



Short Proceeding paper

DEVELOPMENT OF POLYPHENOLICS EXTRACTS FROM MEXICAN CROPS AS NATURAL ANTIMICROBIAL AGENTS FOR POSTHARVEST TREATMENTS

Laura M. Aguilar-Veloz*, J. Arturo Olguín-Rojas, Diana Gómez-Flores, Cecilia Vázquez-González, Alfredo Salvador Castro-Díaz, Manuel González-Pérez, Montserrat Calderón-Santoyo and Juan Arturo Ragazzo-Sánchez

- ¹ Ingeniería en Procesos Bioalimentarios, Universidad Tecnológica de Tecamachalco. Avenida, Universidad Tecnológica 1, 75483 Tecamachalco, Puebla, México
- ² Laboratorio Integral de Investigación en Alimentos, Tecnológico Nacional de México/Instituto Tecnológico de Tepic, Av. Tecnológico #2595, Colonia Lagos del Country, Tepic C.P. 63175, Nayarit, México
- * Correspondence: lauraguilarveloz@gmail.com

Abstract: Last decade, the use of natural antimicrobial agents, like polyphenolic extracts in postharvest applications has gained attention. However, a significant challenge lies ahead. Demonstrating commercial-scale production feasibility, practical applications, and economic viability compared to traditional agrochemicals is crucial. This review focuses on the achievements and obstacles in using polyphenolic extracts from Mexican crops as natural antimicrobial agents against postharvest phytopathogens and foodborne microorganisms. Comprehensive knowledge of the molecular mechanisms in vitro and in vivo, as well as plant systems, is essential for better results. Toxicity assessment and impact on fruit quality need evaluation. Incorporating coating and encapsulation techniques can enhance extract effectiveness. An integrated approach is needed for efficient, cost-effective control throughout the preharvest, harvest, and post-harvest phases. Successful commercialization depends on cost-benefit analysis, infrastructure, raw materials, local needs, and technical facilities, among other factors. Additionally, incorporating strategies related to circular economy can improve plant residue utilization and enhance technological and marketing approaches.

Keywords: polyphenolic extracts; natural antimicrobials; postharvest; Mexican crops

1. Introduction

Mexico is a megadiverse country with a high potential of natural resources, occupying the fourth place in the world for its richness in plant species. However, high food losses, the intensive use of agrochemicals, agro-food waste dimension and the lack of infrastructure for their use and management, affect its agro-industrial development [1]. According to the circular economy principles, the valorization of agro-food waste has showed growing attention due to the possibilities to be a source of high biological active compounds (HBAC), with applications in food, pharmaceutical and cosmetic industries, among others. Furthermore, HBAC are environmental and human health friendly alternatives for pathogens control in different contexts. Particularly, the polyphenols (PPhs) are considered very attractive compounds because are responsible of the antioxidant and antimicrobial properties of some vegetable extracts [2-4]. Postharvest process is focused on food properties (nutritional, taste, aroma, and good appearance) and food safety. Them, the studies will be focused to phytopathogenic microorganisms and those foodborne bacteria, which threaten the health of consumers [5]. The present review aims to evidence the success and challenges of research on PPhs extracts from Mexican crops as NA, against phytopathogens and foodborne microorganisms on postharvest processes.

Citation: To be added by editorial staff during production.

Academic Editor: Firstname Lastname

Published: date



Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licens es/by/4.0/).

2. Mexican crops as potential sources of PPhs extracts

Among the main crops in Mexico, there are some traditional cultivars that serve as sources of PPhs extracts, which are known for their antimicrobial properties. The selection of the following crops (Table 1) was based on their native origin in Mexico (Habanero chili pepper, allspice, prickly pear), commercial importance (cinnamon), their impact on agricultural development in various Mexican states (broccoli and jackfruit), and the attention they have received from the Mexican and international scientific communities. In addition, a brief description of each crop is provided below:

Source	Extract / PPhs concentration	Pathogen	Concentration	Inhibition	Reference
		Foodborne microorganisms			
Prickly pear (Opuntia ficus indica (L.) Mill.) (Cladode)	Aqueous PPhs extract 700 mg GAE/100g	Salmonella typhi Helicobacter pylori Escherichia coli Staphylococcus aureus	3.40 mg/mL 1.37 mg/mL 1.41 mg/mL, 1.41 mg/mL	> 50%	[19].
Piper dioica L.merril	Aqueous eugenol extract*	Streptococccus mutans	0.1-1mg/mL	15.9 mm a 24 h 18.6 mm a 48 h	[24].
Clove (Eugenia caryophyl- lata Thunb.) Mexican oregano (Lippia berlandieri Schauer)	Alcoholic extracts of euge- nol and carvacrol*	Listeria monocytogenes ATCC 19114 Echerichia coli ATCC 25922		> than non encapsu- lated extracts	[25]
,	- Habanero Pepper Peel Etha- nolic Extract	C. albicans—ATCC 90028 C. tropicalis—CI C. glabrata—ATCC2001 C. krusei—ATCC 6258	3000 μg/mL 750 μg/mL 3000 μg/mL 3000 μg/mL	100%	[26]
		Phytopatogens			
Jackfruit leaf (Artocarpus heterophyllus Lam)	Hydroalcoholic PPhs extract	Alternaria alternate	1 mg/mL**	40 %	[4]
Jack ruit leaf (Artocarpus heterophyllus Lam)	Hydroalcoholic PPhs extract	Colletotrichum gloesporioides	1-5 mg/mL**	40-60 %	[5]
Pepper (Piper Dioica L. mer- ril)	Nanoencapsulated eugenol*	Colletotrichum gloesporioides	0.5 % (p/p) de nanoformulación con eugenol incorporado	100%	[27]
Cinnamon (Cinnamomum zeylanicum J.Presl)	Methanolic extract	Fusarium spp.	300 ppm	31.8-45.6 %	[28]
Xoconostle (Opuntia oli- gacantha C.F. Först)	Nanoencapsulated emulsion 409.37 mg GAE/Ml	Colletotrichum gloesporioides		> that non encapsu- lated extracts	[29]

Table 1. Antimicrobial effect of phenolic extracts of different origin on pathogenic microorganisms (in vitro).

*pure compound. **concentration of PPhs.

2.1. Alslpice (Piper Dioica L. Merril)

Allpice is native from Mexico and Central America and has been domesticated and 2 naturalized in various tropical countries. In Mexico, it is ecologically distributed on the 3 slope of the Gulf of Mexico, from the north of Puebla and Veracruz to the south of the 4 Yucatan, relating some similar climatic and edaphic characteristics [16]. Its national pro-5 duction is channeled mainly to the international market. Allspice production in the coun-6 try is located under two contexts: the traditional backyard agriculture, family and manual 7 labor, as an alternative crop, with a lack of support and technology; and another most 8 sophisticated international trade system [17]. According the authors, the fruit contains 9 $1.89\% \pm 0.94$ of essential oil. The eugenol content in the leaves and fruit, varies from 38.23 10 to 46.52%, with respect to its origin and location. 11

2.2. Broccoli (Brassica oleracea var. Italica)

Broccoli is an annual crop, which falls within the cruciferous family and the genus 13 Brassica oleracea. In Mexico, Guanajuato accounts for over 67% of the national broccoli pro-14 duction, alongside other contributing states [6] Within broccoli, the presence of HBAC has 15 been empirically substantiated to exert antimicrobial properties. These compounds en-16 compass vitamin C, carotenoids, PPhs, glucosinolates, sulforaphane, and enzymes, such 17 as peroxidases and lyases, among others. Diverse extraction techniques facilitate the iso-18 lation of these NA agents. For instance, ethyl acetate and chloroform ethyl acetate extracts 19 have exhibited discernible activity against Escherichia coli and Candida albicans, respec-20 tively. Furthermore, aqueous extracts derived from lyophilized broccoli have demon-21 strated efficacy against Bacillus cereus, Streptococcus faecalis, Staphylococcus aureus, Listeria 22 monocytogenes, Salmonella typhimurium, Pseudomonas aeruginosa, E. coli, Shigella sonnei, and 23 C. albicans [7]. Notably, the presence of NA is not confined solely to the broccoli florets but 24 extends to agro-industrial byproducts, such as stems and leaves. Specifically, aqueous ex-25 tracts derived from broccoli stems, at a concentration ratio of 1:20 w/v, have demonstrated 26 antimicrobial activity against L. monocytogenes, with a minimum inhibitory concentration 27 (MIC) of 102.4 mg/mL [8]. 28

