



Novel Formula

Mosquito Larvicides



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Vector-borne diseases account for more than 17% of all infections, causing more than 1 million deaths annually

Mosquitoes are responsible for the transmission of many medically important pathogens and parasites <u>such as:</u>

Viruses

Bacteria

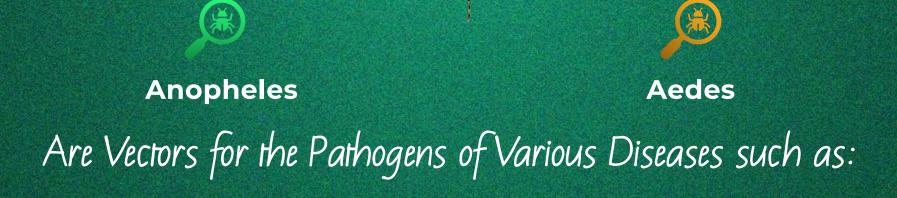
Protozoans

Nematodes

Which Cause Serious Diseases

INTRODUCTION Several Mosquito Species Belonging to Genera

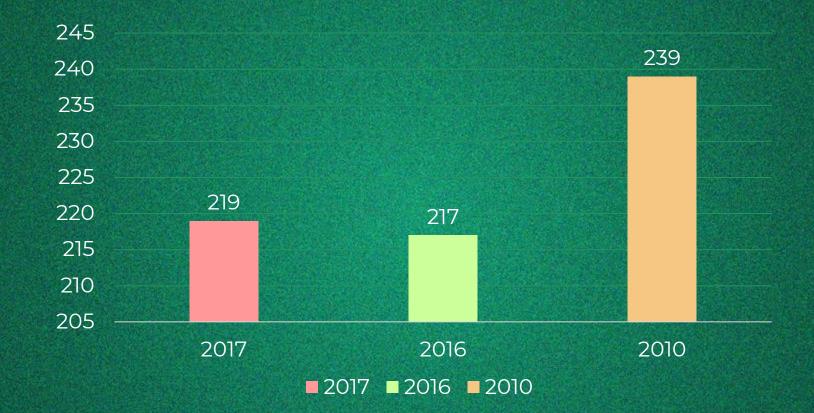
Culex



Several Mosquito Species Belonging to Genera Are Vectors for the Pathogens of Various Diseases such as:



The Global and Regional Malaria Burden, in Numbers



Also, WHO reported that 80% of the global malaria was estimated in 15 countries in sub-Saharan Africa and India in 2017

WHO has Considered Mosquito Vector Control as one of the Important measures to Control Diseases caused by Mosquitoes.



The Use of Chemical Insecticides such as Organophosphate is the Effective Method of Choice.

The common Mosquito Larvicides, Nowadays, include Temephos and Spinosad.

Spinosad is a Mixture of Chemical Compounds in the Spinosyn Family.

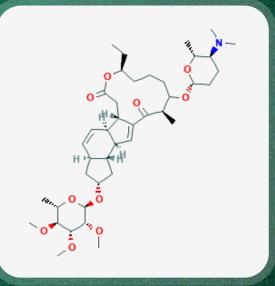
It Consists of:

A unique tetracyclic ring system

Attached to:

An amino sugar (D-forosamine)

A neutral sugar (tri-0-methyl-L-rhamnose)



However, their side effects on human health, environment and development of resistance in mosquitoes against pesticides are causes of concern

About 1 million world-wide deaths and chronic diseases per year were due to the poisoning effect of the pesticide

Production workers, formulators, sprayers, mixers, loaders, and agricultural farm workers are at high risk

INTRODUCTION That because Pesticides Affect by causing

Cancer

Immunosuppression

Congenital Abnormalities

Mental Retardation

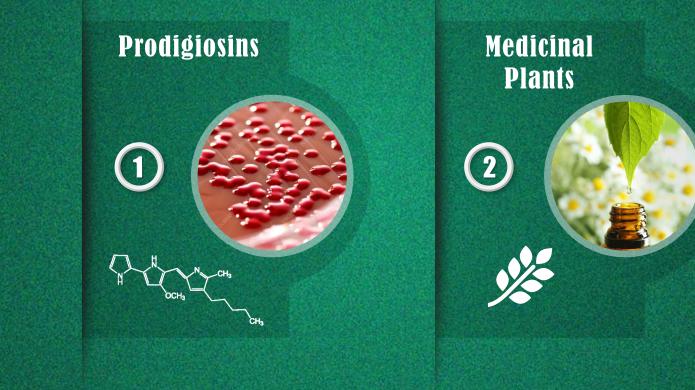
Hormonal Disruption where they act as Hormone Antagonist

Recently, concerns increased with respect to public health and environmental security requiring detection of natural products that may be used against insect pests

It is known that larvicides play a vital role in controlling mosquitoes in their breeding sites as mosquitoes breed in water, and thus, it is easy to deal with them in this habitat

Because of the worsening problems of drug resistance, there has been an urgent need for the discovery of a new chemical class of antimalarial agents

As part of an ongoing natural product research program, microbial extracts have been screened for in vitro antimalarial activity





Prodigiosins constitute a class of natural products isolated from bacterial strains such as *Serratia marcescens*.

S. marcescens, a G-ve Entero bactericeae has got its attention because of Tripyrrylmethene, a naturally occurring dark red pigment.

Serratia is commonly found in water, soil and is also associated with plants and animals.

They are opportunistic pathogens, especially in compromised hosts, and may cause sepsis, bacteremia, meningitis, osteomyelitis, endocarditis, infections of the urinary and respiratory tracts.



Prodigiosin has been shown to be associated in extracellular vesicles, cell associated or present in intracellular granules.

It belongs to the family Prodigiosinide, which is common to all the members of this family <u>such as:</u>







Metacyclo Prodigiosin



Dipyrrolildipirromethan Prodigiosin



Prodigiosin, a bioactive secondary metabolite is characterized by a common pyrrolyl pyrromethane skeleton.

In the PDG molecule (C20H25N30) and a molecular weight of 323.44 Da, the pyrrolyldipyrromethene structure (linear tripyrrole) is composed of two of the rings linked to each other and the third is attached via a methane bridge.

PDG and undecylprodigiosin belong to linear tripyrroles while metacycloprodigiosin, prodigiosin R1, and streptorubin belong to cyclic prodigiosins.

The highly conjugated system of seven double bonds is responsible for the intense pigmentation as it forms dark red square pyramidal crystals with a green reflex.



Prodigiosin revealed a broad range of inhibitory activities against many bacterial, fungal, and protozoan species

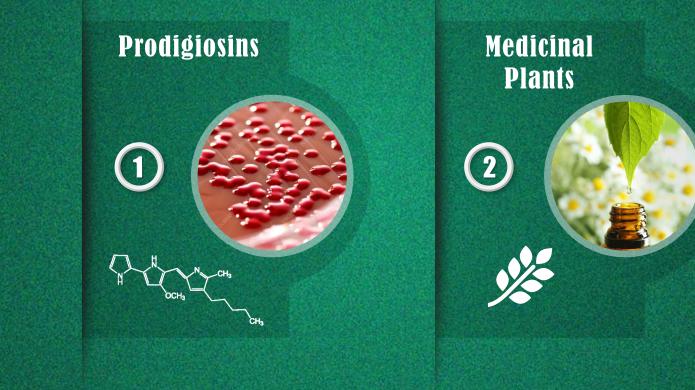
It has antibacterial activity due to Its ability to pass through the outer membrane and inhibiting target enzymes such as DNA gyrase and topoisomerase IV, which inhibited the cell growth.

Prodigiosins are strong therapeutic molecules especially for their immunosuppressive properties and anticancer properties.



Four possible mechanisms are suggested attributed to prodigiosins as pH modulators, cell cycle inhibitors, DNA cleavage agents and mitogen activated protein kinase regulators.

These molecules when combined with some other anticancer agents can greatly help in fighting cancer.





Additionally, during the last few decades there has been an increasing interest in the study of medicinal plants and their traditional use in different parts of the world.

Plants generally produce many secondary metabolites which were constituted an important source of many pharmaceutical drugs.

