



2

3

4

5

6 7

8

9

10

11

12

13 14

25

26

Proceedings The *ex vivo* and *in vitro* antithrombotic properties of fermented Irish ovine yogurt drink

⁺Sakshi Hans ¹, Harishkumar Rajendran ¹, Katie Shiels ³, Sushanta Kumar Saha ³, Alexandros Tsoupras ³, Ronan Lordan ^{4,*} and Ioannis Zabetakis ^{1,*}

- ¹ Department of Biological Sciences, University of Limerick, (sakshi.hans@ul.ie), (harishkumar.rajendran@ucd.ie), (ioannis.zabetakis@ul.ie)
- ² Shannon Applied Biotechnology Centre, Technological University of the Shannon, Moylish Park, Limerick, Ireland, (katie.shiels@tus.ie), (sushanta.saha@tus.ie).
- ³ Department of Chemistry, School of Sciences, International Hellenic University, Saint Luke, GR65404, Kavala, Greece, (atsoupras@gmail.com).
- ⁴ Institute for Translational Medicine and Therapeutics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, 19104, USA, **(ronan.lordan@pennmedicine.upenn.edu)**.
- * Correspondence: ronan.lordan@pennmedicine.upenn.edu; ioannis.zabetakis@ul.ie

Abstract: Platelet function is closely linked with cardiovascular health. Functional foods such as 15 yogurt and oily fish are enriched in bioactive polar lipids which can reduce platelet activation and 16 incidence of cardiovascular disease. This project thus investigated the effect of fermented ovine yo-17 gurt on platelet sensitivity in human plasma. Overall, in vitro studies established inhibitory effect 18 on platelet aggregation by yogurt lipid fractions. Results from dietary intervention showed that yo-19 gurt intake reduces platelet activation against thrombin pathway, compared to placebo. Larger scale 20 studies are required to robustly establish the effect of yogurt intake on platelet sensitivity, following 21 this interim analysis. 22

Keywords:yogurt;polarlipids;plateletaggregation;thrombin-activatorreceptorpep-tide-623(TRAP-6);nutraceuticals;antithrombotic;functional foods24

1. Introduction

While a measured immune response is a necessary protective physiological reaction 27 in response to tissue damage or pathogenic insults, unresolved inflammation is implicated 28 in the pathology of many chronic diseases (Neher et al., 2011). The initiation of cardiovas-29 cular diseases (CVD) and other conditions have been linked to low-grade systemic inflam-30 mation (Furman et al., 2019, Zotova et al., 2010). Platelets are important effectors of these 31 immune responses by mechanisms such as recruitment of neutrophils to the site of in-32 flammation, and by the release of proinflammatory factors and chemokines that further 33 activate leukocytes and intensify the immune response (Stokes and Granger, 2012, Gros 34 et al., 2014). Platelet activity can be modulated by stimuli such as platelet agonists which 35 contribute to platelet aggregation, or platelet inhibitors, which deactivate platelets and 36 discourage formation of thromboses (Willoughby, 2002). Thrombin receptor activator 37 peptide-6 (TRAP-6) is a peptide fragment that selectively activates the thrombin receptor 38 protease-activated receptor 1 (PAR-1), thus inducing platelet activation similar to throm-39 bin. Thrombin is a serine protease that is a key enzyme in the coagulatory pathway. It 40signals by binding to the protease-activated receptors (PARs) which are a family of trans-41 membrane peptide receptors (Coughlin, 2000, Reiter et al., 2003). Platelet-activating factor 42 (PAF) is a phospholipid mediator of inflammation that also induces platelet activation 43 and aggregation. PAF functions by binding to the PAF receptor (PAF-R) a G-protein 44

Citation: To be added by editorial staff during production.

Academic Editor: Firstname Lastname

Published: date



Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/).

2

coupled receptor (GPCR) located on cells involved in immune function (Yost et al., 2010, Harishkumar et al., 2022).

