



Proceeding Paper Myco-Chemical Constituents and Anti-Inflammatory Activity of *Terfezia claveryi* Chatin from Algeria ⁺

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Abstract: Mushrooms are receiving particular attention as a new source of valuable biotherapeutics. The aim of the current study is the valorization of *Terfezia claveryi* Chatin from Algeria. The mycochemical constituents, polyphenol, flavonoid and condensed tannin composition and the in vitro anti-inflammatory activity was examined using the heat denaturation protein inhibition method. Myco-chemical tests presented a very interesting richness in terms of secondary metabolites, the polyphenol, flavonoid and condensed tannin contents of the hydro-methanol extract were respectively: $82.27 \pm 1.44 \ \mu g \text{ GAE/mg}$, $14.94 \pm 0.98 \ \mu g \text{ CE/mg}$, $27.50 \pm 2.50 \ \mu g \text{ CE/mg}$. The extract at the 1.5 mg/mL level showed an inhibition of denaturation $83.53 \pm 1.57\%$ compared for that the diclofenac sodium $98.43 \pm 0.52\%$. This research revealed its interesting anti-inflammatory properties, which confirms its value in traditional use.

Keywords: Terfezia claveryi; myco-chemical constituents; anti-inflammatory activity

1. Introduction

Truffles are a naturally-occurring foodstuff, renowned for its beneficial effects on health as an excellent bio-source. The desert truffles are hypogeous fruiting bodies, symbiotic fungi of ascomycetes found in arid and semi-arid regions, widely used in traditional medicine for their nutritional richness and therapeutic effects [1–3]. Due to their content of protein, vitamins, minerals and dietary fiber, truffles have a particularly high nutritive value [4]. In recent years, truffle secondary metabolites have received considerable attention for their anti-inflammatory, antioxidant, antimicrobial and antimutagenic effects [5]. The truffles harvest usually varies from season to season, according to the abundance of rainfall [6]. An interesting observation is that that the chemical composition of truffles of the same species from different regions is not always the same, it is Most probably determined by a wide range of environmental factors, such as the amount and timing of rainfall, types of soil and climatic variations [7]. In spite of its nutritional advantages, little attention has been paid to the biological activities and phytochemical substances of *T. claveryi* [8].

The present study was carried out to evaluate the in vitro anti-inflammatory activity and myco-chemical constituents of the desert truffle *T. claveryi*.

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2. Materials and Methods

2.1. Extract Preparation

The desert truffles were harvested during December 2020, around Tlemcen region (El Aricha), Algeria. They were air-dried at room temperature. The dried parts were then ground into powder. The powder was extracted with a methanol/water mixture (70:30 v/v) by maceration at room temperature for 48 h. The mixture was then filtered and concentrated using a rotary evaporator [9,10].

2.2. Mycochemical Analysis

The extract obtained from *T. claveryi* was subjected to various qualitative myco-chemical tests to identify the different families of secondary metabolites present by coloration and precipitation reactions. These tests were carried out according to the techniques described by [11,12].

2.3. The Polyphenol Content

The polyphenol content of the extract was determined using the Folin-Ciocalteau method described by [13,14]. 0.1 mL extract (1 mg/mL) was mixed with 2 mL of sodium carbonate (Na₂CO₃) solution (2% w/v). After 5 min incubation, 0.1 mL of Folin-Ciocalteu reagent (1N) was added. The mixture was incubated for 30 min at room temperature in the dark. Absorbance readings were taken against a blank using a spectrophotometer at 725 nm. A calibration curve was performed under the same experimental conditions, using gallic acid as a standard. Total polyphenol levels were calculated using the linear regression equation. Results were expressed in equivalent micrograms of gallic acid per milligram of dry extract (μ g EAG/mg).