Cupressaceae is a conifer family, the cypress family, with worldwide distribution.

A decoction of the cones and leaves of *Cupressus sempervirens* was used for treating hemorrhoids, varicose veins and venous circulation disorders, bronchitis, head colds and coughs externally.



Internally, the dried cones and young branches are used as deodorant and diuretic. anti-parasitic, antipyretic, antirheumatic, antiseptic, balsamic, vasoconstrictive, and antifungal.

Thuja orientalis has been exhibiting extensive biological activities including anticancer, antiepileptic, antiinflammatory, antibacterial, antifungal activities, hair growth-promoting, antiviral, antiallergic, antioxidant and molluscicidal.

Previously, it was reported that there are volatile oils have more than 50% larvicidal effect after 24 hours.





Essential oils are called volatile or ethereal oils, due to their evaporation when exposed to heat in contrast to fixed oils.

They are hydrophobic, and are soluble in alcohol, non-polar or weakly polar solvents.

Their density is usually lower than water, and they are oxidizable by light, heat, and air due to their molecular structures.

Essential oils are highly complex mixtures of volatile compounds containing about 20 to 60 individual constituents, and some may contain more than 100 different components such as:



Jasmine





The major volatile constituents are classified into two main categories:

Terpenoids

Phenylpropanoids

Hydrocarbons

Oxygenated Compounds



Pharmacological and medicinal importance of Essential oils:

Perfumes

Phytotherapy

Insecticides

Coronary Heart Diseases

Cosmetics

Aromatherapy

Herpes Simplex Virus

Leukemia





Novel Formula as Mosquito Larvicides

PLAN OF THE WORK

PLAN OF THE WORK

Plan 01

Isolation, Purification and Characterization of PDG from Serratia Marcescens

Plan 02

Preparation and characterization of four essential oils from fresh leaves of *Thuja orientalis* (TOL).

Plan 03

Preparation and characterization of four extracts from dried leaves of studied plants

Plan 04

Maintaining the mosquito by rearing the culture of Cx pipiens

PLAN OF THE WORK

Plan 05

Dose response bioassay separately of all preparations and dry extract.

Plan 06

Investigation for the synergistic effect of PDG with each E.O, and dry extract individually as mosquito larvicidal potential.

Plan 07

Investigating the mode of action of PDG, E.O and dry extract for mosquito larvicidal potentially.

Plan 08

Probit analysis for calculating the lethal concentration of PDG, E.O and dry E.





STUDY SETTING



Materials

Bacteria Strain Serratia Marcescens Strain

Serratia Marcescens Strain was obtained from other colleagues in the Biotechnology Department - IGSR

Plant Materials

Plant materials Fresh leaves and cones of *Thuja orientalis* were collected from Anotoniadis Botanical Garden in Alexandria.



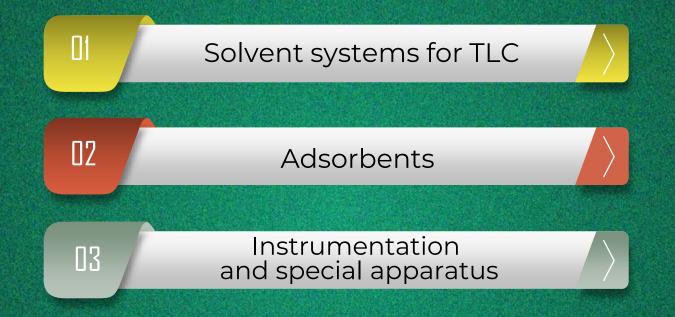
Animals and insects Larvae and Pupae

• Larvae and pupae of Culex pipiens were purchased from the Institute of Medical Insects in El-Dokki, Cairo-Egypt.

Pigeons were purchased from the market and used as a source of blood meal for the adult female insects.

Silica Gel and PDG Standard

Silica Gel and PDG standard Were obtained from Sigma Aldrich (USA)



03

COMMON CO.

Instrumentation and special apparatus



Hood

03

Instrumentation and special apparatus



Shaker incubator

Instrumentation and special apparatus



03

COMMON CO.

pH meter

03

COMMON CO.

Instrumentation and special apparatus



Autoclave

03

Lines

Instrumentation and special apparatus



Digital Balance

03

Instrumentation and special apparatus



Fermenter

03

E.11992

Instrumentation and special apparatus



GC-MS

03

Instrumentation and special apparatus

Rotary Evaporator

03

Instrumentation and special apparatus



UV-Spectrophotometer

03

E.11992

Instrumentation and special apparatus



FT-IR

03

COMMON CO.

Instrumentation and special apparatus



HPLC

03

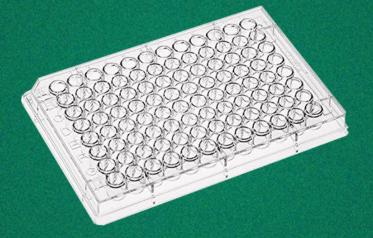
Instrumentation and special apparatus



Eppendorf Cooling Centrifuge

03

Instrumentation and special apparatus



Microplates

03

Instrumentation and special apparatus



Glass Wares, Automatic Pipettes

03

Instrumentation and special apparatus



Stereomicroscope



Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Method 01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

PY Media

500

400

300

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Method

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Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

USCO /

500

400

300

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Method 01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

PY Media

500

400

300

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Method 01

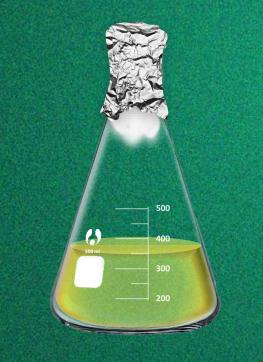
Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*



Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* **Growth and activation of** *S. marcescens*



Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

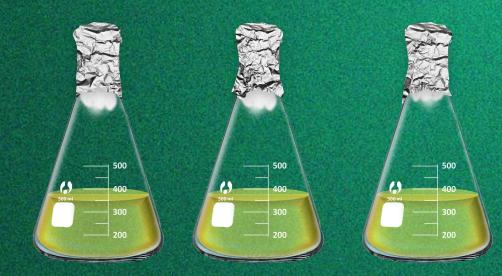




Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*







01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Peanut Media



Method 01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Peanut Media

500

400

300

6







Method Isc 01 Gr

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Peanut Media



Method 01 Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Peanut Media

500

400

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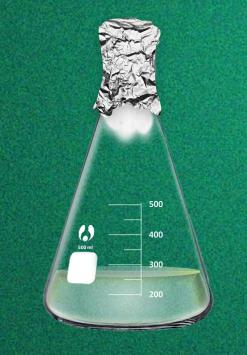




01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Peanut Media





Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

6

500 m

Peanut Media





Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*





Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*





Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*



Method

01

500

400

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Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Inoculation of PY Media

400

400

Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* **Growth and activation of** *S. marcescens*





Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Inoculation of Peanut Media







Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Inoculation of Peanut Media

500



Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens*

Inoculation of Peanut Media



Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Inoculation of Peanut Media

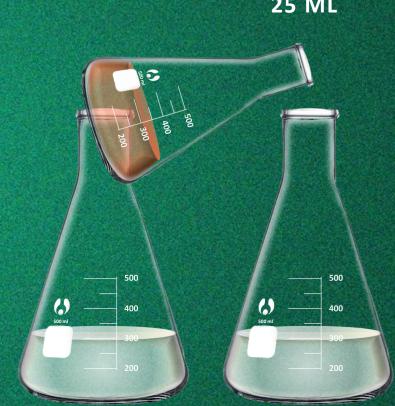


Method

01

Isolation, Purification and Characterization of PDG from Serratia Marcescens Growth and activation of S. marcescens

Inoculation of Peanut Media



25 ML

Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Inoculation of Peanut Media





Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* **Growth and activation of** *S. marcescens*