Consumption of foods or nutraceuticals rich in bioactive lipids (dietary polar lipids) 3 that target platelet signalling pathways such as thrombin and PAF may be beneficial 4 against inflammatory diseases (Harishkumar et al., 2022). However, research is still at a 5 preliminary stage to date, which necessitates the need for further study. Numerous stud-6 ies have been conducted to investigate the cardioprotective properties of polar lipids such 7 as marine and dairy-derived lipids in vitro (Tsoupras et al., 2018, Lordan et al., 2019b, 8 Glenn-Davi et al., 2022, Lordan et al., 2019a). These dietary polar lipids appear to inhibit 9 platelet aggregation induced by agonists such as PAF and TRAP-6. 10

Ovine or sheep's milk has been noted for its many potential health benefits besides 11 its rich nutritional content, such as its anti-inflammatory, antiplatelet, and antimicrobial 12 properties (Balthazar et al., 2017, Mohapatra et al., 2019). Fermentation of ovine milk has 13 been shown to further enhance its antithrombotic properties by altering the fatty acid pro-14 file of monounsaturated fatty acids (MUFA) (Lordan et al., 2019b, Glenn-Davi et al., 2022). 15

The aim of this study was to assess the antiplatelet properties of ovine yogurt drink 16 (YD) enriched in polar lipids using platelet aggregometry *in vitro* and to assess the effect 17 of its long-term consumption on platelet sensitivity in a randomized controlled trial. 18

2. Materials and Methods

2.1. Materials and Instrumentation

All consumables and solvents were purchased from Fisher Scientific Ltd. 22G safety 21 needles and evacuated S-monovettes were purchased from Sarstedt Ltd. Platelet ag-22 gregometry consumables were purchased from Labmedics LLP. Other reagents and 23 chemicals were purchased from Sigma Aldrich. Platelet aggregation testing was per-24 formed on a Chronolog-490 two channel turbidimetric platelet aggregometer and its ac-25 companying AGGRO/LINK software. 26

2.2. Production of the yogurt drink and placebo yogurt drink

Ewe's whole milk was obtained from Velvet Cloud Ltd. (Claremorris, Co Mayo, Ire-28 land), from Friesland and Lacaune breed of dairy ewes. The percentage of fat in raw milk 29 was approximately 7%. The milk was pasteurised on site by heating to 91 °C for 15 s, and 30 then cooled to 42 °C before being packaged and refrigerated (4 °C ± 1 °C). F.

After pasteurization, the milk was divided into two smaller batches and refrigerated. 32 From one of these batches, skimmed milk was produced by double centrifugation; initially 33 by centrifugation at 6900 x g for 30 minutes at 40°C, and afterwards at 7440 x g for 30 min 34 also at 40°C. After both centrifugations, the settled layer of fat was removed to produce 35 skimmed milk. Both milk batches were then inoculated with commercially available YO-36 MIX 205 LYO 250 DCU starter culture (Danisco France SAS, France; 5 mg/100 mL whole 37 milk) to induce fermentation at a temperature of 40.5 °C for a period of 7h. The milk 38 batches were placed in a water bath to ferment and reach a pH = 4.6 and desired con-39 sistency (thick and creamy). The cultures contained the following probiotics- Streptococcus 40 thermophilus, Lactococcus delbrueckii subsp. Bulgaricus, Lactobacillus acidophilus, and 41 Bifidobacterium lactis. After fermentation the milk was then poured into 250 ml bottles and 42 sealed. 43

2.3. Extraction of lipids and determination of nutrient content

Total lipids (TL) of yogurt drink (YD) and placebo drink (PD) were extracted from 45 100 g of sample according to the Bligh and Dyer technique [21]. Total neutral lipids (TNL) 46 and total polar lipids (TPL) were further isolated from TL by counter-current distribution 47 method [22]. Samples were stored under nitrogen at -20 °C. 48

