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Synthesis of Some 2,6-Disubstituted 4-Pyridinecarboxamides and -Carbothioamides, and Their Antimycobacterial and Photosynthesis-Inhibiting Activity

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Abstract: A group of 26 new 6-halogeno- or 6-alkylsulfanyl-2-alkylsulfanylpyridine-4-carboxamides and corresponding carbothioamides was synthesised. Some of the amides and all thioamides were tested for their antimycobacterial activity against atypical mycobacterial strains. Promising photosynthesis-inhibiting activity was also found for some of the amides.

Keywords: Pyridininecarbothioamides, antimycobacterial activity, photosynthesis-inhibiting activity.

Introduction Results and Discussion Experimental References

Introduction

Some events during the past decade have dramatically changed the nature and magnitude of the problem of tuberculosis. The HIV epidemic and increasing resistance to antituberculous drugs dictate the need of development of new antituberculotics [1 -- 3].

In our recent study [4], we modified the structure of therapeutically used antituberculous drugs ethionamide and prothionamide. Some of the more lipophilic derivatives showed promising activity against atypical mycobacterial strains. The present study extends the scope of lipophilic derivatives of 4-pyridinecarbothioamide. Since it has been recently reported that 2-alkylsulfanylpyridine-4-carbothioamides showing antimycobacterial [5] and antifungal activity [6] inhibit photosynthetic processes in algae and plant chloroplasts [7, 8], the synthesised compounds were tested for their both antimycobacterial and photosynthesis-inhibiting activity.

Results and Discussion

The synthesis of 2,6-disubstituted 4-pyridinecarboxamides and -carbothioamides is shown in Scheme 1. 2,6-Dichloro- or -dibromopyridine-4-carboxamide [9] was treated with an equimolar amount of the respective thiolate to give 2-alkylsulfanyl-6-halogenopyridine-4-carboxamides (**1** -- **8**). 6-Alkylsulfanyl-2hexylsulfanylpyridine-4-carboxamides (**9** -- **13**) were prepared from 6-chloro-2-hexylsulfanylpyridine-4carboxamide (**3**) in a similar fashion. Thionation of carboxamides (**1** -- **13**) with the Lawesson's reagent afforded the thioamides (**14** -- **26**). The melting points, yields, and elemental analyses for compounds **1** --**26** are given in Table 1, and IR and ¹H NMR spectroscopic data in Table 2.

Scheme 1.



Selected compounds were evaluated for their activity against *Mycobacterium tuberculosis*, *M. kansasii*, *M. avium*, and *M. fortuitum*. The minimum inhibitory concentrations (MIC) for the tested compounds are listed in Table 3, along with isoniazid as a reference standard. Compounds in the amide series (1, 4, 7, 9, 12) were inactive with the exception of 9 and, in part, 7. Compound 9 showed greater activity against atypical strains, especially *M. kansasii* (MIC = 60 μ mol dm⁻³), than isoniazid.

The thioamide series was more active in the antimycobacterial screening than the amide one. 2-Alkylsulfanyl-6-halogenopyridine-4-carbothioamides (**14** -- **21**) exhibited increasing antimycobacterial activity with prolonging the carbon chain in the alkylsulfanyl substituent up to seven carbons. Overall, 6chloro substituted thioamides were more active than their 6-bromo analogues. Compounds **16** and **20** were the most promising, with MICs of 60 μ mol dm⁻³ against *M. tuberculosis* and *M. kansasii* (as well as *M. avium* for **16**).

Among 2-alkylsulfanyl-6-hexylsulfanylpyridine-4-carbothioamides (**22** -- **26**), compound **22** was found to be the most active of the compounds studied. It showed, similar to compounds **23** and **24**, moderate activity against *M. tuberculosis* (MIC = 30 μ mol dm⁻³). Additionally, it exhibited greater activity against *M.*

kansasii (MIC = 60 μ mol dm⁻³), *M. avium* (MIC = 60 μ mol dm⁻³), and *M. fortuitum* (MIC = 250 μ mol dm⁻³) than isoniazid, as well as **16**. Increasing the total number of carbon atoms in both alkylsulfanyl side chains above ten caused a decrease in antimycobacterial activity, which is in agreement with our previous findings [4].