Inoculation of Peanut Media



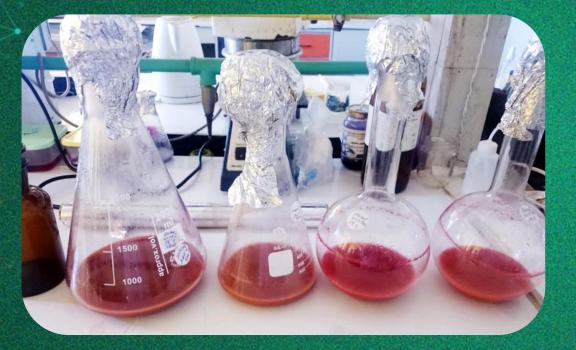


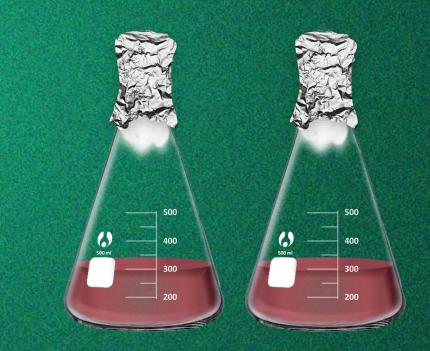


01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Red pigment







Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Growth and activation of *S. marcescens*

Fermenter



Method 01

Method

01

500

() 500 ml Isolation, Purification and Characterization of PDG from Serratia Marcescens Extraction of Prodigiosin



NaOH 100 gm

Method

01

() 500 ml 500

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Extraction of Prodigiosin



NaOH 100 gm

Method

01

Isolation, Purification and Characterization of PDG from Serratia Marcescens Extraction of Prodigiosin







Petroleum Ether

Ethanol

Method

01

500

6) 500 ml



Method

01







Method

01

500

6) 500 ml 3

Method

01





Method

01







Method

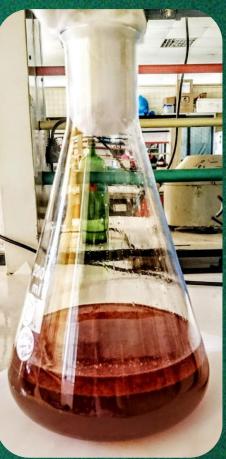
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Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Extraction of Prodigiosin

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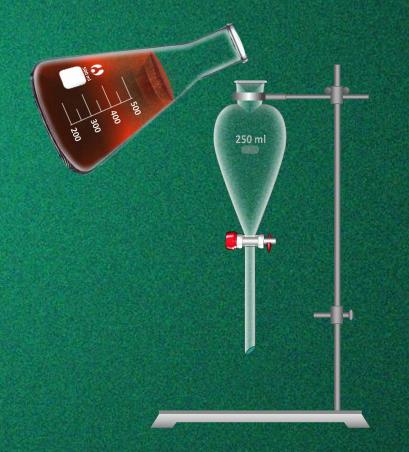
Method

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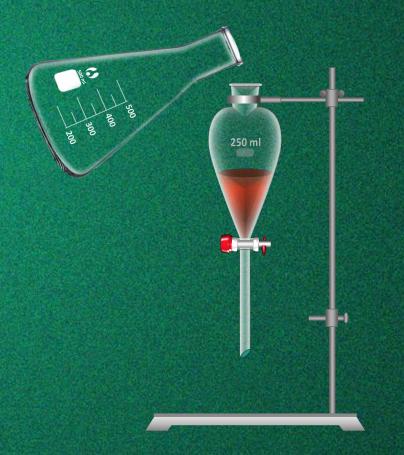
Method

01



Method

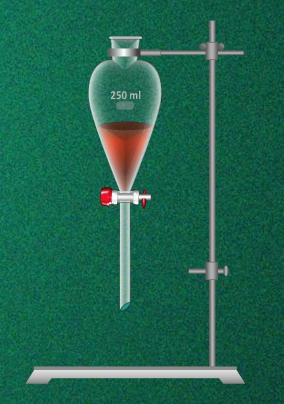
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Method

01





Method

01



Method

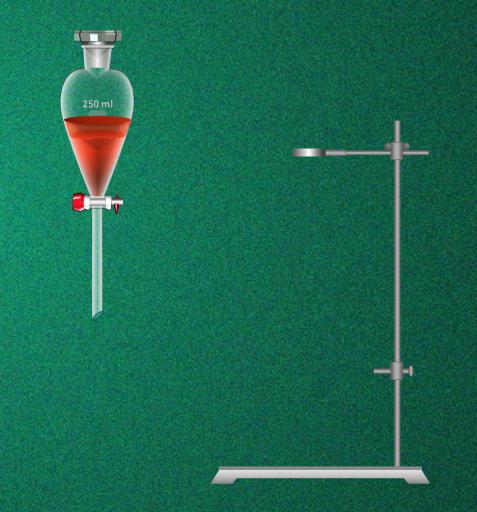
01





Method

01



Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Extraction of Prodigiosin

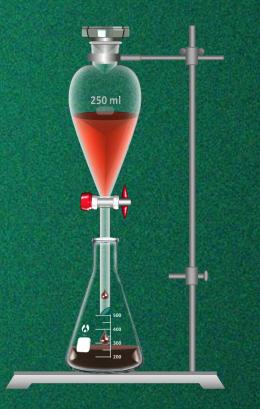
250 ml

Mode

Open

Method

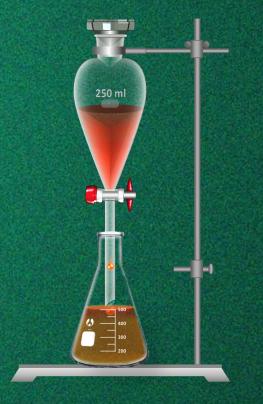
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Method

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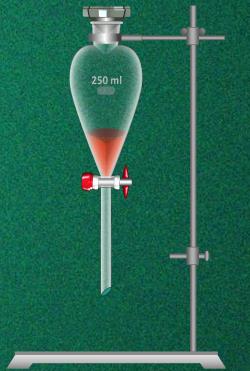




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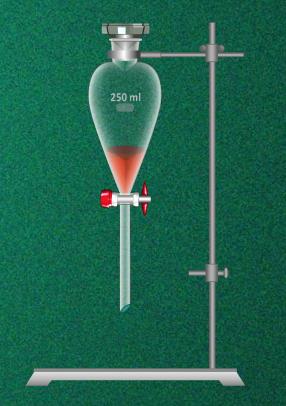






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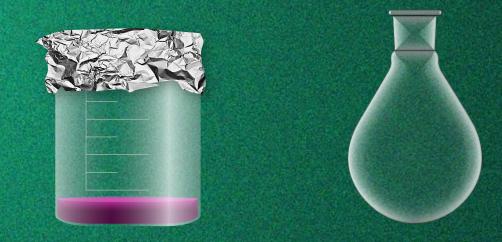


Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Extraction of Prodigiosin





Method

01

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* Extraction of Prodigiosin





Method 01

Method

01

500

Isolation, Purification and Characterization of PDG from Serratia Marcescens Purification of prodigiosin (Column Chromatography)







Isolation, Purification and Characterization of PDG from Serratia Marcescens Purification of prodigiosin (Column Chromatography)



Method

01



Method

01

500

400

0

Isolation, Purification and Characterization of PDG from *Serratia Marcescens* **Purification of prodigiosin (Column Chromatography)**





Method

01

500 400

0

Isolation, Purification and Characterization of PDG from Serratia Marcescens Purification of prodigiosin (Column Chromatography)







01

Method Isolation, Purification and Characterization of PDG from Serratia Marcescens Purification of prodigiosin (Column Chromatography)





Isolation, Purification and Characterization of PDG from Serratia Marcescens **Purification of prodigiosin (Column Chromatography)**



A

Method



Method

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Method

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500



Method

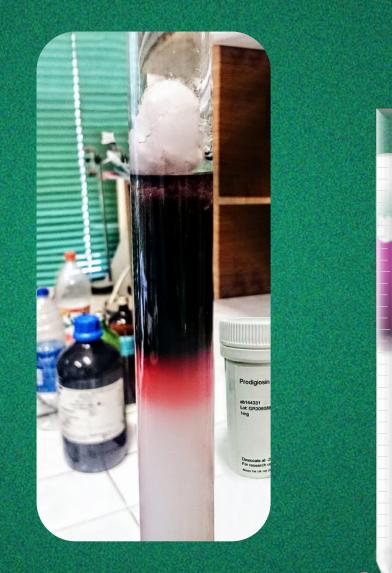
01



Method

01

Isolation, Purification and Characterization of PDG from Serratia Marcescens Purification of prodigiosin (Column Chromatography)