19

20

27

2.4. In vitro assessment of antithrombotic properties of lipid fractions against TRAP-6 and PAFinduced platelet aggregation

Platelet inhibition was assessed for each lipid fraction using platelet aggregometry 3 as detailed in Table 2. Various subclasses of TPL were isolated using preparative thin-4 layer chromatography (TLC) and the IC50 of these was determined similarly (Table 3). 5 The half-maximal inhibitory concentration (IC50) was expressed as the mean mass of lipid 6 fraction (μ g) in the aggregometer cuvette ± standard deviation (SD). All experiments were 7 performed in triplicate. 8

2.5. Dietary intervention study of Irish ovine yogurt drink

The dietary intervention was designed as a randomized double-blind crossover 10 study with the control group provided a placebo drink containing significantly lower 11 amounts of polar lipids (n = 18) produced as described in section 2.2. Two study phases 12 were organized with equally divided groups and each phase of trial was conducted for 28 13 days. Figure 1 is a schematic representation of the study design. 14

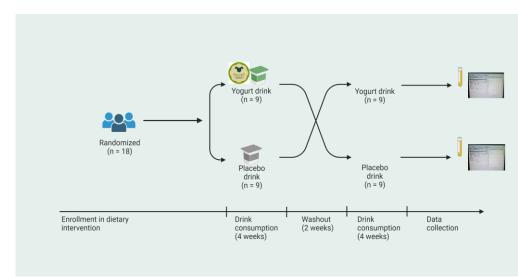


Figure 1. Schematic representation of the design of the crossover dietary intervention study. 18 par-16 ticipants were recruited in the randomized study with equal numbers assigned 250 gm of either the 17 yogurt drink or placebo drink. Blood sampling was performed at four timepoints with a washout period of two weeks prior to the crossover phase. 19

3. Results

3.1. Yield of lipid fractions isolated from ovine yogurt drink

A side-by-side comparison of the ovine yogurt drink and placebo drink with respect to their nutritional content is given in Table 1. The yogurt drink was significantly higher in fat content compared to the placebo (prepared from skimmed milk).

Table 1. Yield of the lipid fractions (TL, total lipid; PL, polar lipids; NL, neutral lipids) from raw sheep's milk, skimmed milkderived placebo drink and full fat yogurt drink (g/100mL). Data is presented as mean ± SD.

Total lipids (TL) in grams	Polar lipids (PL) in gm	Neutral lipids (NL) in gm
9.96 ± 0.30 a	0.16 ± 0.01 a	8.24 ± 0.23 a
8.17 ± 0.22 ^a	0.15 ± 0.004 a	7.75 ± 0.19 a
0.36 ± 0.01 b	0.06 ± 0.002 b	0.29 ± 0.01 b

a,b, Different superscripts in the same column indicate significant differences among different lipid 27 extracts within the same lipid class (p < 0.05) when means are compared using a Tukey's HSD 28

1

2

9

18

20

21

22

23

24

25

26

2

3

4

5

6

12

13

19

23

multiple comparison test. Polar lipids (PL) are amphiphilic and possess a hydrophilic head and hydrophobic tail. Neutral lipids (NL) have a hydrophobic structure and lack charge.

3.2. In vitro assessment of antithrombotic properties of lipid fractions against TRAP-6 and PAFinduced platelet aggregation

Lipid fractions (TL, TNL, and TPL) were extracted and assessed for bioactivity against PAF and TRAP-6 (**Table 2**.).