2.3. Cinnamon (Cinnamomum Zeylanicum J.Presl)

The Cinnamomum genus includes C. zeylanicum and C. cassia, they are widely used 30 spice worldwide and belongs to the Lauraceae family. In Mexico, cinnamon bark serves 31 various purposes, with its prominent role as an aromatic ingredient in foods. Its applica-32 tion in ethnobotanical medicine due to its antimicrobial properties, as well as its antioxi-33 dant and antimutagenic capabilities [9]. C. cassia contains a higher cumarin concentration, 34 up to 1% [10]. This characteristic is linked to the fact that isocoumarin, a secondary me-35 tabolite found in cinnamon, is hepatotoxic to humans. The European Food Safety Author-36 ity (EFSA) regulates the tolerable daily intake (TDI) of coumarin, setting limits between 37 0.1 mg to 2.0 mg/kg of body weight for general food products. 38

2.4. Habanero chili pepper (Capsicum chinense Jacq.)

Habanero chili pepper represents an integral part of Mexican cultural tradition and 40identity, having left its mark on Mexican cuisine for at least eight centuries [11]. Besides, 41 it holds substantial economic importance, in 2019, global production reached approxi-42 mately 42 million tons [1]. The increase in demand for habanero peppers and their prod-43 ucts has led to a growth in waste generation (peduncles, stems, and leaves) [12]. Different 44 researchers explored the HBAC profile of habanero chili pepper, Mokhtar *et al.* [13] eval-45 uated the *in vitro* antimicrobial activity of pepper PPhs and capsaicinoids against thirteen 46 pathogenic bacteria. The principal PPhs present are caffeic acid, quercetin, rutin, 47 kaempferol, coumarin, and narangin. Vuerich et al. [14] analyzed the antifungal activity 48of the ethyl acetate extract against some of the major fungal and *Oomycetes pathogens* of 49

1

12

29

grapevine, the total concentration of PPhs in the oleoresin accounted for 268.5±15.4 mg g⁻¹ (dw), with vanillic acid (65%) being the predominant compound, followed by protocatechuic acid (13%), both of which are hydroxycinnamic derivatives. Habanero chili and its by-products present an interesting alternative as potential sources of PPhs extracts as natural antimicrobial agents for postharvest treatments.

2.5. Jackfruit (Artocarpus heterophyllus Lam.)

Jackfruit is exhibiting applications in traditional medicine, agriculture and industry 7 in some geographical areas, and in Mexico its major economic impact has been in Nayarit 8 state [15]. It is one of the most common evergreen trees found in the tropical regions, 9 which must periodically be pruned to facilitate fruit harvest. A considerable volume of 10 this biomass (about 10,500 tons/ha of leaf per year), its high phytochemical and protein 11 content. Regarding the PPhs extracts recovery, some advantages of the emerging solid-12 liquid techniques such as high hydrostatic pressure, ultrasound-assisted extraction, and 13 microwave-assisted extraction have been demonstrated by Mexican authors. Extracts con-14 taining mainly flavonoids, tannins, glycosides, phenolic and organic acids, with antimi-15 crobial action against different foodborne and phytopathogenic microorganisms were ten-16 tatively identified [4,5]. 17

2.6. Prickly pear (Opuntia ficus indica (L.) Mill.)

Opuntia is a diverse family that predominates in the arid and semi-arid regions of 19 the Americas. About 1400–1800 cacti's species have been described in the world, and Mex-20 ico is the country with the greatest diversity (850 species, 84% endemic) [18]. PPhs in dif-21 ferent parts (fruit, cladode, and pulp) of prickly pear are known to contribute to its antioxidant and antimicrobial activities. The studies indicated that cladode possessed a higher 23 quantity of phenolics compared with that observed in fruit and pulp. Gallic acid was the most abundant phenolic compound (66.19 μ g/g) [19]. 25