2.4. The Flavonoids Content

Flavonoids were quantified in the extract using the aluminum trichloride method described by [15]. 250 μ L of the sample was mixed with 75 μ L of a 15% (w/v) NaNO₂ solution and 1 mL of distilled water, after a 6 min rest, 75 μ L of a 10% (w/v) AlCl₃ solution was added, following a 5 min incubation, 1 mL of 4% (w/v) NaOH was introduced. The mixture was adjusted to 2.5 mL with distilled water, and the absorbance was read at 510 nm against a blank after 15 min. Total flavonoid content was calculated using catechin as standard (μ g CE/mg).

2.5. The Total Tannin

The total tannin content of the extract was estimated using the method [16]. 1.5 mL of 4% (v/w) vanillin/methanol solution and 750 µL of concentrated HCl were added to 50 µL of the extract (1 mg/mL). Measurement was taken at 550 nm after 20 min of reaction at room temperature. The condensed tannin content was determined from the linear regression equation of the catechin calibration curve. Results are expressed as microgram catechin equivalent per milligram extract (µg CE/mg).

2.6. Anti-Inflammatory Activity

Anti-inflammatory activity in vitro was assessed by the procedure of [17] with slight modifications. The basic principle of this method is the inhibition of heat (72 °C) induced BSA denaturation by extracts. 1 mL of 0.2% (w/v) BSA solution (prepared in Tris-HCl, pH: 6.3) was mixed with 1 mL of different concentrations of extract preparations (or standard). After incubation for 15 min at 37 °C, denaturation was induced by heating in a water bath for 5 min at 72 °C, followed by cooling. The control consisted of 1 mL distilled water with 1 mL 0.2% BSA, under the same operating conditions. Absorbance was measured using a UV-VIS spectrophotometer at a wavelength of 660 nm.

Denaturation inhibition percentage (%) = [(Ab control × Ab sample)/Ab control] × 100.

3. Results and Discussion

3.1. Mycochemical Screening

The mycochemical tests consist in detecting the various existing chemical families by means of qualitative characterization reactions. The results of mycochemical screening are shown in the Table 1.

Extract of T. claveryi
+
+
+
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+
+
+

Table 1. Myco-chemical screening results obtained from the hydro-methanolic extract of *T. claveryi*.

+: positive; -: negative.

The measurement of total polyphenols, flavonoids and condensed tannins revealed the presence of these compounds in the hydro-methanolic extract in varying quantities. Total phenolic components are the main bioactive constituents found in the extract, followed by condensed tannins and flavonoids. The results obtained are shown in Table 2. A large number of scientific studies demonstrate that the consumption of a polyphenol-rich diet helps reduce the risk of chronic diseases, with health benefits for the human body [18,19].

Table 2. Total phenolic content, total flavonoid content and total condensed tannins of the hydromethanolic extract of *T. claveryi*.

	Extract of T. claveryi
Total polyphenols (µg GAE/mg)	82.27 ± 1.44
Flavonoids (µg CE/mg)	14.94 ± 0.98
Condensed tannins (µg CE/mg)	27.50 ± 2.50

Values are expressed as mean ± standard deviation; GAE: gallic acid equivalent; CE: Catechin equivalent positive; –, negative.

3.2. Anti-Inflammatory Activity

Inflammation plays a crucial role in the elimination of pathogens, serving as a protective immune response. However, an uncontrolled or chronic inflammatory response can contribute to the genesis of many chronic inflammatory diseases [20].

The results of the in vitro anti-inflammatory activity of the extract are shown in the Table 3. It can be seen that the percentage inhibition of BSA denaturation (0.2%) is proportional to the different concentrations of the extract, where the strongest protection rate $83.53 \pm 1.57\%$ is registered at a concentration of 1.5 (mg/mL). However, these results are nearly proximate to those obtained for the diclofenac sodium (a non-steroidal anti-inflammatory drug) applied as a standard, which almost completely prevents BSA denaturation with a percentage of $98.43 \pm 0.52\%$ at the same concentration. This potency may be attributable to the presence of polyphenols. These components exert considerable anti-inflammatory properties, and can be regarded as contributing to their prevention [21–23].

	0.75 (mg/mL)	1.5 (mg/mL)
Extract of T. claveryi	69.41 ± 1.57	83.53 ± 1.57
Diclofenac sodium	92.16 ± 1.31	98.43 ± 0.52

Table 3. Effect of extracts on inhibition of BSA denaturation (in percentage).

