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Antioxidant and antihypertensive properties from muscle hydrolysates of farm rainbow trout ⁺

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Abstract: Fish proteins are a promising source for multifunctional bioactive peptide production.19Thus, this work aimed to establish the potentiality of rainbow trout muscle protein for the obtention20of antioxidant and antihypertensive hydrolysates. Alcalase application produced a hydrolysate21with inhibition of angiotensin-converting enzyme equivalent to $56.43\pm2.05\%$ after one hour. The22same hydrolysate exhibited a scavenging and ferric reducing power of $2.65\pm0.07 \mu$ M Trolox equivalents, respectively. Results showed that rainbow trout muscle could24be essential for identifying peptides with bifunctional properties.25

Keywords: rainbow trout; hydrolysates; antioxidant; antihypertensive

1. Introduction

Aquaculture has become a potential strategy for worldwide fishery production. Its 29 growth was slow from 2009, but just in 2018, a historical record was achieved, producing 30 81.1 million tons [1,2]. In this sense, farming finfish has taken such relevance because, in 31 some countries, species are produced highly by this method instead of by fishing. Also, 32 finfish farming represents 90% of total fishery production, where the grass and silver carp, 33 like the Nile tilapia, are the highest farming species [2]. In Mexico, the main farming specie 34 is tilapia. Still, specifically in Hidalgo, a state located in the center of the country with 35 temperate weather, rainbow trout has been considered an emerging farm finfish with hu-36 man feeding and tourism aims [3]. 37

Fish proteins have been recognized as a valuable source for antioxidant and antihypertensive hydrolysates generation, with salmon, tuna, and tilapia as actual examples. In contrast, trout have been a freshwater fish with short exploration to produce hydrolysates with these functions [4]. Indeed, research in the bioactive peptides obtention from rainbow trout has been mainly oriented to using by-products [5-9]. Contrary, the investigation related to muscle exploitation has been scarce [10].

Cardiovascular diseases are a severe public health problem because they are the most common non-communicable disease in the world. It estimates that in the year 2030, there will be 22.2 million cases [11]. Nowadays, a novel approach for cardiovascular disease 46

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Copyright: © 2022 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/). treatment has been proposed, where the application of multifunctional peptides is highly 1 desirable to attend to different pathologies simultaneously [12]. Thus, the objective of the 2 present work was to determine the potentiality for the obtention of hydrolysates with both 3 antioxidant and antihypertensive activities from rainbow trout muscle. 4

2. Methods

Sample obtention: Rainbow trout (*Oncorhynchus mykiss*) specimens were acquired from a local Huasca de Ocampo, Hidalgo, Mexico farm. The sample was eviscerated and muscle processed to obtain a homogenized paste, which was used in protein determination and freeze-dried at -43°C and 286X10⁻³ mbar to generate systems for the enzymatic reaction.

Protein determination: Protein content in trout homogenized paste was realized through the Kjeldahl method [13].

Enzymatic hydrolysis: Hydrolysis systems were prepared as suspensions at 5% (w/v) 13 in Tris-HCl buffer (pH=9) from lyophilized trout paste. Afterward, protein trout was hydrolyzed at 55°C with alcalase, adding a mass ratio of 100:10 (soluble protein: enzyme) 15 [14]. Sampling hydrolysis times were 0,1 and 2 h, stopping enzymatic reaction by boiling 16 water treatment for 10 min. Respective supernatants were frozen and used to free amino 17 groups, antioxidant and antihypertensive determinations. 18

Hydrolysis degree determination: Free amino groups from rainbow trout hydrolysates were determined with the trinitrobenzenesulfonic acid method [15]. Results were expressed as a glycine mM concentration.

Antioxidant capacity analysis: Radical scavenging and ferric reduction power was determined by DPPH and FRAP techniques, respectively [16]. Results were expressed as concentrations of Trolox (μ M) and Fe²⁺ (mM) equivalents.

Antihypertensive evaluation: The *in vitro* assay for the Angiotensin Converting Enzyme inhibition was used to evaluate antihypertensive properties, measuring hippuric acid produced by spectrophotometry [17].

3. Results and Discussion

Protein content in homogenized muscle paste: The paste analysis obtained from rain-29 bow trout muscle showed a 17.87±0.31% protein content. The last value was found within 30 the range found by other studies [13,18]. Specifically, the protein determined in this work 31 was equivalent to that reported by Craft et al. [18], where trout specimens were fed with 32 a novel formulation composed mainly of poultry by-products, wheat flour, and menha-33 den fish oil. In contrast, Cano-Estrada et al. [13] found a protein content of 19.46±0.78% 34 for rainbow trout from the exact location of the specimens used in this study. Differences 35 could be linked to the feeding source because Cano-Estrada et al. [13] reported that their 36 samples were fed with commercial fish food, suggesting that the trout diet is not the same 37 in the farms from Huasca de Ocampo. Nonetheless, protein content did not fluctuate 38 highly and, in all cases, could be used as a potential source for bioactive peptide obtention. 39