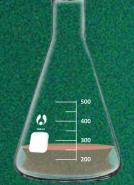
Method 01

Isolation, Purification and Characterization of PDG from Serratia Marcescens **Purification of prodigiosin (Column Chromatography)**



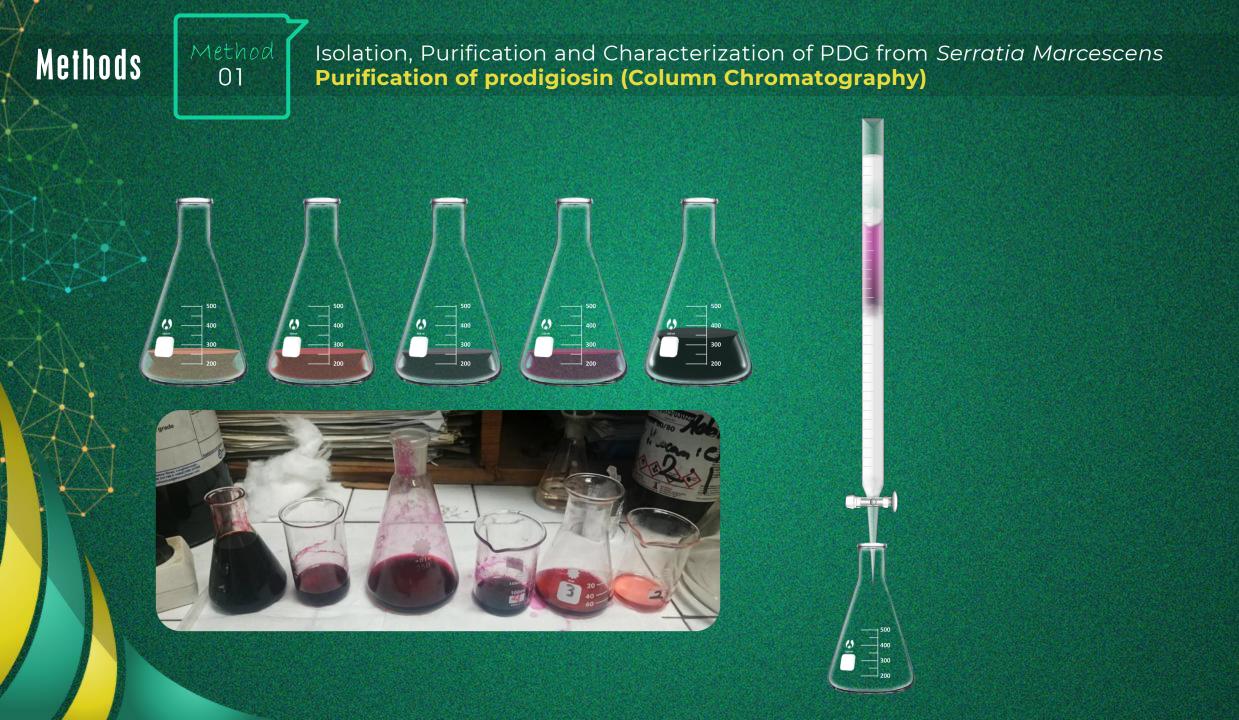
Isolation, Purification and Characterization of PDG from Serratia Marcescens Purification of prodigiosin (Column Chromatography)

500 400



Method





Method 02

Preparation of Four Essential Oils from Fresh Leaves



Preparation of Four Essential Oils from Fresh Leaves



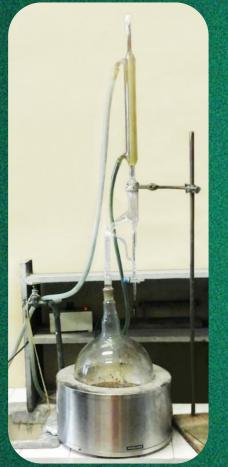
T. orientalis

Leaves

Method

02

Preparation of Four Essential Oils from Fresh Leaves



Steam Distillation of leaves



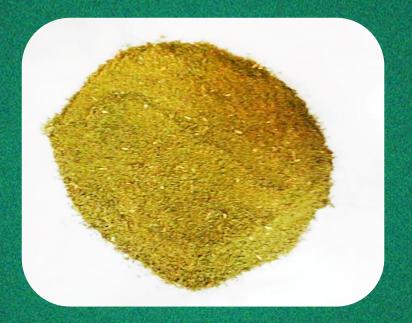
Condensation of the oil

Method Preparation of 03

Preparation of Four Extracts from Dried Leaves of Studied Plant

Method F

Preparation of Four Extracts from Dried Leaves of Studied Plant



Grinding of the dried leaves



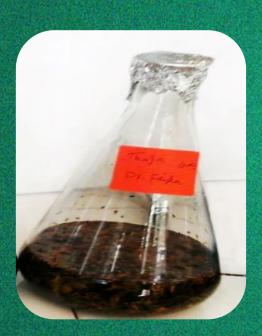
Preparation of Four Extracts from Dried Leaves and Cones of Studied Plants



Method 03

Preparation of Four Extracts from Dried Leaves and Cones of Studied Plants







Method

03

Preparation of Four Extracts from Dried Leaves and Cones of Studied Plants Characterization



Method 04

Maintaining the Mosquito by Rearing the Culture of Culex Pipiens

Method M

Maintaining the Mosquito by Rearing the Culture of Culex Pipiens







Method 05

Dose Response Bioassay Separately of the all Preparations, Extracts and Chemical Insecticide





50 ml Dechlorinated Water + 10 Larvae

Each control and Treatment were Replicated Three Times 0, 20, 30, 40, 50 and 60 ppm PDG





0, 120, 140, 160, 180 & 200 ppm E



Dose Response Bioassay Separately of the all Preparations, and Extract





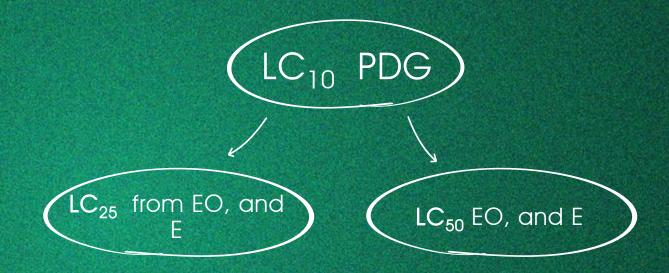


Investigation for the synergistic effect of prodigiosin with essential oil, and extract individually as mosquito larvicidal potential



Method Investigation for the synergistic effect of prodigiosin with essential oil, and extract individually as mosquito larvicidal potential

50 ml Dechlorinated Water + 10 Larvae



The mortality of the larvae was recorded after an hour, 3, 6,12, 24, 48, 72 and 96 hours



Method Investigating the Mode of Action of PDG, Essential Oil and Extract for Mosquito Larvicidal Potentially **Preparation of 'Whole Body Homogenates'**

Investigating the Mode of Action of PDG, Essential Oil and Extracts for Mosquito Larvicidal Potentially **Preparation of 'Whole Body Homogenates'**



02

Method

07

- Untreated Larvae (20-30 larvae) with:
- Treated Larvae (20-30 larvae) with:
 - **Prodigiosin at LC10** concentration
 - **Essential oil, and extracts treated larvae** at the LC₂₅ concentration
 - **Essentia oil, and extract treated larvae** at the LC₅₀ concentration
 - **PDG Treated** at LC_{10} in combination with EO as well as in combination with extract treated at LC_{25} and LC_{50}

After 24 hour

Were Washed with Distilled Water to Remove the Adhering Water

Method Investigating the Mode of Action of PDG, Essential Oil and Extract for Mosquito Larvicidal Potentially **Preparation of 'Whole Body Homogenates'**



07

Treated Larvae (20-30 larvae) with:



The larvae then pooled and homogenized in Eppendorf tubes (held in ice) using a Teflon hand homogenizer in 1 ml of 0.9% w/v saline for eventual estimation of total protein, acetylcholine esterase.