Table 2. Inhibitory effects (IC50 values) of polar lipids (PL), neutral lipids (NL), and total lipids (TL) isolated from the ovine yogurt	7
drink against human platelet aggregation induced by PAF and TRAP-6 in vitro. Data is expressed as mean ± SD.	8

		IC50 against TRAP-6 (µg)	IC50 against PAF (µg)
Yogurt drink	YPL	312.95 ± 15.04	156.72 ± 5.88
	YNL	259.41 ± 13.56	729.15 ± 35.079
	YTL	209.57 ± 10.11	248.23 ± 11.39
Placebo (skimmed milk)	YPL	45.8 ± 2.05	44.0 ± 1.98
	YNL	47.37 ± 2.14	47.20 ± 2.13
	YTL	59.16 ± 3.13	53.84 ± 2.85
	1	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	11/2

a,b,c,d,eDifferent superscripts in the same column indicate significant differences among different lipid extracts within the same lipid class (p < 0.05) when means are compared using a Tukey's HSD10multiple comparison test.11

3.3. In vitro assessment of antithrombotic properties of each PL subclass towards TRAP-6 and PAF-induced platelet aggregation in PRP, expressed as mean \pm SD.

Table 3 depicts the in vitro inhibition of the subclasses of yogurt TPL in response to14PAF and TRAP-6-induced platelet aggregation, represented as IC50 values for each TLC15fraction. The lipid fractions corresponding to PC (phosphatidylcholine) had the lowest16IC50 values against PAF and TRAP and consequently the highest biological activity, closely17followed by the PE fraction against PAF.18

Table 3. Antiplatelet activity of each PL subclass towards PAF-induced platelet aggregation in PRP, expressed as mean ± SD.

Yogurt drink polar lipid fractions		EC50 (-) or IC50 against PAF (μg)	IC50 against TRAP-6 (μg)
1	LPC	-48.1 ± 3.9	348.6 ± 7.9
2	SM	1120 ± 54	250.6 ± 8.3
3	PC	169.5 ± 8.3	205.3 ± 8.9
4	CL	ND	208.0 ± 10.8
5	PE	196.7 ± 6.4	592.1 ± 8.8
	A11		

Abbreviations: CL, cardiolipin; LPC, lyso-phosphatidylcholine; ND, not determined; PAF, platelet-20activating factor; PC, phosphatidylcholine; PE, phosphatidylethanolamine; SM, sphingomyelin;21TRAP-6, thrombin receptor activator peptide 6.22

3.4. Assessment of the impact of dietary yogurt intake on platelet sensitivity against TRAP-6

Table 4 depicts the effect of yogurt consumption on TRAP-6 induced platelet activity,24compared to placebo, at different timepoints during the study. Overall, a trend of in-25creased EC50 in the YD cohort was observed indicating reduced platelet activation status,26although not statistically significant.27

Table 4. The effect of yogurt (YD) consumption on platelet activation by TRAP-6, expressed as the28mean Δ EC50 at different timepoints.29

Treatment group	ΔEC50 Day 0-28	ΔEC50 Day 42-70	Combined ΔEC50
YD	17.22 ± 32.2	10.22 ± 33.35	13.72 ± 31.1
PD	7.45 ± 27.5	-5.97 ± 19.95	0.742 ± 23.6

4. Discussion

Research has shown that fermented ovine dairy has potent antiplatelet effects against 2 the thrombin pathway of platelet activation, in vitro in human PRP. [37]. The effect of 3 daily intake of yogurt on the PAF pathway of platelet activation has also been demon-4 strated as a dietary intervention study model, although no intervention studies so far have 5 demonstrated the ex vivo effect of yogurt on platelet activation through the thrombin 6 pathway [33]. Due to funding constraints the present study did not investigate the effect 7 of non-fermented ovine milk on TRAP-6 platelet activation pathway. However, in vitro 8 data suggests there is less potent activity of non-fermented milk vs fermented yogurt. 9 Thus, the present dietary intervention study sought to determine the effect of consump-10 tion over 28 days of an ovine yogurt drink (YD) enriched with polar lipids on baseline 11 platelet reactivity, through the thrombin pathway of activation. In vitro analyses found 12 that yogurt lipids inhibit platelet activation through both PAF and thrombin pathways. 13 While we observed a clear trend of yogurt intake reducing platelet activation through 14 thrombin pathway in human volunteers, studies on a larger scale are required to establish 15 impact of yogurt lipids more robustly on platelet activity. 16

