); G x T, Genotype by Treatment interactions. *, **, *** indicate probability of significance at 0.05, 0.01 and 0.001, respectively.

4. Discussion

Nutrient solutions containing bicarbonate or complete absence of iron have been 4 used extensively in the literature in the investigation of iron deficiency chlorosis in 5 laboratory conditions [14,15]. Literature has also proven a significant correlation between 6 the results in laboratory conditions and that of field trials as recorded in research 7 conducted by [16,17]. Chlorosis is a common physiological marker used to express 8 bicarbonate stress, thus it informs the selection of tolerant or susceptible genotypes. 9

The selected upland rice genotypes were phenotyped to determine their variations 10 to alkalinity stress in the Laboratory. The reduced SPAD meter reading as a result of 11 increasing NaHCO₃ concentration in the growth medium was corroborated visually with 12 increasing chlorosis. This could be linked to the fact that iron constituted 80% of the 13 chloroplast [18]. Furthermore, 60% of iron was involved in the electron transport chain 14 [19]. This disruption in the functional integrity of the chloroplast could have explained 15 observed reduction in the quantum yield and subsequently leaf photosynthesis with 16 increasing concentration of the NaHCO₃. A similar position was also reported by [20] 17 when maize was subjected to iron deficiency in the growth medium. The reduced 18 19 availability of assimilates from the reduction in leaf photosynthesis under iron deficiency could have explained the reduced growth under this condition among all the upland rice 20 genotypes investigated. 21

An alternative physiological mechanism linking iron deficiency with leaf 22 photosynthesis and concentration of iron in different organs of the plant was proposed by 23 [21]. In their explanation, it was proposed that iron deficiency linked with a reduction in 24

2 3

leaf photosynthesis could result in the disruption of sucrose transportation in the phloem. 1 It could also lead to the differential distribution of iron in the plant shoot and root, with 2 more of iron found in the plant shoot than in the plant root [21]. It was posited that this 3 observed response pattern would have elicited a signal from the plant shoot to the roots 4 5 leading to changes in the reactive oxygen system and consequently resulting in oxidative damage of organs. However, this distribution of iron was observed in matured shoot and 6 roots, unlike what was obtained in our investigation, where the leaf iron concentration 7 was sampled form the flag leaf. 8

9 In context of iron tolerance stress index, it was revealed that the upland rice genotypes were categorized into 6 groups with Caipo and NERICA 7 being the most and 10 least ranked respectively. This could be attributed to their pattern of growth and chlorosis 11 response with increasing bicarbonate concentration in the medium. While the former 12 maintained a steady decline in its growth variables with decreasing iron concentration, 13 the later had a twofold reduction in growth, especially at bicarbonate concentration of 25 14 mM. 15

5. Conclusions

17 These evidences, taken together indicated that physiological responses to iron deficiency, especially under alkalinity stress, is expressed in the reduction of leaf iron 18 concentration. This may perhaps be linked with the observed decreased in the SPAD-19 meter reading, quantum yield and leaf photosynthetic rate. The reduced availability of 20 assimilates would negatively affect carbon budgeted for growth as portrayed in reduced 21 plant height, shoot and root dry weight at 35 DAS. In addition, it was found that within 22 this germplasm evaluated, Caipo and NERICA 7, were regarded as the most and least 23 tolerant to iron deficiency. 24

Funding: This research was funded by The World Bank Centre of Excellence in Agricultural 26 Development and Sustainable Environment (CEADESE), Federal University of Agriculture 27 Abeokuta under Grant Number 0023.