Homogenates were centrifuged at 4,000 rpm for 15 min at 4 °C in a cooling centrifuge



The clear supernatants were kept at -80 °C until use for biochemical analysis

07

Method Investigating the Mode of Action of PDG, Essential Oil and Extract for Mosquito Larvicidal Potentially **Preparation of 'Whole Body Homogenates'**

12112

eppendorf

Eppendorf Cooling Centrifuge

07

Method Investigating the Mode of Action of PDG, Essential Oil and Extract for Mosquito Larvicidal Potentially **Preparation of 'Whole Body Homogenates'**



Anticholinesterase Activity

Total Protein Activity

Methods

07

Method Investigating the Mode of Action of PDG, Essential Oil and Extract for Mosquito Larvicidal Potentially pH Determination by Bromothymol Blue Dye

> pH determination in the 3rd Stage Larval Midgut

Methods

Method 08

Probit Analysis for Calculating the Lethal Concentration of PDG, Essential Oils and Dry Extract Methods

Method

80

Probit Analysis for Calculating the Lethal Concentration of PDG, Spinosad Essential Oils and Dry Extracts

Mortality Rate was Calculated using Abbott's Formula

The dose-response data was Analyzed by Probit Regression

The Lethal Concentrations in ppm (LC₁₀, LC₂₅ and LC₅₀), and the **95% confidence** intervals [upper confidence limit (UCL) and lower confidence limit (LCL) were Calculated

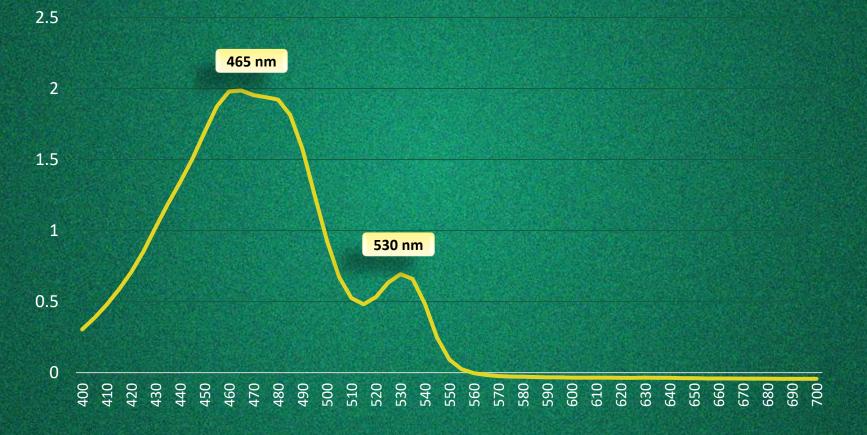






Results

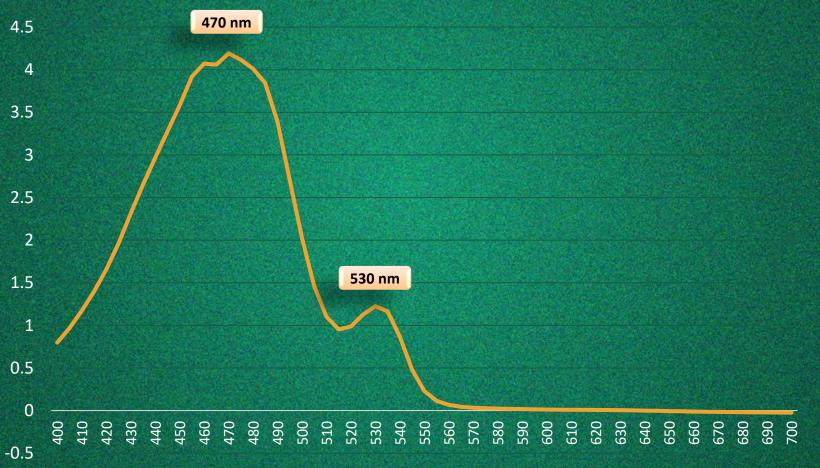
Shows the UV-visible spectrophotometry in the range 400-700 nm for the crude PDG



A) PDG-batch Scale

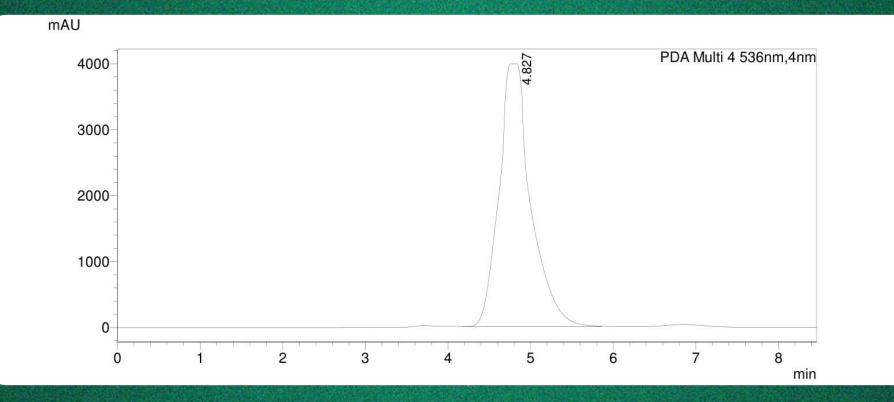
-0.5

Shows the UV-visible spectrophotometry in the range 400-700 nm for the crude PDG



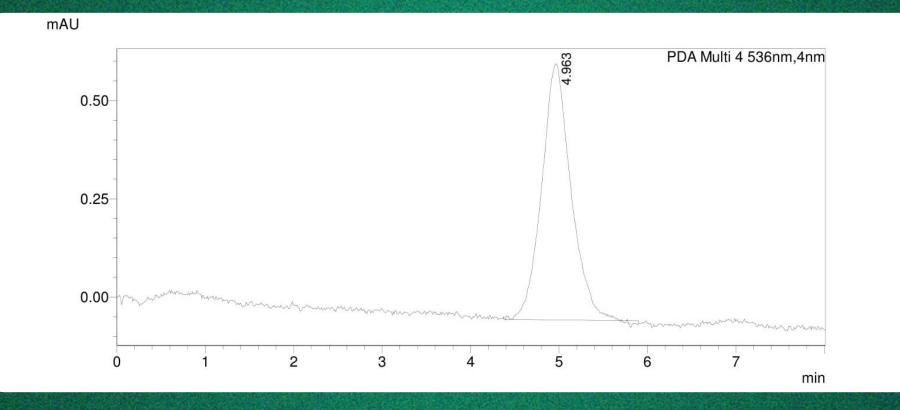
B) PDG-Fermenter (Bioreactor)