To better understand the structure -- activity relationships, log P values were calculated (Table 3). We found that the lipophilicities of the most potent antimycobacterial compounds were different for all four strains employed. In the thioamide series (**14** -- **26**), the highest activities against *M. tuberculosis* were observed for compounds **22** -- **24** with log P values between 5.75 and 7.34. The antimycobacterial activity of thioamides against other three strains showed a sharp dependence on lipophilicity. In the case of *M. kansasii* and *M. avium*, the most active thioamides **16** and **22** showed log P values ranging from 4.86 to 5.75, while the compounds with the highest activity against *M. fortuitum*, **23** and **24**, exhibited log P values 6.81 and 7.34.

The tested compounds also inhibited photochemical activity of spinach chloroplasts. The IC₅₀ values, *i. e.*, concentrations of the compounds causing 50% decrease of oxygen evolution rate in spinach chloroplasts with respect to the untreated control, are listed in Table 3. From the comparison of IC₅₀ values of the 6-halogeno substituted amides (1 -- 7) and thioamides (14 -- 21) it can be concluded that amides exhibit greater inhibitory activity than the corresponding thioamides. For compounds 9 -- 12, a pronounced decrease in photosynthesis-inhibiting activity with the increasing lipophilicity of the compounds has been confirmed. This is in good agreement with the previously obtained results concerning photosynthesis-inhibiting activity of 2-alkylsulfanylpyridine-4-carbothioamides in spinach chloroplasts and *Chlorella vulgaris* [7, 8]. In the carboxamide series, the most active compounds 7, 9, 3, and 6 showed log P in the range of 3.12 -- 5.0, whereas the inhibitory activity of thioamides with log P > 3.27 showed a pronounced decrease.

Using EPR spectroscopy it was found that in the suspension of spinach chloroplasts the studied thioamides interact with D^+ intermediate, i.e., with the radical of tyrosine 161 (Tyr_D) which is located in D_2 protein on the donor side of photosystem 2, [10] and due to this interaction the photosynthetic electron transport from the oxygen evolving complex to the core of photosystem 2 is impaired. The same site of action in the photosynthetic apparatus of spinach chloroplasts has also been confirmed for the structurally similar 2-alkylsulfanylpyridine-4-carbothioamides [7].

		ł	$R^2 \xrightarrow{N} SR^1$ $C \xrightarrow{-NH_2}$						
Compd.	Formula M. w.	R ¹ R ²	X	M. p. °C Yield %		% Calculated % Found			
					С	Н	N	S	CI (Br)
1	C ₈ H ₉ CIN ₂ OS (216.7)	C ₂ H ₅ , Cl	0	162163 75	44.34 44.21	4.19 4.12	12.93 13.11	14.80 14.69	16.36 16.51
2	C ₉ H ₁₁ CIN ₂ OS (230.7)	C ₃ H ₇ , Cl	0	112113 76	46.85 46.65	4.81 4.73	12.14 12.35	13.90 13.62	15.37 15.50
3	C ₁₂ H ₁₇ CIN ₂ OS (272.8)	C ₆ H ₁₃ , Cl	0	129131 72	52.84 52.76	6.28 6.21	10.27 10.39	11.75 11.64	13.00 13.14
4	C ₇ H ₇ BrN ₂ OS	СН ₃ ,	0	178180	34.02	2.86	11.34	12.97	32.34