Hydrolysis degree, antioxidant and antihypertensive properties: As observed in table401, free amino groups increased with higher hydrolysis time, as expected. Hydrolysis41shown at the beginning of the performance is associated with the normal autolysis carried42out by fish muscle. Indeed, that procedure has also been tested as an alternative for bio-43active peptide production from trout by-products [18].44

On the other hand, antioxidant properties were found in the control time, but antihypertensive activity was not detected. The last property was identified until one hour 46 after hydrolysis and decreased with an additional hour. Also, the results showed that 47 while antihypertensive activity is deleted by prolonged hydrolysis times, ferric reducing 48 power is increased, especially during the first hour of hydrolysis. In the case of scavenging 49 activity, it was lost within the first hour but recovered and increased during the second 50

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hydrolysis hour. Results are linked with the alcalase specificity for cutting action, which hydrolyzes preferentially peptide bonds containing aromatic amino acid residues [20].

Hydrolysis time (h)	degree	Radical scavenging properties (µM Trolox equiv- alents)	Ferric reducing power (mM Fe ²⁺)	Angiotensin-con- verting enzyme in- hibition (%)
0	16.18±0.30	3.31±0.12	24.46±0.08	ND
1	19.76±0.59	2.65±0.07	32.12±0.98	56.43±2.05
2	32.64±0.37	3.68±0.09	33.98±0.75	2.14±0.04

Table 1. Hydrolysis degree and bioactive properties from trout muscle hydrolysates .

ND: Not detected

As the novel investigation is focused on the search for multifunctional peptides [12], 5 in this work, the first hydrolysis hour was the best time to achieve that aim, obtaining both 6 antioxidant and antihypertensive activities. Comparing the activities found with other 7 works [6,10], the values obtained here are highly competitive, but with the additional ben-8 efit that the hydrolysate obtained showed bifunctional activity.

4. Conclusion

Rainbow trout muscle protein showed high potentiality to generate hydrolysates 11 with bifunctional bioactivity, where the best time for the obtention of as antioxidant as 12 antihypertensive capacities was after one hour of hydrolysis. In the same way, the ex-13 posed results represent a promising beginning for developing novel alternatives in cardi-14 ovascular treatment.

Supplementary Materials: No supplemental material is presented.

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References

- Rocha, C.P.; Cabral, H.N.; Marques, J.C.; Gonçalves, A.M. A Global Overview of Aquaculture Food Production with a Focus on 1. 30 the Activity's Development in Transitional Systems - The Case Study of a South European Country (Portugal). J. Mar. Sci. Eng. 31 (2022), 10, 417. https://doi.org/10.3390/jmse10030417 32
- 2. FAO. The State of World Fisheries and Aquaculture 2020. Sustainability in action, 1st ed.; FAO: Rome, Italy, 2020; pp. 21-30. 33 https://doi.org/10.4060/ca9229en 34
- Reves-Vera, A.M.; Román-Gutiérrez, A.D.; Guzmán-Ortiz, F.A.; Pulido-Flores, G.; Velasco-Amaro, P.I. An Overview of Aqua-3. culture Activity in Hidalgo State. In Water Availability and Management in Mexico, 1st ed.; Otazo-Sánchez, E.M.; Navarro-Frometa, A.E.; Singh, V.P., Eds.; Springer: Switzerland, 2020: Volume 999, pp. 267-286. https://doi.org/10.1007/978-3-030-24962-5
- 4 Korczek, K.; Tkaczewska, J.; Migdał, W. Antioxidant and antihypertensive protein hydrolysates in fish products-a review. Czech 38 J. Food Sci. (2018), 36, 195-207. https://doi.org/10.17221/283/2017-CJFS 39