4. Conclusions

The current study underlines the importance of bioactive compounds, which could have pharmacological properties and therapeutic potency, thus demonstrating the potential anti-inflammatory properties of the hydro-methanolic extract of *T. claveryi*. More research is required, to identify the main bio-compounds responsible for this remarkable potency, and their mechanism of effect.

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References

- 1. Boonsong, S.; Klaypradit, W.; Wilaipun, P. Antioxidant activities of extracts from five edible mushrooms using different extractants. *Agri. Nat. Resour.* **2016**, *50*, 89–97, https://doi.org/10.1016/j.anres.2015.07.002.
- Zitouni-Haouar, F.E.-H.; Bidartondo, M.I.; Moreno, G.; Carlavilla, J.R.; Manjón, J.L.; Neggaz, S.; Zitouni-Nourine, S.H. Bioclimatic Origin Shapes Phylogenetic Structure of Tirmania (Pezizaceae): New Species and New Record from North Africa. *J. Fungi* 2023, 9, 532. https://doi.org/10.3390/jof9050532.
- Louro, R.; Natário, B.; Santos-Silva, C. Morphological Characterization of the In Vitro Mycorrhizae Formed between Four Terfezia Species (Pezizaceae) with Cistus salviifolius and Cistus ladanifer—Towards Desert Truffles Production in Acid Soils. J. Fungi 2021, 7, 35. https://doi.org/10.3390/jof7010035.
- Beara, I.N.; Lesjak, M.M.; Cetojevic-Simin, D.D.; Marjanovic, Z.S.; Ristic, J.D.; Mrkonjic, Z.O.; Mimica-Dukic, N.M. Phenolic profile, antioxidant, anti-inflammatory and cytotoxic activities of black (Tuber aestivumVittad.) and white (Tuber magnatum Pico) truffles. *Food Chem.* 2014, 165, 460–466. https://doi.org/10.1016/j. foodchem.2014.05.116.
- Khadhri, A.; Aouadhi, C.; Aschi-Smiti, S. Screening of bioactive compounds of medicinal mushrooms collected on Tunisian territory. *Int. J. Med. Mushrooms* 2017, 19, 127–135. https://doi.org/10.1615/IntJMedMushrooms.v19.i2.40.
- 6. Al-Shabibi, M.M.A.; Toma, S.J.; Haddad, B.A. Studies on Iraqi truffles. I. Proximate analysis and characterization of lipids. *Can. Inst. Food Sci. Technol. J.* **1982**, *15*, 200–202.
- Hussain, G.; Al-Ruqaie, I.M. Occurrence, chemical composition, and nutritional value of truffles: An overview. *Pak. J. Biol. Sci.* 1999, 2, 510–514.
- 8. Harbone, J.B. *Phytochimical Methods: A Guide to Modern Techniques of Plant Analysis*, 3rd ed.; Chapman and Hall: London, UK, 1998; p. 303.
- Souna, M.; Tefiani, C.; Chaouche, T.M.; Rachid AZ, Z.I.; Salim HA, B.I.; Chaalel, A.; Souna Née Kerman, S.; Boukezzoula, N.; Lamraoui, G. The first check-list of macro-fungi in Tlemcen Wilaya (North-West of Algeria) reveals the discovery of 5 new records for Algeria. *Genet. Biodivers. J.* 2023, 7, 104–127. https://doi.org/10.46325/gabj.v7i1.319.
- 10. Souna, M. Biodiversité et Valorisation des Champignons Supérieurs de la Wilaya de Tlemcen-Étude Myco-Chimique et Activités Biologiques de Quelques Espèces. Doctoral Dissertation, University of Abou Bekr Belkaïd, Tlemcen, Algeria, 2023.
- 11. Dahham, S.S.; Tabana, Y.M.; Iqbal, M.A.; Ahamed, M.B.; Ezzat, M.O.; Majid, A.S.; Majid, A.M. The anticancer, antioxidantand antimicrobial properties of the sesquiterpene b –caryophyllene from the essential oil of Aquilaria crassna. *Molecules* **2015**, *20*, 11808–11829.
- 12. Trease, G.E.; Evans, W.C. Pharmacognosy. In *Phytochemicals*, 15th ed.; WB Saunders: Edinburgh, NY, USA, 2002; pp. 246–249, 391–393.