Author Contributions: Conceptualization, I.Z. and R.L.; methodology, I.Z., S.H., R.L., and R.H; for-17mal analysis, S.H., R.H., and R.L.; investigation, S.H., R.H., K.S., and S.K.S.; resources, I.Z.; data18curation, R.H., S.H.; writing—original draft preparation, S.H. and R.L.; writing—review and edit-19ing, R.L., S.H. and I.Z; visualization, S.H., and R.H.; supervision, I.Z.; project administration, I.Z.,20R.H., R.L., and S.H.; funding acquisition, I.Z. and R.L.21

Funding: This research was funded by Enterprise Ireland, grant number IP-2021-0972.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki and approved by the Institutional Review Board (or Ethics Committee) of University of Limerick (ethical approval code 2022_01_01_S&E, approved 16.02.2022).

Informed Consent Statement: Informed consent was obtained from all subjects involved.

Acknowledgments: The authors are grateful to the volunteers who took part in the study and to27Ms. Elaine Ahern for her phlebotomy support. The authors acknowledge the financial support of28Enterprise Ireland (study grant reference: IP-2021-0972), the Velvet Cloud private company for their29donation and help preparing the yogurt drinks and placebo drinks. We also thank the Department30of Physical Education and Sports Science and the Department of Biological Sciences at the University of Limerick, Ireland, for their continued support and use of their facilities.32

Conflicts of Interest: The funders had no role in the design of the study; in the collection, analyses,33or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.34The authors declare no conflicts of interest.35

References

- BALTHAZAR, C. F., PIMENTEL, T. C., FERRAO, L. L., ALMADA, C. N., SANTILLO, A., ALBENZIO, M., MOLLAKHALILI, 37 N., MORTAZAVIAN, A. M., NASCIMENTO, J. S., SILVA, M. C., FREITAS, M. Q., SANT'ANA, A. S., GRANATO, D. & 38 CRUZ, A. G. 2017. Sheep Milk: Physicochemical Characteristics and Relevance for Functional Food Development. *Compr* 39 *Rev Food Sci Food Saf*, 16, 247-262.
- 2. COUGHLIN, S. R. 2000. Thrombin signalling and protease-activated receptors. Nature, 407, 258-64.
- FURMAN, D., CAMPISI, J., VERDIN, E., CARRERA-BASTOS, P., TARG, S., FRANCESCHI, C., FERRUCCI, L., GILROY, D.
 W., FASANO, A., MILLER, G. W., MILLER, A. H., MANTOVANI, A., WEYAND, C. M., BARZILAI, N., GORONZY, J. J.,
 RANDO, T. A., EFFROS, R. B., LUCIA, A., KLEINSTREUER, N. & SLAVICH, G. M. 2019. Chronic inflammation in the
 etiology of disease across the life span. *Nat Med*, 25, 1822-1832.
- GLENN-DAVI, K., HURLEY, A., BRENNAN, E., COUGHLAN, J., SHIELS, K., MORAN, D., SAHA, S. K., ZABETAKIS, I. 46 & TSOUPRAS, A. 2022. Fermentation Enhances the Anti-Inflammatory and Anti-Platelet Properties of Both Bovine Dairy 47 and Plant-Derived Dairy Alternatives. *Fermentation*, 8.

