The 4th International Electronic Conference on Applied Sciences (Session: Food Science and Technology; 10/27/2023 – 11/10/2023)



Anti-mycotoxigenic efficacy of redox-active natural compounds and derivatives

Jong H. Kim*, Kathleen L. Chan, DeAngela Ford, Foodborne Toxin Detection and Prevention Research Unit Western Regional Research Center, USDA-ARS 800 Buchanan St., Albany, CA 94710, USA

*Email: jongheon.kim@usda.gov



Background

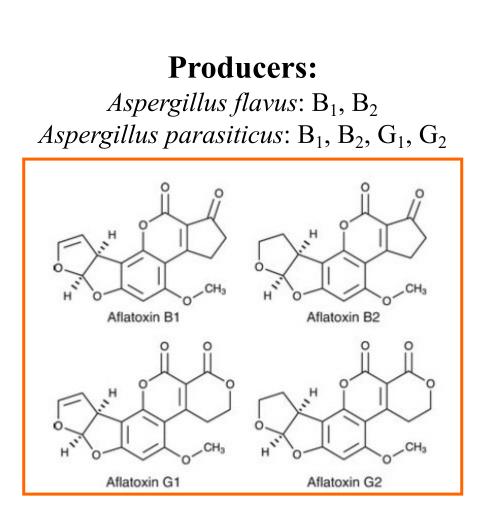
Crops such as tree nuts, peanuts, etc., are susceptible to infestation by the mold *Aspergillus flavus* and *Aspergillus parasiticus* and subsequent aflatoxin (AF) contamination

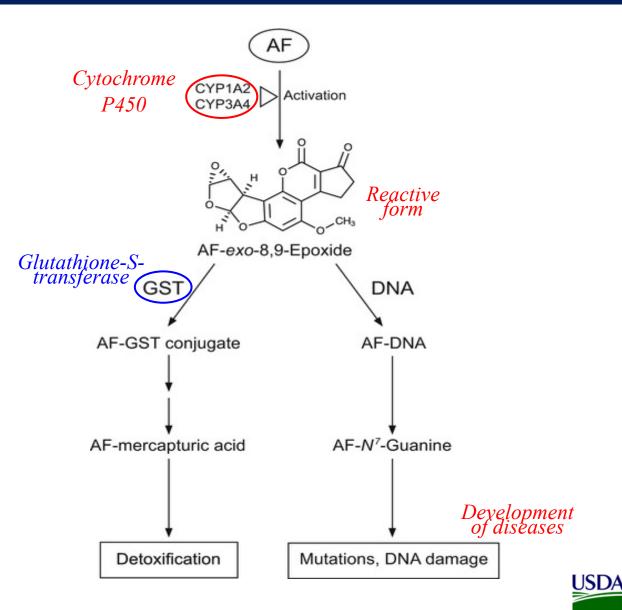


- AF is a serious threat to food safety; many importing countries imposing limits as low as 4 ppb
- The EU's Rapid Alert System for Food and Feed (RASFF; <u>https://food.ec.europa.eu/safety/rasff_en</u>) continually issued border rejection on the AFcontaminated food products imported
- Although the exporting countries set strict standards to ensure the food products maintain high level of quality, it is often difficult to determine the **root causes of AF contamination**



Producers and toxicity of AFs





Fungicide resistance and mycotoxins

• Traditional control of *A. flavus* and *A. parasiticus* has been through the application of fungicides; **fungicide resistance** have been correlated with increased mycotoxin production

Examples of fungicide-potentiation of mycotoxin production in resistant pathogens (Adapted from Kim et al. 2015. Outlooks on Pest Management. 26:172-176)

Fungi

Aspergillus parasiticus Fusarium graminearum Fusarium culmorum Fusarium sporotrichioides Penicillium expansum Penicillium verrucosum

Fungicides

Anilinopyrimidine Benzimidazole Carbendazim Fludioxonil Flusilazole Iprodione Phenylpyrrole Strobilurins Tebuconazole