- 13. Singleton, V.L.; Orthofer, R.; Lamuela-Raventós, R.M. Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods Enzymol.* **1999**, 299, 152–178. https://doi.org/10.1016/j.scienta.2016.11.004.
- 14. Singleton, V.L.; Rossi, J.A. Colorimetry of total phenolic with phosphomolybdic-phosphotungstic acid reagents. *Am. J. Enol. Vitic.* **1965**, *16*, 144–158.
- 15. Dewanto, V.; Xianzhong, W.; Adom, K.K.; Liu, R.H. Thermal processing enhances the nutritional value of tomatoes by increasing total antioxidant activity. *J. Agric. Food Chem.* **2002**, *50*, 3010–3014. https://doi.org/10.1021/jf0115589.
- Sun, B.; Ricardo-da-Silva, J.M.; Spranger, I. Critical Factors of Vanillin Assay for Catechins and Proanthocyanidins. J. Agric. Food Chem. 1998, 46, 4267–4274. https://doi.org/10.1021/jf980366j.
- 17. Sakat, S.S.; Juvekar, A.R.; Gambhire, M.N. International Journal of Pharmacy and Pharmaceutical Sciences. *Int. J. Pharm. Pharm. Sci.* **2010**, *2*, 146–155.
- 18. Ziółkiewicz, A.; Kasprzak-Drozd, K.; Rusinek, R.; Markut-Miotła, E.; Oniszczuk, A. The Influence of Polyphenols on Atherosclerosis Development. *Int. J. Mol. Sci.* 2023, 24, 7146. https://doi.org/10.3390/ijms24087146.
- Hemmami, H.; Seghir, B.B.; Zeghoud, S.; Ben Amor, I.; Kouadri, I.; Rebiai, A.; Zaater, A.; Messaoudi, M.; Benchikha, N.; Sawicka, B.; et al. Desert Endemic Plants in Algeria: A Review on Traditional Uses, Phytochemistry, Polyphenolic Compounds and Pharmacological Activities. *Molecules* 2023, 28, 1834. https://doi.org/10.3390/molecules28041834.
- Germolec, D.R.; Shipkowski, K.A.; Frawley, R.P.; Evans, E. Markers of Inflammation. In *Immunotoxicity Testing: Methods and Protocols*; DeWitt, J.C., Rockwell, C.E., Bowman, C.C., Eds.; Springer: Berlin/Heidelberg, Germany, 2018; pp. 57–79. https://doi.org/10.1007/978-1-4939-8549-4_5.
- Tassinari, V.; Smeriglio, A.; Stillittano, V.; Trombetta, D.; Zilli, R.; Tassinari, R.; Maranghi, F.; Frank, G.; Marcoccia, D.; Di Renzo, L. Endometriosis Treatment: Role of Natural Polyphenols as Anti-Inflammatory Agents. *Nutrients* 2023, 15, 2967. https://doi.org/10.3390/nu15132967.
- 22. Suzuki, T.; Ohishi, T.; Tanabe, H.; Miyoshi, N.; Nakamura, Y. Anti-Inflammatory Effects of Dietary Polyphenols through Inhibitory Activity against Metalloproteinases. *Molecules* **2023**, *28*, 5426. https://doi.org/10.3390/molecules28145426.
- Martínez-Coria, H.; Arrieta-Cruz, I.; Gutiérrez-Juárez, R.; López-Valdés, H.E. Anti-Inflammatory Effects of Flavonoids in Common Neurological Disorders Associated with Aging. Int.J. Mol. Sci. 2023, 24, 4297. https://doi.org/10.3390/ijms24054297.

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