3. Antimicrobial properties vs. antioxidant effect of PPhs

The antioxidant capacity of these crops, depend on the variavility of PPhs and other 27 phytochemicals concentration and composition, and mechanisms of action. High antioxi-28 dant capacities related with scavenging hydroxyl radical action mesure by ABTS and 29 DPPH methods were exhibited [5]. The relationship between the antimicrobial action of 30 PPhs and antioxidant capacity, it is barely analyzed during postharvest studies [4,5]. In 31 fact, PPhs favor increase of permeability of cellular wall and membrane causing their dis-32 integration, and delay of lipid peroxidation, and free radical scavenging [5]. Furthermore, 33 antimicrobial action of PPhs is associated with the presence of a free hydroxyl group, 34 bonded to a C6 aromatic ring as a system for electron delocalization. It provokes modifi-35 cation of the microbial membrane and has a key role in the inactivation of microbial en-36 zymes. These changes disturb cellular respiration and may cause cell death [20]. At the 37 same time, potassium ion leakage and reduction of H2O2 decomposition is observed, and highly reactive oxygen species, promote oxidative damage and act as proton exchangers. 39 It causes collapse of proton motive force and eventually leads to cell death [21]. For in-40stance, the antioxidant activity of in prickly pear and minimum inhibitory concentration 41 (MIC) responses had a significant negative correlation with each other [19].

4. Challenges in research and use of NA in postharvest systems

NA technologies will present a major challenge in next decades, because some tech-44 nical problems must be analized [22]. In fact, the standardization of the commercial NA 45 depends of various factors such as: variety and cultivation conditions, climatic factors; 46 plant maturity and agronomic practices, the part of the plant used, methods of extraction 47 and conservation, and techniques of chemical characterization [23]. 48

6

1

2

3

4

5

18

22

24

26

- 38
- 42

Meanwhile, some technical problems can be considered as opportunities to develop 1 the industrial applications of NA in postharvest processes and commercialization 2 [5,22,30–33] of methods for pathogen and metabolites identification, and the study about 3 mechanisms of pathogens and NA compounds will promote the compound's registration 4 and improve the use of these technologies. Furthermore, the *in vivo* practices, whether 5 conducted under semi-controlled or real conditions, must consider the composition of the 6 microbiota in fruits and vegetables. Additionally, research on the relationship between 7 physiological changes in food products and genetic control will enhance the optimization 8 of treatments. On the other hand, it is crucial to investigate the impact of nanotechnology 9 on preservation strategies and safety [5,22,30–33]. In fact, micro and nanocapsules have 10 been employed as effective components in coatings, edible films, and PPh formulations. 11 Various methods, utilizing either natural biopolymers or synthetic polymeric materials, 12 have been employed based on their specific use. They improve the physicochemical prop-13 erties of the capsules or fibers to guarantee their technofunctional properties and applica-14 tions [27,29,32,33]. For successful technological development, the selection of efficient 15 emerging methods is needed. Furthermore, the implementation of multipurpose biorefin-16 eries is proposed, accompanied by the evaluation of their feasibility, according to the cost 17 of materials, energy sustainability and environmental impact, considering the circular 18 economy principles. The production of AN is a trigger for agro-food waste revaluation, 19 so it is essential to identify and strengthen the research lines that enhance their study and 20 applications, especially from endemic sources. Furthermore, support for primary produc-21 ers in aspects of training and infrastructure must be ensured. 22

5. Conclusions

Mexican agroindustry has the challenge of technologically ensuring the management 24 and use of large volumes of agro-food waste. This involves the development of agricul-25 tural programs at the local, regional and national level based on viable crops and the re-26 design of linkage strategies with other industries. The production of HBVC is a trigger for 27 the revaluation of this biomase, so it is essential to identify and strengthen the investiga-28 tions and applications, especially from endemic sources. Some technical problems can be 29 considered as opportunities to develop the industrial applications of NA in postharvest 30 processes and commercialization. Greater attention should be given to the development 31 of nanotechnologies, due to its socioeconomic impact. Furthermore, the implementation 32 of multipurpose biorefineries is proposed, accompanied by the evaluation of their feasi-33 bility, according to the cost of materials, energy sustainability and environmental impact, 34 considering the circular economy principles. 35