Table 1.	Analytical	data of th	ne prepared	compounds
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	(247.1)	Br		70	<mark>33.96</mark>	<mark>2.78</mark>	11.46	<mark>12.89</mark>	32.48
5	C ₈ H ₉ BrN ₂ OS (261.1)	C ₂ H ₅ , Br	0	160162 73	36.80 36.66	3.47 3.35	10.73 10.85	12.28 12.18	30.60 30.76
6	C ₁₀ H ₁₃ BrN ₂ OS (289.2)	C ₄ H ₉ , Br	0	113115 71	41.53 41.31	4.53 4.41	9.69 9.90	11.09 10.81	27.63 27.81
7	C ₁₃ H ₁₉ BrN ₂ OS (331.3)	C ₇ H ₁₅ , Br	0	120122 68	47.13 47.01	5.78 5.71	8.46 8.57	9.68 9.58	24.12 24.26
8	C ₁₄ H ₂₁ BrN ₂ OS (345.3)	C ₈ H ₁₇ , Br	0	123125 65	48.70 48.43	6.13 6.01	8.11 8.31	9.28 9.11	23.14 23.26
9	C ₁₄ H ₂₂ N ₂ OS ₂ (298.5)	C ₆ H ₁₃ , SC ₂ H ₅	0	8384 75	56.34 56.29	7.43 7.41	9.39 9.45	21.48 21.43	
10	C ₁₆ H ₂₆ N ₂ OS ₂ (326.5)	C ₆ H ₁₃ , SC ₄ H ₉	0	8688 72	58.86 58.93	8.03 8.09	8.58 8.51	19.64 19.71	
11	C ₁₇ H ₂₈ N ₂ OS ₂ (340.6)	C ₆ H ₁₃ , SC ₅ H ₁₁	0	109111 70	59.96 60.05	8.29 8.34	8.23 8.17	18.83 18.92	
12	C ₁₉ H ₃₂ N ₂ OS ₂ (368.6)	C ₆ H ₁₃ , SC ₇ H ₁₅	0	106108 67	61.91 61.93	8.75 8.72	7.60 7.55	17.40 17.43	
13	C ₂₀ H ₃₄ N ₂ OS ₂ (382.6)	C ₆ H ₁₃ , SC ₈ H ₁₇	0	9698 64	62.78 62.67	8.96 8.90	7.32 7.38	16.76 16.67	
14	C ₈ H ₉ CIN ₂ S ₂ (232.8)	C ₂ H ₅ , Cl	S	8081 89	41.28 41.16	3.90 3.81	12.04 12.17	27.55 27.41	15.23 15.38
15	C ₉ H ₁₁ CIN ₂ S ₂ (246.8)	C ₃ H ₇ , Cl	S	oil 87	43.81 43.73	4.49 4.45	11.35 11.43	25.98 25.87	14.37 14.49
16	C ₁₂ H ₁₇ CIN ₂ S ₂ (288.9)	C ₆ H ₁₃ , Cl	S	4547 91	49.90 49.69	5.93 5.78	9.70 9.81	22.20 22.01	12.27 12.42
17	C ₇ H ₇ BrN ₂ S ₂ (263.2)	CH ₃ , Br	S	127129 85	31.95 31.82	2.68 2.61	10.64 10.51	24.36 24.22	30.36 30.48
18	C ₈ H ₉ BrN ₂ S ₂ (277.2)	C ₂ H ₅ , Br	S	107108 92	34.66 34.58	3.27 3.22	10.11 10.03	23.13 22.96	28.83 28.98
19	C ₁₀ H ₁₃ BrN ₂ S ₂ (305.3)	C ₄ H ₉ , Br	S	5355 90	39.35 39.15	4.29 4.20	9.18 9.07	21.01 20.89	26.18 26.39
20	C ₁₃ H ₁₉ BrN ₂ S ₂ (347.3)	C ₇ H ₁₅ , Br	S	4345 89	44.96 44.78	5.51 5.47	8.07 8.15	18.46 18.27	23.01 23.27
21	C ₁₄ H ₂₁ BrN ₂ S ₂ (361.4)	C ₈ H ₁₇ , Br	S	4446 91	46.53 46.31	5.86 5.77	7.75 7.92	17.74 17.59	22.11 22.35
22	C ₁₄ H ₂₂ N ₂ S ₃ (314.5)	C ₆ H ₁₃ , SC ₂ H ₅	S	oil 90	53.46 53.41	7.05 7.02	8.91 9.03	30.58 30.41	
23	C ₁₆ H ₂₆ N ₂ S ₃ (342.6)	C ₆ H ₁₃ , SC ₄ H ₉	S	5860 91	56.10 56.25	7.65 7.71	8.18 8.03	28.08 28.23	
24	C ₁₇ H ₂₈ N ₂ S ₃ (356.6)	C ₆ H ₁₃ , SC ₅ H ₁₁	S	6263 89	57.26 57.02	7.91 7.78	7.86 8.05	26.97 26.72	
25	C ₁₉ H ₃₂ N ₂ S ₃	C ₆ H ₁₃ ,	S	7173	59.33	8.39	7.28	25.00	