1

32 33

36

41

22

23

24

25

5.	GROS, A., OLLIVIER, V. & HO-TIN-NOE, B. 2014. Platelets in inflammation: regulation of leukocyte activities and vascular	1
	repair. Front Immunol, 5, 678.	2
6.	HARISHKUMAR, R., HANS, S., STANTON, J. E., GRABRUCKER, A. M., LORDAN, R. & ZABETAKIS, I. 2022. Targeting	3
	the Platelet-Activating Factor Receptor (PAF-R): Antithrombotic and Anti-Atherosclerotic Nutrients. Nutrients, 14.	4
7.	JAIN, A. P., AGGARWAL, K. K. & ZHANG, P. Y. 2015. Omega-3 fatty acids and cardiovascular disease. Eur Rev Med	5
	Pharmacol Sci, 19, 441-5.	6
8.	LORDAN, R., O'KEEFFE, E., DOWLING, D., MULLALLY, M., HEFFERNAN, H., TSOUPRAS, A. & ZABETAKIS, I. 2019a.	7
	The in vitro antithrombotic properties of ale, lager, and stout beers. Food Bioscience, 28, 83-88.	8
9.	LORDAN, R., TSOUPRAS, A. & ZABETAKIS, I. 2017. Phospholipids of Animal and Marine Origin: Structure, Function, and	9
	Anti-Inflammatory Properties. Molecules, 22.	10
10.	LORDAN, R., WALSH, A. M., CRISPIE, F., FINNEGAN, L., COTTER, P. D. & ZABETAKIS, I. 2019b. The effect of ovine	11
	milk fermentation on the antithrombotic properties of polar lipids. Journal of Functional Foods, 54, 289-300.	12
11.	MOHAPATRA, A., SHINDE, A. K. & SINGH, R. 2019. Sheep milk: A pertinent functional food. Small Ruminant Research,	13
	181, 6-11.	14
12.	NASOPOULOU, C., GOGAKI, V., PANAGOPOULOU, E., DEMOPOULOS, C. & ZABETAKIS, I. 2013. Hen egg yolk lipid	15
	fractions with antiatherogenic properties. Anim Sci J, 84, 264-71.	16
13.	NEHER, M. D., WECKBACH, S., FLIERL, M. A., HUBER-LANG, M. S. & STAHEL, P. F. 2011. Molecular mechanisms of	17
	inflammation and tissue injury after major traumais complement the "bad guy"? J Biomed Sci, 18, 90.	18
14.	REITER, R., DERHASCHNIG, U., SPIEL, A., KEEN, P., CARDONA, F., MAYR, F. & JILMA, B. 2003. Regulation of protease-	19
	activated receptor 1 (PAR1) on platelets and responsiveness to thrombin receptor activating peptide (TRAP) during systemic	20
	inflammation in humans. <i>Thromb Haemost</i> , 90, 898-903.	21
15.	STOKES, K. Y. & GRANGER, D. N. 2012. Platelets: a critical link between inflammation and microvascular dysfunction. J	22
	Physiol, 590, 1023-34.	23
16.	TSOUPRAS, A., LORDAN, R., DEMURU, M., SHIELS, K., SAHA, S. K., NASOPOULOU, C. & ZABETAKIS, I. 2018.	24
	Structural Elucidation of Irish Organic Farmed Salmon (Salmo salar) Polar Lipids with Antithrombotic Activities. Mar Drugs,	25
	16.	26
17.	WILLOUGHBY, S. 2002. Platelets and cardiovascular disease. European Journal of Cardiovascular Nursing, 1, 273-288.	27
18.	XANTHOPOULOU, M. N., KALATHARA, K., MELACHROINOU, S., ARAMPATZI-MENENAKOU, K.,	28
	ANTONOPOULOU, S., YANNAKOULIA, M. & FRAGOPOULOU, E. 2017. Wine consumption reduced postprandial	29
	platelet sensitivity against platelet activating factor in healthy men. Eur J Nutr, 56, 1485-1492.	30
19.	YOST, C. C., WEYRICH, A. S. & ZIMMERMAN, G. A. 2010. The platelet activating factor (PAF) signaling cascade in	31
	systemic inflammatory responses. Biochimie, 92, 692-7.	32
20.	ZOTOVA, E., NICOLL, J. A., KALARIA, R., HOLMES, C. & BOCHE, D. 2010. Inflammation in Alzheimer's disease:	33
	relevance to pathogenesis and therapy. Alzheimers Res Ther, 2, 1.	34