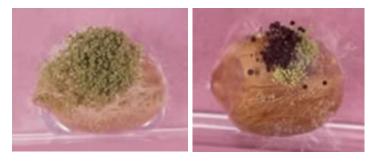
Mycotoxins

Aflatoxin Citrinin Patulin Trichothecene



Example: Prevalence of fungicide resistant A. flavus (Ali et al. 2021. J of Fungi. 7: 284)

- In 2019: High frequency of **AF contamination** in peanuts (USA)
- In 2020: **Poor seed quality**; peanut seeds had lower than expected germination and a high frequency of *A*. *flavus* contamination
- **Mitochondrial** *cytochrome b* gene mutations: These *A. flavus* had a single nucleotide mutations at CytB G143A (36.8% isolates) and at CytB F129L (15.8% isolates), resulting in fungicide resistance
- Ineffectiveness of current seed treatments: Thus, **needs for new fungicides** to avoid AF contamination



Aspergillus-contaminated peanut seeds (Courtesy: USDA)

USDA

Rationale

- **Natural compounds** that do not have any significant environmental impact are a potential source of antimycotic agents, either in their nascent form or as lead structures for more effective derivatives
- Especially, **natural phenolic compounds** can serve as potent **redox cyclers** that inhibit microbial growth through destabilization of cellular **redox homeostasis** and/or **antioxidation systems**
- However, as determined in commercial fungicides, if **natural phenolic compounds** are applied at **suboptimal concentrations** than that required for fungal control, the compounds would **potentiate mycotoxin production** by *A. flavus* and *A. parasiticus*
- In this study, two sub-inhibitory concentrations of **natural phenolic compounds/derivatives** were tested against *A. flavus* and *A. parasiticus* for determining their anti-mycotoxigenecity & potentiation of mycotoxin production (**Risk assessment**)



Materials and Methods

• AF determination/analysis:

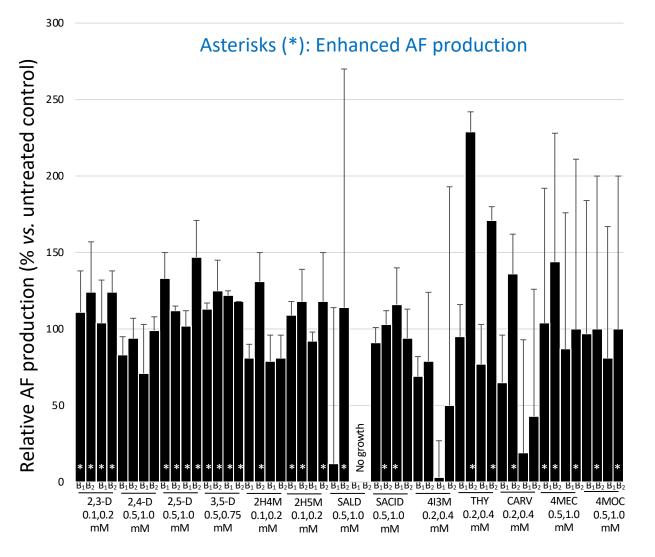
Aflatoxin determination was performed (w/ or w/o treatments of natural phenolic compounds) using an Agilent 1260 system (Palo Alto, CA, USA) with fluorescence detection at 365 nm excitation and 455 nm emission, as previously described Hua et al. 2019. *Mycotoxin Res.* 35: 381-389. <u>https://doi.org/10.1007/s12550-019-00364-w</u>. Analysis was performed using OpenLAB CDS Chemstation Edition for LC & LC/MS Systems (Rev. C.01.08) (Agilent Technologies, Palo Alto, CA, USA).