	(384.7)	SC ₇ H ₁₅		89	<mark>59.47</mark>	<mark>8.42</mark>	7.15	<mark>24.78</mark>	
26	C ₂₀ H ₃₄ N ₂ S ₃ (398.7)	C ₆ H ₁₃ , SC ₈ H ₁₇	S	6264 87	60.25 60.03	8.60 8.47	7.03 7.25	24.12 23.91	

 Table 2. IR and ¹H NMR spectroscopic data of the prepared compounds

Compd.	IR (cm ⁻¹)	¹ H NMR delta (ppm)				
1	3019, 2972 (CH aliph.) 1690 (C=O)	7.38 d, J = 1, 1 H, arom.; 7.27 d, J = 1, 1 H, arom.; 6.09 bs, 1 H, NH; 5.90 bs, 1 H, NH; 3.18 q, J = 7, 2 H, SCH ₂ ; 1.37 t, J = 7, 3 H, CH ₃				
2	3013, 2968 (CH aliph.) 1689 (C=O)	7.38 d, J = 1, 1 H, arom.; 7.27 d, J = 1, 1 H, arom.; 6.53 bs, 1 H, NH; 6.42 bs, 1 H, NH; 3.14 t, J = 7, 2 H, SCH ₂ ; 1.73 sext, J = 7, 2 H, CH ₂ ; 1.02 t, J = 7, 3 H, CH ₃				
3	3014, 2959, 2931 (CH aliph.) 1689 (C=O)	7.38 d, J = 1, 1 H, arom.; 7.27 d, J = 1, 1 H, arom.; 6.19 bs, 2 H, NH ₂ ; 3.15 t, J = 7, 2 H, SCH ₂ ; 1.69-1.26 m, 4 H, $(CH_2)_2$; 0.87 t, J = 7, 3 H, CH ₃				
4	3019, 2970 (CH aliph.) 1696 (C=O)	7.44-7.46 m, 2 H, arom.; 6.15 bs, 1 H, NH; 5.75 bs, 1 H, NH; 2.60 s, 3 H, CH ₃				
5	3018, 2969, 2936 (CH aliph.) 1695 (C=O)	7.40-7.42 m, 2 H, arom.; 6.06 bs, 1 H, NH; 5.88 bs, 1 H, NH; 3.18 q, J = 7, 2 H, SCH ₂ ; 1.37 t, J = 7, 3 H, CH ₃				
6	3014, 2962, 2933 (CH aliph.) 1694 (C=O)	7.40-7.42 m, 2 H, arom.; 6.24 bs, 2 H, NH_2 ; 3.14 t, J = 7, 2 H, SCH ₂ ; 1.73 m, 2 H, CH ₂ ; 1.44 m, 2 H, CH ₂ ; 0.92 dist. t, J = 5, 3 H, CH ₃				
7	3014, 2958, 2930 (CH aliph.) 1690 (C=O)	7.40-7.42 m, 2 H, arom.; 6.19 bs, 2 H, NH_2 ; 3.14 t, J = 7, 2 H, SCH_2 ; 1.70-1.25 m, 10 H, $(CH_2)_5$; 0.87 dist. t, J = 5, 3 H, CH_3				
8	3013, 2957, 2929 (CH aliph.) 1689 (C=O)	7.39-7.41 m, 2 H, arom.; 6.14 bs, 2 H, NH_2 ; 3.14 t, J = 7, 2 H, SCH_2 ; 1.70-1.26 m, 12 H, $(CH_2)_6$; 0.87 dist. t, J = 5, 3 H, CH_3				
9	3013, 2960, 2931 (CH aliph.) 1686 (C=O)	7.14 s, 2 H, arom.; 6.32 bs, 2 H, NH_2 ; 3.21 q overlapping with 3.17 t, 4 H both, 2 x SCH ₂ ; 1.1 - 1.9 m, 8 H, 4 x CH ₂ ; 1.38 t, J = 7, 3 H, SCH ₂ CH ₃ ; 0.90 dist. t, J = 5, 3 H, CH ₃				
10	3010, 2961, 2932 (CH aliph.) 1686 (C=O)	7.14 s, 2 H, arom.; 6.03 bs, 2 H, NH_2 ; 3.20 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1 - 1.9 m, 12 H, 6 x CH ₂ ; 0.95 t, J = 6, 3 H, S(CH ₂) ₃ CH ₃ ; 0.90 dist. t, J = 5, 3 H, CH ₃				
11	3010, 2960, 2931 (CH aliph.) 1686 (C=O)	7.14 s, 2 H, arom.; 6.14 bs, 2 H, NH_2 ; 3.19 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1 - 1.9 m, 14 H, 7 x CH ₂ ; 0.90 dist. t, J = 5, 6 H, 2 x CH ₃				
12	3009, 2959, 2930 (CH aliph.) 1686 (C=O)	7.14 s, 2 H, arom.; 6.02 bs, 2 H, NH_2 ; 3.19 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1 - 1.9 m, 18 H, 9 x CH ₂ ; 0.90 dist. t, J = 5, 6 H, 2 x CH ₃				
13	3010, 2959, 2929 (CH aliph.) 1686 (C=O)	7.14 s, 2 H, arom.; 6.20 bs, 2 H, NH_2 ; 3.19 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1 - 1.9 m, 20 H, 10 x CH ₂ ; 0.90 dist. t, J = 5, 6 H, 2 x CH ₃				
14	2991, 2931, 2874 (CH aliph.)	7.64 bs, 1 H, NH; 7.37 d, J = 1, 1 H, arom.; 7.29 d, J = 1, 1 H, arom.; 7.3 bs, 1 H, NH; 3.18 q, J = 7, 2 H, SCH ₂ ; 1.38 t, J = 7, 3				