HPLC for the prepared and the standard PDG



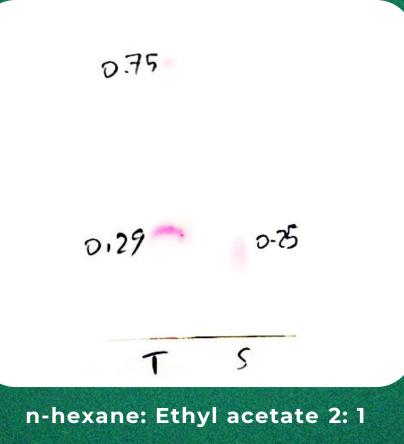
A) Purified Red Pigment

HPLC for the prepared and the standard PDG

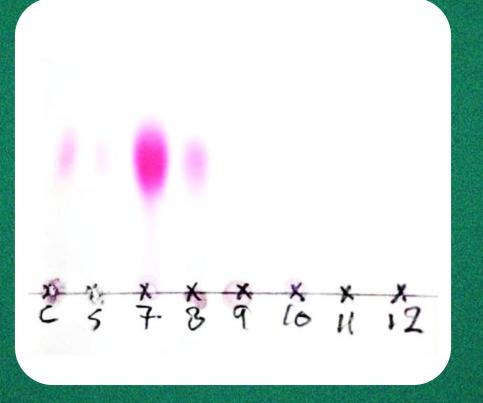


B) Standard PDG

Trials of TLC application to determine the best mobile phase

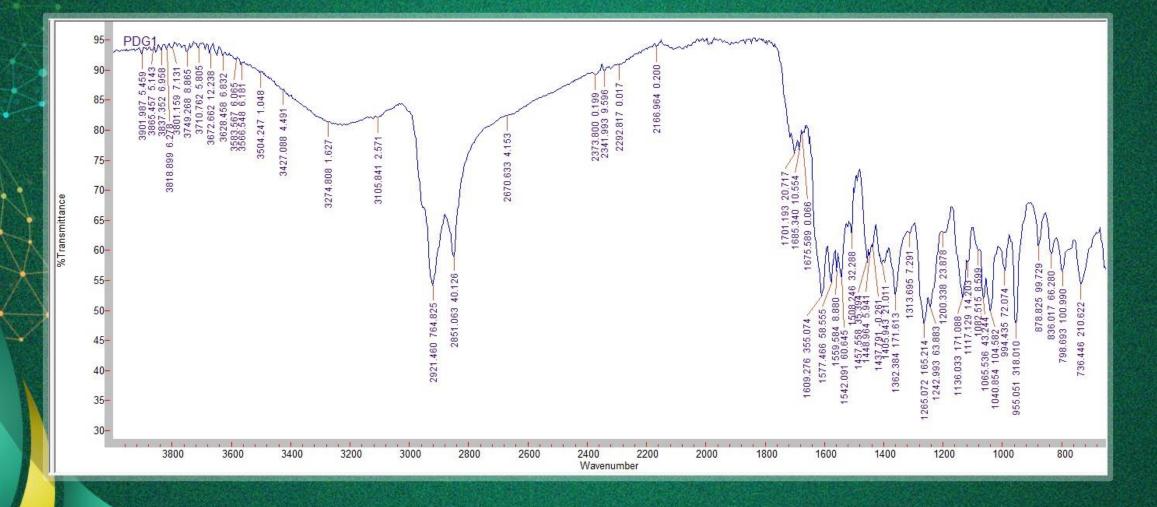


Application of the purified PDG fractions and the standard on TLC



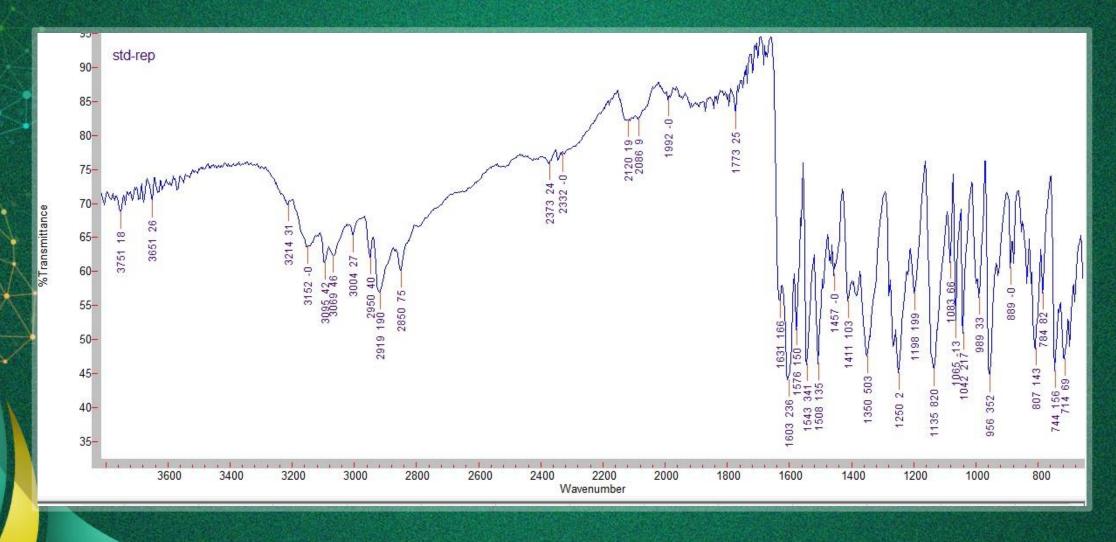
n-hexane: Ethyl acetate 2:1

• FT-IR analysis of the purified Red Pigment



Pigment

FT-IR analysis of the PDG Standard



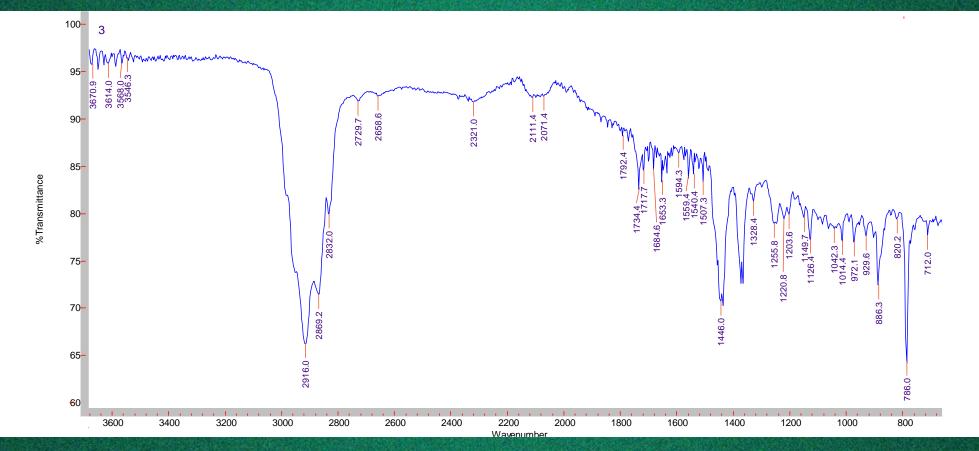
Standard

Preparation and Characterization of Four Essential Oils from Fresh Leaves and Cones of Studied Plants

Results



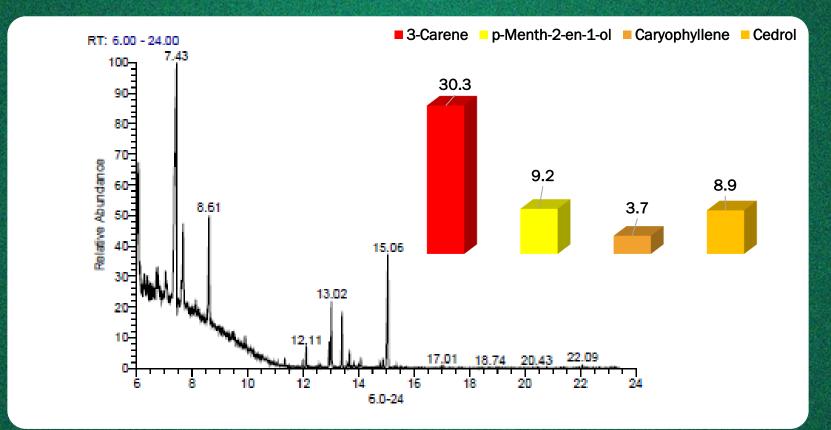
FT-IR Spectra



FT-IR analysis for the Essential Oil Prepared from Fresh Leaves

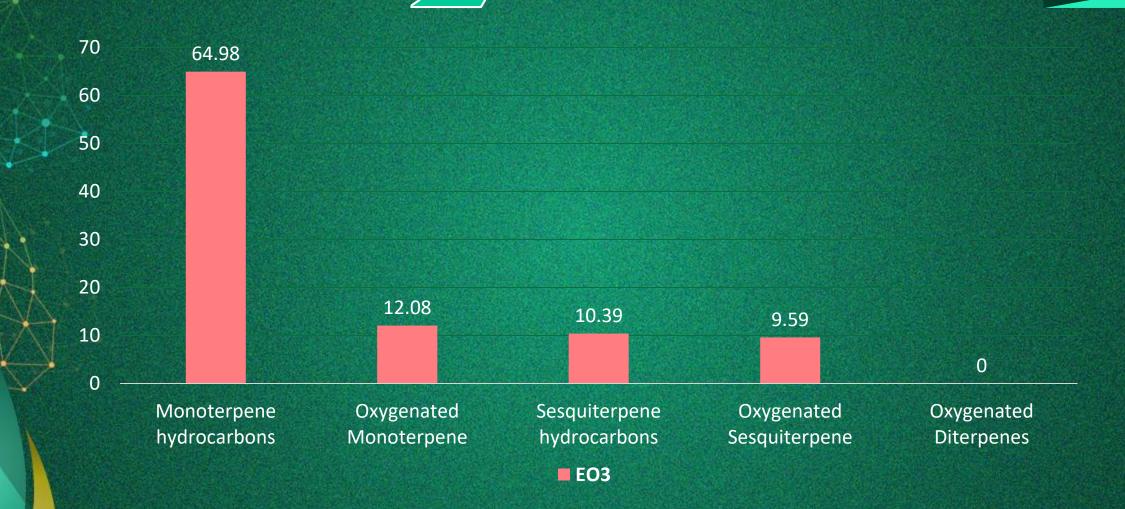
Preparation and Characterization of Four Essential Oils from Fresh Leaves of *Thuja orientalis*