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- Yaghoubzadeh, Z.; Peyravii Ghadikolaii, F.; Kaboosi, H.; Safari, R.; Fattahi, E. Antioxidant activity and anticancer effect of bioactive peptides from rainbow trout (*Oncorhynchus mykiss*) skin hydrolysate. *Int. J. Pept. Res.* (2020), 26, 625-632. https://doi.org/10.1007/s10989-019-09869-5
- 6. Ramezanzade, L.; Hosseini, S.F.; Nikkhah, M.; Arab-Tehrany, E. Recovery of bioactive peptide fractions from rainbow trout (*Oncorhynchus mykiss*) processing waste hydrolysate. *Ecopersia*. (2018), *6*, 31-40.
- Ketnawa, S.; Suwal, S.; Huang, J.Y.; Liceaga, A.M. Selective separation and characterisation of dual ACE and DPP-IV inhibitory peptides from rainbow trout (*Oncorhynchus mykiss*) protein hydrolysates. *Int. J. Food Sci.* (2019), 54, 1062-1073. https://doi.org/10.1111/ijfs.13939
- 8. Vásquez, P.; Zapata, J.E.; Chamorro, V.C.; Fillería, S.F.G.; Tironi, V.A. Antioxidant and angiotensin I-converting enzyme (ACE) inhibitory peptides of rainbow trout (*Oncorhynchus mykiss*) viscera hydrolysates subjected to simulated gastrointestinal digestion and intestinal absorption. *LWT*. (2022), *154*, 112834. https://doi.org/10.1016/j.lwt.2021.112834
- 9. Nikoo, M.; Benjakul, S.; Yasemi, M.; Gavlighi, H.A.; Xu, X. Hydrolysates from rainbow trout (*Oncorhynchus mykiss*) processing by-product with different pretreatments: Antioxidant activity and their effect on lipid and protein oxidation of raw fish emulsion. *LWT*. (2019), *108*, 120-128. https://doi.org/10.1016/j.lwt.2019.03.049
- 10. Kim, S.R.; Byun, H.G. The novel angiotensin I converting enzyme inhibitory peptide from rainbow trout muscle hydrolysate. *Fish Aquatic Sci.* (2012), *15*, 183-190. https://doi.org/10.5657/FAS.2012.0183
- 11. Şahin, B; İlgün, G. Risk factors of deaths related to cardiovascular diseases in World Health Organization (WHO) member countries. *Health Soc. Care Community.* (2022), 30, 73-80. https://doi.org/10.1111/hsc.13156
- 12. Lammi, C.; Aiello, G.; Boschin, G.; Arnoldi, A. Multifunctional peptides for the prevention of cardiovascular disease: A new concept in the area of bioactive food-derived peptides. *J. Funct. Foods.* (2019), *55*, 135-145. https://doi.org/10.1016/j.jff.2019.02.016
- 13. Cano-Estrada, A.; Castañeda-Ovando, A.; Ramírez-Godinez, J.; Contreras-López, E. Proximate and fatty acid composition in raw and cooked muscle tissue of farmed rainbow trout (*Oncorhynchus mykiss*) fed with commercial fishmeal. *J. Food Process. Preserv.* (2018), 42, e13674. https://doi.org/10.1111/jfpp.13674
- 14. Sbroggio, M.F.; Montilha, M.S.; Figueiredo, V.R.G.D.; Georgetti, S.R.; Kurozawa, L.E. Influence of the degree of hydrolysis and type of enzyme on antioxidant activity of okara protein hydrolysates. *Food Sci. Technol.* (2016), 36, 375-381. https://doi.org/10.1590/1678-457X.000216
- 15. Adler-Nissen, J. Determination of the degree of hydrolysis of food protein hydrolysates by trinitrobenzenesulfonic acid. *J. Agric. Food Chem.* (1979), 27, 1256-1262. https://doi.org/10.1021/jf60226a042
- Ramírez-Godínez, J.; Jaimez-Ordaz, J.; Castañeda-Ovando, A.; Añorve-Morga, J.; Salazar-Pereda, V.; González-Olivares, L.G.; Contreras-López, E. Optimization of physical conditions for the aqueous extraction of antioxidant compounds from ginger (*Zingiber officinale*) applying a box-Behnken design. *Plant Foods Hum. Nutr.* (2017), 72, 34-40. https://doi.org/10.1007/s11130-016-0582-1
- Sebastián-Nicolas, J.L.; Contreras-López, E.; Ramírez-Godínez, J.; Cruz-Guerrero, A.E.; Rodríguez-Serrano, G.M.; Añorve-Morga, J.; Jaimez-Ordaz, J.; Castañeda-Ovando, A.; Pérez-Escalante, E.; Ayala-Niño, A.; González-Olivares, L.G. Milk fermentation by *Lacticaseibacillus rhamnosus* GG and *Streptococcus thermophilus* SY-102: Proteolytic profile and ace-inhibitory activity. *Fermentation* (2021), 7, 215. https://doi.org/10.3390/fermentation7040215
- 18. Craft, C.D.; Ross, C.; Sealey, W.M.; Gaylord, T.G.; Barrows, F.T.; Fornshell, G.; Myrick, C.A. Growth, proximate composition, and sensory characteristics of Rainbow Trout *Oncorhynchus mykiss* consuming alternative proteins. *Aquaculture* (2016), 459, 223-231. https://doi.org/10.1016/j.aquaculture.2016.03.039
- 19. Nikoo, M.; Regenstein, J.M.; Noori, F.; Gheshlaghi, S.P. Autolysis of rainbow trout (*Oncorhynchus mykiss*) by-products: Enzymatic activities, lipid and protein oxidation, and antioxidant activity of protein hydrolysates. *LWT*. (2021), 140, 110702. https://doi.org/10.1016/j.lwt.2020.110702
- 20. Cui, Q.; Sun, Y.; Zhou, Z.; Cheng, J.; Guo, M. Effects of enzymatic hydrolysis on physicochemical properties and solubility and bitterness of milk protein hydrolysates. *Foods* (2021), *10*, 2462. https://doi.org/10.3390/foods10102462
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