Conflicts of Interest: The authors declare no conflict of interest

References

- 1 Saito, K.; Asai, H.; Zhao, D.; Laborte, A.G.; Grenier, C. Progress in varietal improvement for increasing upland rice 30 productivity in the tropics. Plant Prod. Sci. 2018, 21, 145–158, doi:10.1080/1343943x.2018.1459751. 31
- van Ittersum, M.K.; van Bussel, L.G.J.; Wolf, J.; Grassini, P.; van Wart, J.; Guilpart, N.; Claessens, L.; de Groot, H.; Wiebe, K.; 32 2 Mason-D'Croz, D.; et al. Can sub-Saharan Africa feed itself? Proc. Natl. Acad. Sci. 2016, 113, 14964–14969, doi:10.1073/pnas.1610359113.
- Mengel, K. Iron availability in plan tissues-iron chlorosis on calcareous soils. In J. Abadia Ed., Iron nutrition in soils and 3. plant. Kluwer academic publishers, 1995, pp 389-397.
- 4. Ramírez, L.; Bartoli, C.G.; LaMattina, L. Glutathione and ascorbic acid protect Arabidopsis plants against detrimental effects 37 of iron deficiency. J. Exp. Bot. 2013, 64, 3169-3178, doi:10.1093/jxb/ert153. 38
- Kumar, P.; Tewari, R.K.; Sharma, P.N. Sodium nitroprusside-mediated alleviation of iron deficiency and modulation of 5. antioxidant responses in maize plants. AoB PLANTS 2010, 2010, plq002, doi:10.1093/aobpla/plq002.
- Roosta, H.R. INTERACTION BETWEEN WATER ALKALINITY AND NUTRIENT SOLUTION PH ON THE VEGETATIVE 6. GROWTH, CHLOROPHYLL FLUORESCENCE AND LEAF MAGNESIUM, IRON, MANGANESE, AND ZINC CONCENTRATIONS IN LETTUCE. J. Plant Nutr. 2011, 34, 717-731, doi:10.1080/01904167.2011.540687.
- 7. Bavaresco, L., Fregoni, M., & Perino, A. Physiological aspects of lime-induced chlorosis in some Vitis species. I. Pot trial on calcareous soil. Vitis, 1994, 33(2), 123-126.
- Pestana, M.; de Varennes, A.; Abadia, J.; Faria, E.A. Differential tolerance to iron deficiency of citrus rootstocks grown in 8. 46 nutrient solution. Sci. Hortic. 2005, 104, 25-36, doi:10.1016/j.scienta.2004.07.007. 47
- 9 Yoshida, S., Forno, D., Cook, J., & Gomez, K. Laboratory manual for physiological studies of rice. International Rice Research 48 49 Institute, 1976.
- 10. Mamidi, S.; Chikara, S.; Goos, R.J.; Hyten, D.L.; Annam, D.; Moghaddam, S.M.; Lee, R.K.; Cregan, P.B.; McClean, P.E. 50 Genome-Wide Association Analysis Identifies Candidate Genes Associated with Iron Deficiency Chlorosis in Soybean. Plant 51 Genome 2011, 4, 154–164, doi:10.3835/plantgenome2011.04.0011. 52

16

25

28

29

33 34 35

36

39

40

41

42

43

- 11. AOAC. Official methods of analysis. Mineral Analysis flame absorption spectroscopy, 17th ed. Association of Analytical Community, 2006.
- 12. Akhtar, S., Shahzad, A., Arshad, M. and Hassan, F. Morpho-physiological evaluation of groundnut (Arachhypogaea L.) genotypes for iron deficiency tolerance. *Pak J Bot*, **2013**, 45, 893–899.
- 13. Zeng, L.; Shannon, M.C.; Grieve, C.M. Evaluation of salt tolerance in rice genotypes by multiple agronomic parameters. *Euphytica* **2002**, 127, 235–245, doi:10.1023/a:1020262932277.
- 14. Alcántara, E.; Romera, F.; Cañete, M.; De La Guardia, M. Effects of bicarbonate and iron supply on Fe(III) reducing capacity of roots and leaf chlorosis of the susceptible peach rootstock "Nemaguard." *J. Plant Nutr.* **2000**, *23*, 1607–1617, doi:10.1080/01904160009382127.
- 15. Zribi, K.; Gharsalli, M. EFFECT OF BICARBONATE ON GROWTH AND IRON NUTRITION OF PEA. J. Plant Nutr. 2002, 25, 2143–2149, doi:10.1081/pln-120014066.
- 16. Coulombe, B.A.; Chaney, R.L.; Wiebold, W.J. Use of bicarbonate in screening soybeans for resistance to iron chlorosis. *J. Plant Nutr.* **1984**, *7*, 411–425, doi:10.1080/01904168409363208.
- 17. Sakariyawo, O.S.; Oyedeji, O.E.; Soretire, A. Effect of iron deficiency on the growth, development and grain yield of some selected upland rice genotypes in the rainforest. *J. Plant Nutr.* **2020**, *43*, 851–863, doi:10.1080/01904167.2020.1711936.
- 18. Kim, S.A.; Guerinot, M.L. Mining iron: Iron uptake and transport in plants. *FEBS Lett.* 2007, 581, 2273–2280, doi:10.1016/j.febslet.2007.04.043.
- 19. Terry, N.; Abadia, J. Function of iron in chloroplasts. J. Plant Nutr. **1986**, 9, 609–646, doi:10.1080/01904168609363470.
- Chen, J.; Wu, F.-H.; Shang, Y.-T.; Wang, W.-H.; Hu, W.-J.; Simon, M.; Liu, X.; Shangguan, Z.-P.; Zheng, H.-L. Hydrogen sulphide improves adaptation ofZea maysseedlings to iron deficiency. *J. Exp. Bot.* 2015, 66, 6605–6622, doi:10.1093/jxb/erv368.
- 21. Chen, L.; Wang, G.; Chen, P.; Zhu, H.; Wang, S.; Ding, Y. Shoot-Root Communication Plays a Key Role in Physiological Alterations of Rice (Oryza sativa) Under Iron Deficiency. *Front. Plant Sci.* **2018**, *9*, 9, doi:10.3389/fpls.2018.00757.