Author Contributions: Conceptualization, L.M.A.V, J.A.O.R.; investigation, L.M.A.V., J.A.O.R.,36D.G.F., C.V.G., M.G.P, M.C.S, J.A.R.S data curation, L.M.A.V.; J.A.O.R. writing – original draft prep-
aration, L.M.A.V.; writing – review and editing, L.M.A.V., J.A.O.R., J.A.R.S. visualization, L.M.A.V.,37J.A.O.R.; supervision, L.M.A.V.; J.A.O.R; project administration, L.M.A.V, J.A.O.R.; funding acqui-
sition, M.G.P, M.C.S, J.A.R.S. All authors have read and agreed to the published version of the man-
uscript.39

Funding: This research received no external funding.	42
Institutional Review Board Statement: Not applicable.	43
Informed Consent Statement: Not applicable.	44
Data Availability Statement: Not applicable.	45
Acknowledgments: The authors express their gratitude to CONAHCYT and Technological University of Tecamachalco. To Ms. Paulina Aguirre-Lara for audiovisual material.	46 47
Conflicts of Interest: The authors declare no conflict of interest.	48

49

- 1. FAO El Sistema Alimentario En México-Oportunidades Para El Campo Mexicano En La Agenda 2030 de Desarrollo Sostenible 2019.
- Aguilar-Veloz, L.M.; Calderón-Santoyo, M.; Vazquez Gonzalez, Y.; Ragazzo-Sánchez, J.A. Application of Essential Oils and Polyphenols as Natural Antimicrobial Agents in Postharvest Treatments: Advances and Challenges. Food Sci Nutr 2020, 8, 2555–2568.
- Aguilar-Veloz, L.M.; Calderón-Santoyo, M.; Carvajal-Millan, E.; Martínez-Robinson, K.; Ragazzo-Sánchez, J.A. Artocarpus Heterophyllus Lam. Leaf Extracts Added to Pectin-Based Edible Coating for Alternaria Sp. Control in Tomato. LWT 2022, 156, 113022, doi:10.1016/J.LWT.2021.113022.
- 4. Aguilar-Veloz, L.M.; Calderón-Santoyo, M.; Ragazzo-Sánchez, J.A. Optimization of Microwave Assisted Extraction of Artocarpus Heterophyllus Leaf Polyphenols with Inhibitory Action against Alternaria Sp. and Antioxidant Capacity. Food Sci Biotechnol **2021**, *30*, 1695–1707, doi:10.1007/s10068-021-00996-8.
- Aguilar-Veloz, L.M.; Calderón-Santoyo, M.; Vázquez González, Y.; Ragazzo-Sánchez, J.A. Application of Essential Oils and Polyphenols as Natural Antimicrobial Agents in Postharvest Treatments: Advances and Challenges. Food Sci Nutr 2020, 8, 2555–2568, doi:https://doi.org/10.1002/fsn3.1437.
- 6. Servicio de Información Agroalimentaria y Pesquera El Brocoli Casi Un Super Alimento.
- Pacheco-Cano, R.D.; Salcedo-Hernández, R.; López-Meza, J.E.; Bideshi, D.K.; Barboza-Corona, J.E. Antimicrobial Activity of Broccoli (Brassica Oleracea Var. Italica) Cultivar Avenger against Pathogenic Bacteria, Phytopathogenic Filamentous Fungi and Yeast. J Appl Microbiol 2018, 124, 126–135, doi:10.1111/jam.13629.
- 8. Corrêa, C.B.; Martin, J.G.P.; Alencar, S.M. de; Porto, E. Antilisterial Activity of Broccoli Stems (Brassica Oleracea) by Flow Cytometry. Int Food Res J **2014**, *21*, 395.
- 9. Alizadeh Behbahani, B.; Falah, F.; Lavi Arab, F.; Vasiee, M.; Tabatabaee Yazdi, F. Chemical Composition and Antioxidant, Antimicrobial, and Antiproliferative Activities of Cinnamomum Zeylanicum Bark Essential Oil. Evidence-Based Complementary and Alternative Medicine **2020**, *2020*, 5190603, doi:10.1155/2020/5190603.
- 10. Carmen Muñoz Ezcurra Variabilidad de Los Parámetros de Calidad de Muestras Comerciales de Canela Molida. Tesis de master, Universidad Miguel Hernández de Elche: Alicante, España, **2021**.