	1603 (C=O)	Н, СН ₃
15	2996, 2968, 2934 (CH aliph.) 1603 (C=O)	7.7 bs, 1 H, NH; 7.38 d, J = 1, 1 H, arom.; 7.29 d, J = 1, 1 H, arom.; 7.2 bs, 1 H, NH; 3.17 t, J = 7, 2 H, SCH ₂ ; 1.74 sext, J = 7, 2 H, CH ₂ ; 1.05 t, J = 7, 3 H, CH ₃
16	2996, 2959, 2931 (CH aliph.) 1603 (C=O)	7.82 bs, 1 H, NH; 7.36 d, J = 1, 1 H, arom.; 7.28 d, J = 1, 1 H, arom.; 7.3 bs, 1 H, NH; 3.17 t, J = 7, 2 H, SCH ₂ ; 1.69-1.26 m, 8 H, $(CH_2)_4$; 0.90 dist. t , J = 5, 3 H, CH ₃
17	3001, 2932 (CH aliph.) 1603 (C=O)	7.76 bs, 1 H, NH; 7.40 d, J = 1, 1 H, arom.; 7.29 d, J = 1, 1 H, arom.; 7.26 bs, 1 H, NH; 2.59 s, 3 H, CH ₃
18	2992, 2932 (CH aliph.) 1603 (C=O)	7.79 bs, 1 H, NH; 7.39 d, J = 1, 1 H, arom.; 7.29 d, J = 1, 1 H, arom.; 7.26 bs, 1 H, NH; 3.19 q, J = 7, 2 H, SCH ₂ ; 1.38 t, J = 7, 3 H, CH ₃
19	2999, 2962, 2933 (CH aliph.) 1603 (C=O)	7.88 bs, 1 H, NH; 7.37 d, J = 1, 1 H, arom.; 7.29 d, J = 1, 1 H, arom.; 7.3 bs, 1 H, NH; 3.18 t, J = 7, 2 H, SCH ₂ ; 1.25-1.86, 4 H, $(CH_2)_2$; 0.95 t, J = 6, 3 H, CH ₃
20	2997, 2958, 2929 (CH aliph.) 1603 (C=O)	7.85 bs, 1 H, NH; 7.38 d, J = 1, 1 H, arom.; 7.29 d, J = 1, 1 H, arom.; 7.3 bs, 1 H, NH; 3.17 t, J = 7, 2 H, SCH ₂ ; 1.25-1.86, 10 H, $(CH_2)_5$; 0.89 dist. t, J = 5, 3 H, CH ₃
21	2998, 2957, 2928 (CH aliph.) 1603 (C=O)	7.77 bs, 1 H, NH; 7.37 d, J = 1, 1 H, arom.; 7.28 d, J = 1, 1 H, arom.; 7.26 bs, 1 H, NH; 3.16 t, J = 7, 2 H, SCH ₂ ; 1.2-1.8 m, 12 H, $(CH_2)_6$; 0.88 dist. t, J = 5, 3 H, CH ₃
22	3004, 2960, 2930 (CH aliph.) 1601 (C=O)	7.13 s, 2 H, arom.; 6.1 bs, 1 H, NH; 5.8 bs, 1 H, NH; 3.20 q overlapping with 3.19 t, 4 H both, 2 x SCH ₂ ; 1.2 -1.8 m, 8 H, 4 x CH ₂ ; 1.34 t, J = 7, 3 H, SCH ₂ CH ₃ ; 0.90 dist. t, J = 5, 3 H, CH ₃
23	2996, 2961, 2931 (CH aliph.) 1601 (C=O)	7.76 bs, 1 H, NH; 7.13 s, 2 H, arom.; 7.2 bs, 1 H, NH; 3.19 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1-1.9 m, 12 H, 6 x CH ₂ ; 0.95 t, J = 6, 3 H, $S(CH_2)_3CH_3$; 0.90 dist. t, J = 5, 3 H, CH ₃
24	2995, 2960, 2931 (CH aliph.) 1601 (C=O)	7.76 bs, 1 H, NH; 7.13 s, 2 H, arom.; 7.2 bs, 1 H, NH; 3.19 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1-1.9 m, 14 H, 7 x CH ₂ ; 0.90 dist. t, J = 5, 6 H, 2 x CH ₃
25	2996, 2959, 2930 (CH aliph.) 1601 (C=O)	7.61 bs, 1 H, NH; 7.13 s, 2 H, arom.; 7.2 bs, 1 H, NH; 3.19 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1-1.9 m, 18 H, 9 x CH ₂ ; 0.90 dist. t, J = 5, 6 H, 2 x CH ₃
26	2995, 2958, 2929 (CH aliph.) 1601 (C=O)	7.66 bs, 1 H, NH; 7.13 s, 2 H, arom.; 7.2 bs, 1 H, NH; 3.18 t, J = 7, 4 H, 2 x SCH ₂ ; 1.1-1.9 m, 20 H, 10 x CH ₂ ; 0.90 dist. t, J = 5, 6 H, 2 x CH ₃