GC-MS

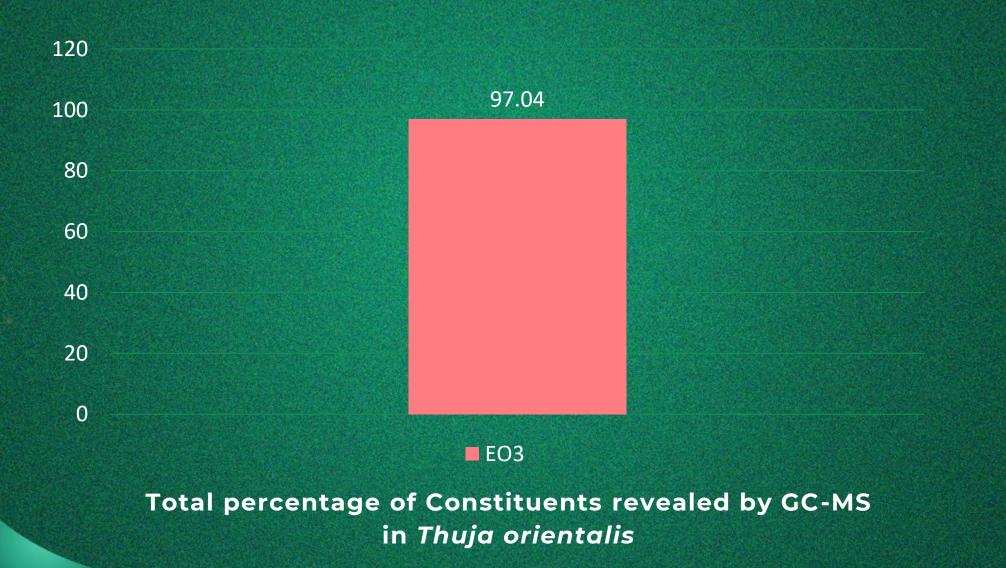


GC-MS analysis for the essential oil prepared from fresh leaves of *Thuja orientalis*

⁷ Preparation and Characterization of Four Essential Oils from Fresh Leaves of *Thuja orientalis*



GC-MS analysis for the essential oil prepared from fresh leaves of Thuja orientalis Preparation and Characterization of Four Essential Oils from Fresh Leaves of *Thuja orientalis*



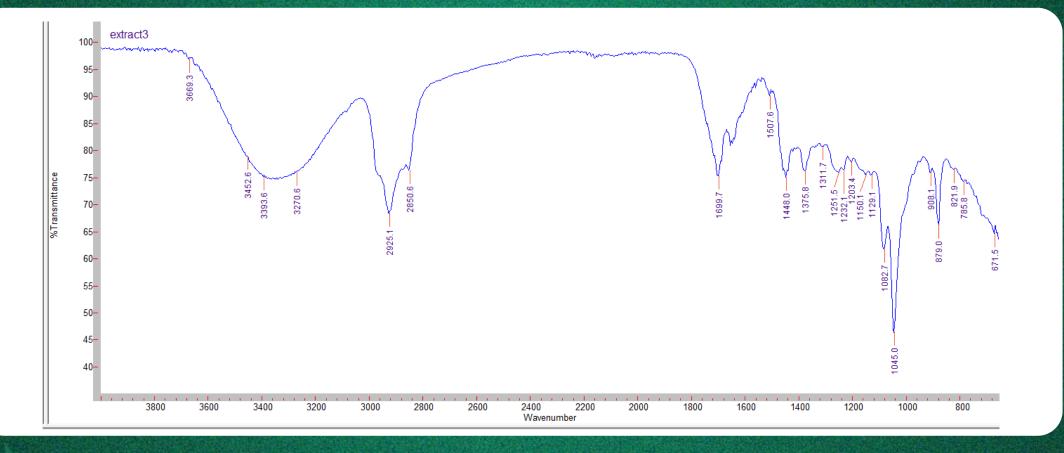
Preparation and Characterization of Four Crude Extracts from Dried Leaves of Studied Plants

Results



Preparation and Characterization of Four Crude Extracts from Dried Leaves of Studied Plants

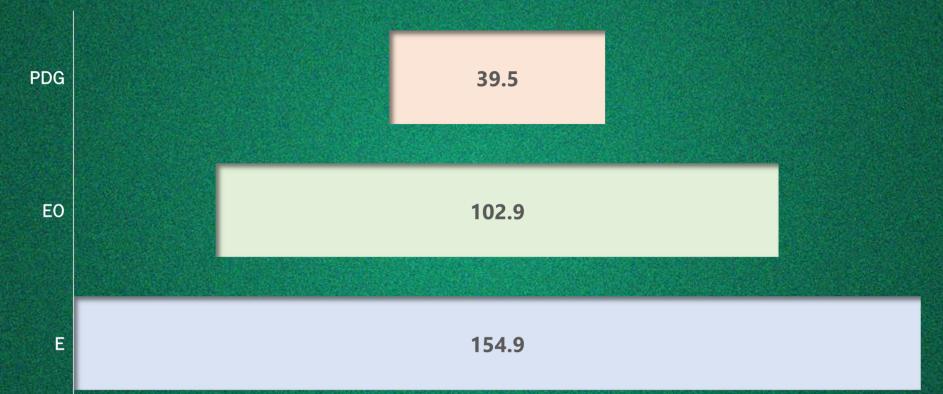
FT-IR analysis for the extract of dry leaves of *Thuja orientalis* (E3)





Results





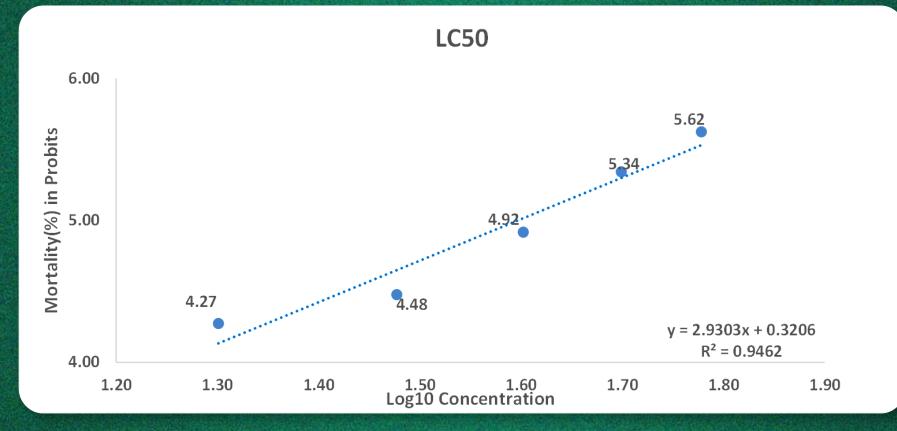
Larvicidal activity of the studied preparations after 24 hours against the 3rd larval stage of Cx. pipiens





Dead Larvae after Treatment with PDG & E.O



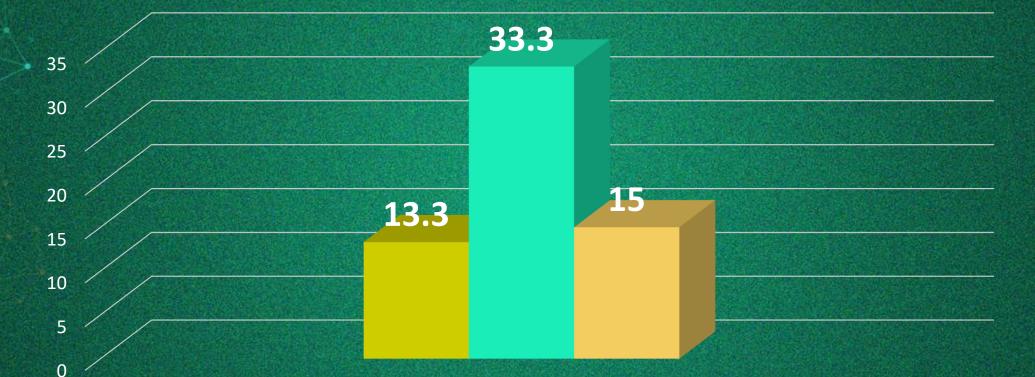


The concentration of PDG at LC50 after 24 Hours

Synergistic Effect of PDG with the natural Pds Individually as a Mosquito Larvicidal Potential after 24 hrs.