- Cisneros-Pineda, O.; Torres-Tapia, L.W.; Gutiérrez-Pacheco, L.C.; Contreras-Martín, F.; González-Estrada, T.; Peraza-Sánchez, S.R. Capsaicinoids Quantification in Chili Peppers Cultivated in the State of Yucatan, Mexico. Food Chem 2007, 104, doi:10.1016/j.foodchem.2006.10.076.
- 12. Avilés-Betanzos, K.A.; Oney-Montalvo, J.E.; Cauich-Rodríguez, J.V.; González-Ávila, M.; Scampicchio, M.; Morozova, K.; Ramírez-Sucre, M.O.; Rodríguez-Buenfil, I.M. Antioxidant Capacity, Vitamin C and Polyphenol Profile Evaluation of a Capsicum Chinense By-Product Extract Obtained by Ultrasound Using Eutectic Solvent. Plants **2022**, *11*, doi:10.3390/plants11152060.
- Mokhtar, M.; Ginestra, G.; Youcefi, F.; Filocamo, A.; Bisignano, C.; Riazi, A. Antimicrobial Activity of Selected Polyphenols and Capsaicinoids Identified in Pepper (Capsicum Annuum L.) and Their Possible Mode of Interaction. Curr Microbiol 2017, 74, 1253–1260, doi:10.1007/s00284-017-1310-2.
- 14. Vuerich, M.; Petrussa, E.; Filippi, A.; Cluzet, S.; Fonayet, J.V.; Sepulcri, A.; Piani, B.; Ermacora, P.; Braidot, E. Antifungal Activity of Chili Pepper Extract with Potential for the Control of Some Major Pathogens in Grapevine. Pest Manag Sci **2023**, *79*, 2503–2516, doi:https://doi.org/10.1002/ps.7435.
- 15. Sreeja Devi, P.S.; Kumar, N.S.; Sabu, K.K. Phytochemical Profiling and Antioxidant Activities of Different Parts of Artocarpus Heterophyllus Lam.(Moraceae): A Review on Current Status of Knowledge. Futur J Pharm Sci **2021**, *7*, 1–7.
- 16. García, E.W.H. Posibilidad de Desarrollo y Cultivo de La Pimienta Gorda (Pimienta Dioica) En El Estado de Tabasco. Chapingo, México 1971.
- 17. Martínez, M.Á.; Evangelista, V.; Mendoza, M.; Basurto, F.; Mapes, C. Allspice [Pimenta Dioica (L.) Merrill], A Non-Timber Forest Product of Sierra Norte de Puebla, Mexico; Case Studies of Non-Timber Forest Product Systems; Center for International Forestry Research, **2004**.
- 18. Guerrero, P.C.; Majure, L.C.; Cornejo-Romero, A.; Hernández-Hernández, T. Phylogenetic Relationships and Evolutionary Trends in the Cactus Family. Journal of Heredity **2019**, *110*, 4–21.
- Iftikhar, K.; Siddique, F.; Ameer, K.; Arshad, M.; Kharal, S.; Mohamed Ahmed, I.A.; Yasmin, Z.; Aziz, N. Phytochemical Profiling, Antimicrobial, and Antioxidant Activities of Hydroethanolic Extracts of Prickly Pear (Opuntia Ficus Indica) Fruit and Pulp. Food Sci Nutr 2023, 11, 1916–1930.
- 20. Batiha, G.E.-S.; Beshbishy, A.M.; Ikram, M.; Mulla, Z.S.; El-Hack, M.E.A.; Taha, A.E.; Algammal, A.M.; Elewa, Y.H.A. The Pharmacological Activity, Biochemical Properties, and Pharmacokinetics of the Major Natural Polyphenolic Flavonoid: Quercetin. Foods **2020**, *9*, 374.
- Vázquez-González, Y.; Ragazzo-Sánchez, J.A.; Calderón-Santoyo, M. Characterization and Antifungal Activity of Jackfruit (Artocarpus Heterophyllus Lam.) Leaf Extract Obtained Using Conventional and Emerging Technologies. Food Chem 2020, 330, 127211, doi:10.1016/J.FOODCHEM.2020.127211.
- 22. Mari, M.; Bautista-Baños, S.; Sivakumar, D. Decay Control in the Postharvest System: Role of Microbial and Plant Volatile Organic Compounds. Postharvest Biol Technol **2016**, *122*, 70–81, doi:10.1016/J.POSTHARVBIO.2016.04.014.
- 23. Dini, I.; Grumetto, L. Recent Advances in Natural Polyphenol Research. Molecules 2022, 27, 8777.