• Natural phenolic compounds/derivatives examined:

| Compounds | Low concentration (mM) | High concentration (mM) |
|--|------------------------|-------------------------|
| 2,3-Dihydroxybenzaldehyde (2,3-D) | 0.1 | 0.2 |
| 2,4-Dihydroxybenzaldehyde (2,4-D) | 0.5 | 1.0 |
| 2,5-dihydroxybenzaldehyde (2,5-D) | 0.5 | 1.0 |
| 3,5-Dimethoxybenzaldehyde (3,5-D) | 0.5 | 0.75 |
| 2-Hydroxy-4-methoxybenzaldehyde (2H4M) | 0.1 | 0.2 |
| 2-Hydroxy-5-methoxybenzaldehyde (2H5M) | 0.1 | 0.2 |
| Salicylaldehyde (SLAD) | 0.5 | 1.0 |
| Salicylic acid (SACID) | 0.5 | 1.0 |
| 4-Isopropyl-3-methylphenol (4I3M) | 0.2 | 0.4 |
| Thymol (2-Isopropyl-5-methylphenol; THY) | 0.2 | 0.4 |
| Carvacrol (5-Isopropyl-2-methylphenol; CARV) | 0.2 | 0.4 |
| 4-Methylcinnamic acid (4MEC) | 0.5 | 1.0 |
| 4-Methoxycinnamic acid (4MOC) | 0.5 | 1.0 |



Results: AF production in A. flavus

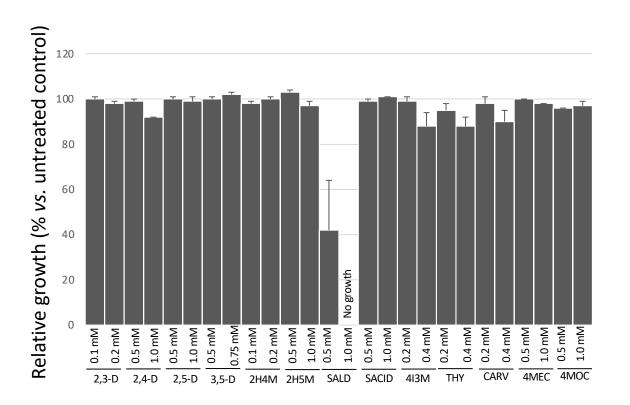


KEYS:

- Twenty-seven (marked as "asterisks") out of fifty-two concentrations of test compounds exhibited enhanced AF (B₁, B₂) production compared to the untreated control
- 4I3M and CARV at 0.4 mM showed the most potent antimycotoxigenic activity whereas THY at 0.2 mM exhibited the highest mycotoxin enhancement
- 4I3M and 2,4-Dihydroxybenzaldehyde (2,4-D) did not enhance AF production at all concentrations (0.5, 1.0 mM) tested; viz., no risks identified in *A. flavus*
- AFB₁ and AFB₂ production seems to be **differentially** influenced by test compounds at different concentrations

USDA

Results: A. flavus growth

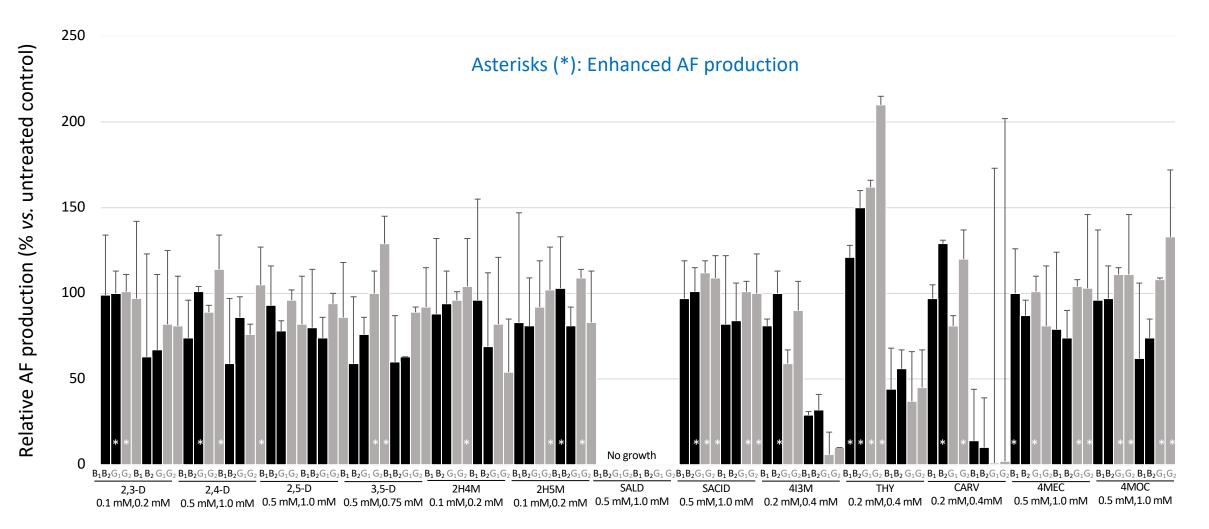


KEYS:

• Except salicylaldehyde (SALD), the concentrations of most compounds **marginally** affected the growth of *A. flavus*



Results: AF production in A. parasiticus





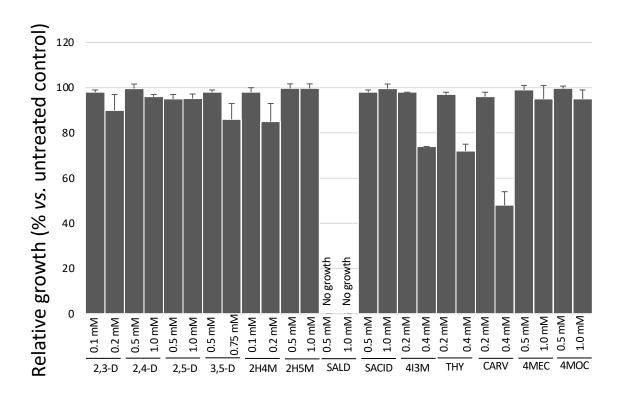
Results (continued): AF production in A. parasiticus

KEYS:

- Thirty-one (marked as "asterisks") out of fifty-two concentrations of test compounds exhibited enhanced AF (B₁, B₂, G₁, G₂) production compared to untreated control
- As determined in *A. flavus*, 4I3M and CARV at 0.4 mM showed the most potent antimycotoxigenic activity while THY at 0.2 mM exhibited the highest mycotoxin enhancement
- Except AFB₁ at 0.5 mM, 2,5-Dihydroxybenzaldehyde (2,5-D) did not enhance AF production at all concentrations (0.5, 1.0 mM) tested
- As determined in *A. flavus*, AFB₁, AFB₂, AFG₁ and AFG₂ production seems to be differentially influenced by test compounds at different concentrations



Results: A. parasiticus growth



KEYS:

• Except salicylaldehyde (SALD; 0.5 & 1.0 mM), 4I3M (0.4 mM), THY (0.4 mM) and CARV (0.4 mM), the concentrations of most compounds tested **marginally affected the growth** of *A. parasiticus*



Summary

- In both *A. flavus* and *A. parasiticus*, **4I3M** and **CARV** at 0.4 mM showed the **most potent** antimycotoxigenic activity while **THY** at 0.2 mM exhibited the highest **mycotoxin enhancement**
- Twenty-seven (*A. flavus*) to thirty-one (*A. parasiticus*) out of fifty-two concentrations of natural phenolic compounds tested exhibited enhanced AF (B₁, B₂, G₁, G₂) production compared to the untreated control
- **2,4-Dihydroxybenzaldehyde (2,4-D)** or **2,5-Dihydroxybenzaldehyde (2,5-D)** did not enhance AF production at all concentrations (0.5, 1.0 mM) tested in *A. flavus* or *A. parasiticus*, respectively (except AFB₁ at 0.5 mM in *A. parasiticus*)
- AFB₁, AFB₂, AFG₁ and AFG₂ production were **differentially influenced/regulated** by test compounds at different concentrations
- In conclusion, **natural phenolic agents** could be applied as anti-mycotoxigenic agents but **should be used at optimum concentrations**, thus preventing the enhanced AF production in *A. flavus* and *A. parasiticus*



Acknowledgement

WRRC, USDA-ARS, Albany, CA, USA

Kathleen Chan, DeAngela Ford Siov Sarrell, Jeff Palumbo William Hart-Cooper, William J. Orts

Almond Board of California, Modesto, CA, USA Guangwei Huang (Associate Director, Food Research & Technology)

Ipura Food Safety, Fresno, CA, USA Keith Meeks (CEO)