Results

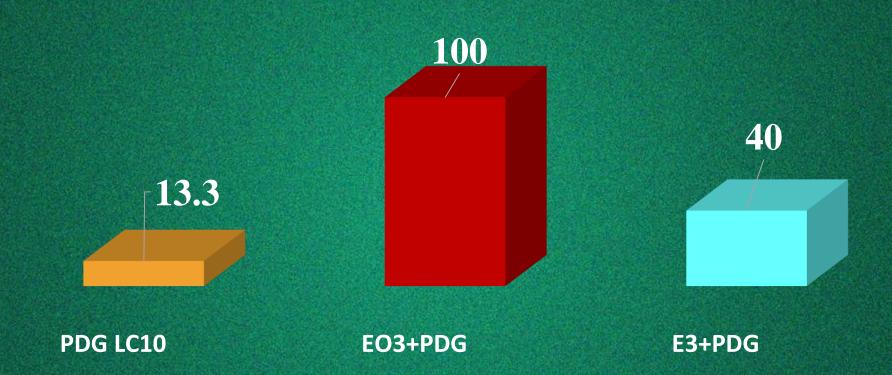




PDG LC10 O3+PDG E3+PDG

Synergistic Larvicidal activity of the LC₁₀ of PDG with LC₂₅ of Oil, and the extract Larvicide **(Spinosad)** after 24 hrs.

Synergistic Effect of PDG with the natural Pds Individually as a Mosquito Larvicidal Potential after 24 hrs.

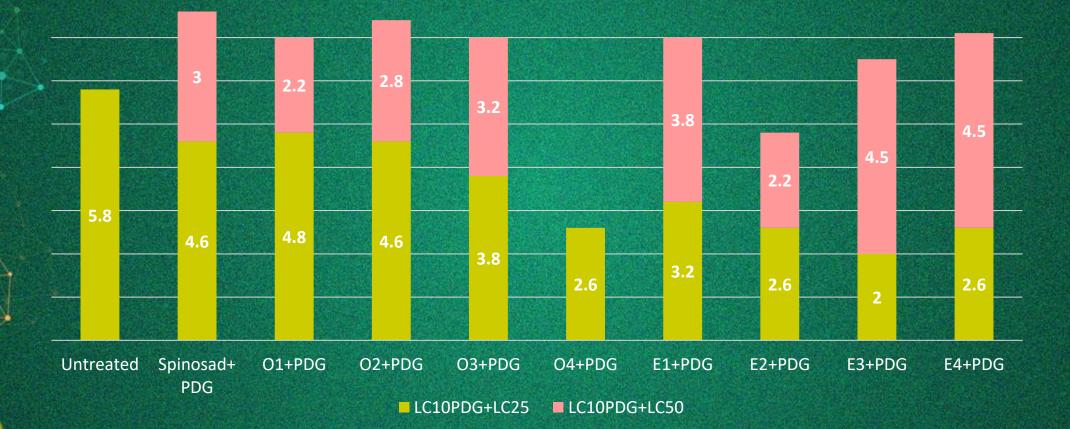


Synergistic Larvicidal activity of the LC₁₀ of PDG with LC₅₀ of Oils, extracts and the Chemical Larvicide **(Spinosad)** after 24 hrs.

Investigating the Mode of Action of PDG, Essential Oil, and the extract for Mosquito Larvicidal Potentially

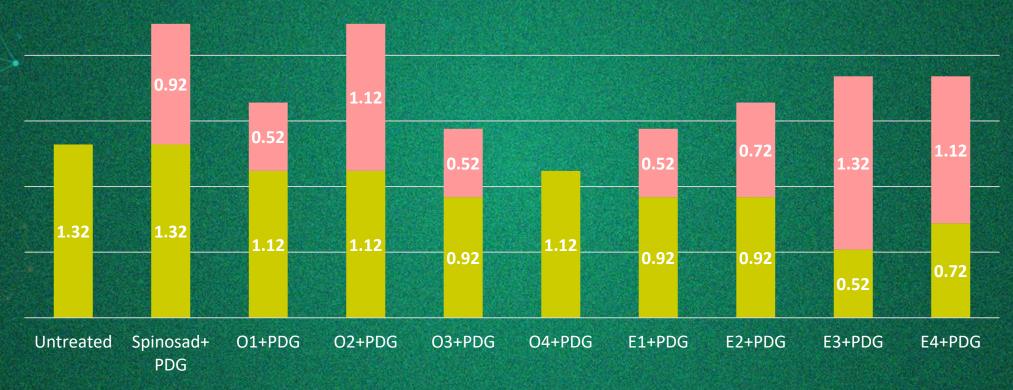
Results

Investigating the Mode of Action of PDG, Essential Oil, and extracts for Mosquito Larvicidal Potentially



AChE Arbitrary Activity unit/gm Tissue (%)

Investigating the Mode of Action of PDG, Volatile Oils, extracts and Spinosad for Mosquito Larvicidal Potentially



■ LC10PDG+LC25 ■ LC10PDG+LC50

Total Protein in mg /gm tissue $(\%)^3$





LC10PDG+LC25 LC10PDG+LC50

AChE Arbitrary Specific Activity² (%)³

Investigating the Mode of Action of PDG, Essential Oil, and extract for Mosquito Larvicidal Potentially

pH Indicator



Midgut of Untreated 3rd Larval Stage showed alkaline media by using Bromothymol blue dye that act as pH indicator **(1.6 X**) Investigating the Mode of Action of PDG, Essential Oil, and extract for Mosquito Larvicidal Potentially

pH Indicator



Midgut of PDG Treated 3rd Larval Stage showed acidic media by using Bromothymol blue dye that act as pH indicator (1.6 X) Investigating the Mode of Action of PDG, Essential Oil, and extract for Mosquito Larvicidal Potentially

pH Indicator



Midgut of E.O Treated 3rd Larval Stage showed acidic media by using Bromothymol blue dye that act as pH indicator (1.6 X) Investigating the Mode of Action of PDG, Essential Oils and extract for Mosquito Larvicidal Potentially

pH Indicator

Midgut of PDG+E.O Treated 3rd Larval Stage showed acidic media by using Bromothymol blue dye that act as pH indicator (1.6 X)

1 mm



Conclusion Recommendation

CHAPTER FIVE

Purified rodigiosin showed the lowest LC₅₀ followed by crude essential oils of *Thuja orientalis* leaves and its **Crude extract**

Π

02

The combination between LC₁₀ of prodigiosin and LC₅₀ of *T. orientalis* E.O, showed the highest synergistic effect (100%) The treated 3rd larval *Cx. pipiens* showed reduction in the acetylcholine esterase and total protein content as compared to the **untreated ones**

03

04

05

The midgut of the treated 3rd larval Cx. pipiens showed acidic medium in contrast the untreated ones showed alkaline medium by stereomicroscope after using bromothymol blue dye

The present study recommended that the combination of prodigiosin and E.O. from **T. orientalis** leaves is a promising potent larvicid against the 3rd larval **Cx. pipiens.**