1 2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

- Erazo Guijarro, M.J.; Arroyo Bonilla, F.A.; Arroyo Bonilla, D.A.; Castro García, M.R.; Santacruz Terán, S.G.; Armas Vega, A.D.C.
 Efecto Antimicrobiano Del Cinamaldehído, Timol, Eugenol y Quitosano Sobre Cepas de Streptococcus Mutans. Rev Cubana
 Estomatol 2017, 54, 1–9.
- Anaya-Castro, M.A.; Ayala-Zavala, J.F.; Muñoz-Castellanos, L.; Hernández-Ochoa, L.; Peydecastaing, J.; Durrieu, V. β-Cy- 4 clodextrin Inclusion Complexes Containing Clove (Eugenia Caryophyllata) and Mexican Oregano (Lippia Berlandieri) Essential 5 Oils: Preparation, Physicochemical and Antimicrobial Characterization. Food Packag Shelf Life **2017**, *14*, 96–101, 6 doi:10.1016/J.FPSL.2017.09.002.
- 26. Menezes, R. de P.; Bessa, M.A. de S.; Siqueira, C. de P.; Teixeira, S.C.; Ferro, E.A.V.; Martins, M.M.; Cunha, L.C.S.; Martins, C.H.G. Antimicrobial, Antivirulence, and Antiparasitic Potential of Capsicum Chinense Jacq. Extracts and Their Isolated Compound Capsaicin. Antibiotics **2022**, *11*, 1154.
- Ochoa Fuentes, Y.M.; Cerna Chávez, E.; Landeros Flores, J.; Hernández Camacho, S.; Delgado Ortiz, J.C. Evaluación in Vitro de La Actividad Antifúngica de Cuatro Extractos Vegetales Metanólicos Para El Control de Tres Especies de Fusarium Spp. Phyton (Buenos Aires) 2012, *81*, 69–73.
- 28. Barrera, A.M.P.; Pérez, M.S.R.; González, J.G.B.; Amaya-Guerra, C.A.; Román, R.Á.; Rodríguez, S.A.G. Recubrimiento Comestible a Base de Alginato En Combinación Con Eugenol Nanoencapsulado y Su Efecto Conservador En La Vida Útil de Jitomate (Solanum Lycopersicum). Biotecnia **2021**, 23.
- Solís-Silva, A.; Reyes-Munguía, A.; Madariaga-Navarrete, G.; Medina-Pérez, R.G.; Campos-Montiel, A.J.; Cenobio-Galindo, J. Evaluación de La Actividad Antifúngica y Antioxidante de Una Nanoemulsión W/O de Opuntia Oligacantha y Aceite Esencial de Citrus X Sinensis. Investigación y Desarrollo en Ciencia Y Tecnología de Alimentos 2018, 3, 182–187.
- 30. Albonico, M.; Schutz, L.F.; Caloni, F.; Cortinovis, C.; Spicer, L.J. In Vitro Effects of the Fusarium Mycotoxins Fumonisin B1 and Beauvericin on Bovine Granulosa Cell Proliferation and Steroid Production. Toxicon 2017, *128*, 38–45.
- 31. Basak, S.; Guha, P. A Review on Antifungal Activity and Mode of Action of Essential Oils and Their Delivery as Nano-Sized Oil Droplets in Food System. J Food Sci Technol **2018**, *55*, 4701–4710.
- 32. Kothalawala, S.G.; Sivakumaran, K. An Overview of Nanotechnology Applications in Food Industry. Int. J. Sci. Res. Publ **2018**, *8*, 340.
- 33. Liu, W.; Zhang, M.; Bhandari, B. Nanotechnology A Shelf Life Extension Strategy for Fruits and Vegetables. Crit Rev Food Sci Nutr **2020**, *60*, 1706–1721, doi:10.1080/10408398.2019.1589415.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27 28

29

30