Table 3. Antimycobacterial activity, inhibition of oxygen evolution rate in spinach chloroplasts, and lipophilicity of the prepared compounds

		MIC	IC ₅₀	
Compd		(µmol dm ⁻³)	(umol dm^{-3})	Calculated
	Compa.	M. tuberculosis M. kansasii M. avium M. fortuitum	spinach	log P

	H ₃₇ R _v	PKG8	80/72	1021	chloroplasts	
1	500	1000	1000	1000	101.5	1.96
2					58.4	2.49
3					10.2	4.08
4	> 1000	> 1000	> 1000	> 1000	76.7	1.52
5					34.2	2.06
6					10.6	3.12
7	250	250	> 1000	> 1000	5.9	4.71
8						5.24
9	60	60	250	250	9.1	5.00
10					203.5	6.06
11					249.3	6.59
12	1000	> 1000	> 1000	> 1000	543.6	7.66
13					258.8	8.19
14	125	250	250	500	104.8	2.73
15	125	125	250	500	9.3	3.27
16	60	60	60	250	29.8	4.86
17	500	500	500	1000	187.7	2.34
18	250	250	500	500	19.6	2.87
19	125	250	250	500	20.9	3.93
20	60	60	125	500	61.0	5.52
21	125	125	250	500	105.1	6.06
22	30	60	60	250		5.75
23	30	125	125	125	99.7	6.81
24	30	250	125	125	157.5	7.34
25	60	250	250	500		8.41
26	125	250	250	500		8.94
isoniazid	7	250	1000	1000		-0.89

Experimental

General

Melting points were determined on a Kofler block, and are uncorrected. IR spectra were recorded on a Nicolet Impact 400 spectrometer in chloroform. ¹H NMR spectra were determined for solutions in CDCl₃ with TMS as the internal standard with a BS 587 (Tesla, Brno) 80 MHz apparatus. Column chromatography was performed on silica gel (Silpearl, Kavalier Votice). Elemental analyses were performed on a EA 1110 CHNS-O CE INSTRUMENTS elemental analyser.

Lipophilicity of the compounds was computed using a program ACD/LogP version 1.0 (Advanced Chemistry Development Inc., Toronto).

Synthesis of 2-alkysufanyl-6-halogenopyridine-4-carboxamides 1 -- 8

2,6-Dichloro- or 2,6-dibromopyridine-4-carboxamide [9] (10 mmol) and the appropriate thiol (10 mmol)

were dissolved in anhydrous *N*,*N*-dimethylformamide (10 ml). To the stirred solution was added a sodium methoxide (10 mmol) solution dropwise. (In preparing 2-methylsulfanyl-6-bromopyridine-4-carboxamide, sodium methanethiolate (10 mmol) was added in several portions to the stirred solution of 2,6-dibromopyridine-4-carboxamide (10 mmol) in anh. *N*,*N*-dimethylformamide.) The reaction mixture was stirred at room temperature until TLC indicated a complete reaction. TLC was performed using petroleum ether : ethyl acetate (2:1) as the mobile phase. The mixture was poured into cold water. The crude product was filtered off, purified by column chromatography (petroleum ether : ethyl acetate, 2:1), and recrystallised from aqueous ethanol. The yields and melting points are given in Table 1, and the IR and NMR spectroscopic data in Table 2.

Synthesis of 2-alkylsulfanyl-6-hexylsulfanylpyridine-4-carboxamides 9 -- 13

To a stirred solution of 2-chloro-6-hexylsulfanylpyridine-4-carboxamide (**3**) (10 mmol) and the appropriate thiol (10 mmol) in anhydrous *N*,*N*-dimethylformamide (10 ml) was added a sodium methoxide solution (10 mmol) dropwise. The reaction mixture was heated to about 50 °C, stirred and maintained at this temperature until TLC indicated a complete reaction. TLC was performed using petroleum ether : ethyl acetate (2:1) as the mobile phase. The mixture was poured into cold water. The crude product was filtered off, purified by column chromatography (petroleum ether : ethyl acetate, 2:1) and recrystallised from aqueous ethanol. The yields and melting points are given in Table 1, and the IR and NMR spectroscopic data in Table 2.

Synthesis of 2,6-disubstituted pyridine-4-carboxthioamides 14 -- 26

To a solution of 2,6-disubstituted pyridine-4-carboxamide (10 mmol) in anhydrous toluene (10 ml) was added Lawesson's reagent (5 mmol) and the reaction mixture was heated at reflux until TLC indicated a complete reaction. TLC was performed using petroleum ether : ethyl acetate (4:1) as the mobile phase. The mixture was then evaporated under reduced pressure, the crude product was purified by column chromatography (petroleum ether : ethyl acetate, 4:1), and recrystallised from aqueous ethanol. The melting points and yields are given in Table 1, and the IR and NMR spectroscopic data in Table 2.

Biological assays

Antimycobacterial evaluation was carried out on a semisynthetic liquid protein-containing Sula medium (IMUNA, Sarisske Michalany) buffered to pH 7.2. The following mycobacterial strains were used: *Mycobacterium tuberculosis* H₃₇Rv, *M. kansasii* PKG8, *M. avium* 80/72 and *M. fortuitum* 1021. The

concentrations of the compounds in the medium were 1000, 500, 250, 125, 60 and 30 µmol dm⁻³. The MIC values were determined after 14 days of incubation at 37 °C.

The oxygen evolution rate (OER) in spinach chloroplasts was determined spectrophotometrically (Specord UV VIS Zeiss Jena, Germany) by the Hill reaction. The measurements were carried out in phosphate buffer (20 mmol, pH = 7.2) containing sucrose (0.4 mol dm⁻³), MgCl₂ (5 mmol dm⁻³) and NaCl (15 mmol dm⁻³) using 2,6-dichlorophenol-indophenol as electron acceptor. Chlorophyll content in the samples was 30 µg dm⁻³ and the samples were irradiated (~ 100 W m⁻²) from 10cm distance with a halogen lamp (250 W) using a water filter to prevent warming of the samples (suspension temperature 22 °C). The compounds were dissolved in dimethyl sulfoxide (DMSO) because of their limited water solubility. The applied DMSO concentration (up to 5 %) did not affect OER.

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Comments

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