



Proceedings

# **Evaluation of Organophosphate Pesticide Residues in** Food using the Partial Least Squares Method <sup>†</sup>

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Abstract: Organophosphourus (OP) chemicals were broadly used as insecticides and in the treatment of human diseases such as malaria mosquitoes, parasitosis, myasthenia, and glaucoma. The OP toxicity is well known, they can cause environmental and health problems and the possibility to accumulate in the food chain. The acceptable daily intake (ADI) can be considered as a measure of the effect of pesticide residues in food on human health. In this paper, the partial least squares (PLS) approach is used to evaluate the ADIs (expressed as pADIs) of a series of 46 structurally diverse OPs. OP structures were pre-optimized using the MMFF94s force field and structural descriptors were calculated for the minimum energy conformers. This dataset was divided into 26 training compounds, and 20 pesticides were included in the prediction set. Several criteria to check the model robustness, overfitting, and the potential outliers in the X and Y space were employed. The PLS results indicate that new experimental toxicological data would be needed for five out of the 46 OPs, to improve their known ADI values, for qualitative and quantitative dietary long-term risk assessments.

Keywords: organophosphourus pesticides; PLS; Omega; ADI; risk assessment

## 1. Introduction

Pesticides are generally used to prevent and control insects, pests, and diseases in the field crops, as animal and bird repellents, food storage protectants, mould-killing substances, antifouling products, soil sterilants, and wood preservatives [1,2]. Initially, the main use of pesticides was to diminish the pest attack. Simultaneously, increased use of chemical pesticides has resulted in pollution of the environment and also caused many long-term changes in society. Pesticides are necessary to the farmer in his fight against plant pests and diseases. Today, it is anticipated that as much as 45% of the world's crop is damaged by plant pests and diseases. Thus, it is important to employ pesticides to protect the crops, both during growth and their later storage and transport. But the arbitrary and incautious use of pesticides generated extensive contamination in the food chain.

The organophosphorus pesticides (OPs) were introduced as replacements for the organochlorine pesticides after the tendency of DDT and its metabolites to bioaccumulate in ecosystems and to cause adverse health effects, particularly in top predators, led to the legal forbid or restraint of their use in the 1970s [3]. As a result of the increased use of OPs, even though originally they were considered to be less dangerous to the environment due to their low persistence, different ecotoxicological problems appeared related to their high acute toxicity. The

unreasonable use of organophosphate pesticides can generate environmental pollution problems due to their stability, high toxicity, and capacity to accumulate in the food chain [4].

The organophosphorus insecticides have a common mechanism the inhibition of acetylcholinesterase enzyme. Their relative potential toxicity in humans, rodents, and insects differ in their biotransformation and accumulation among these species [5]. The binding of OPs to carboxylesterases, cholinesterases, and other targets, which have been identified as receptors and enzymes involved in the hydrolysis of endobiotics, plays a key role in limiting the binding of OP compounds to acetylcholinesterase (AChE) [6]. Phosphorylation of AChE, which hydrolyzes acetylcholine and thus finishes its neurotransmitter activity, is the principal mechanism of OP toxicity in mammals, insects, and nematodes, with 70% to 90% inhibition usually proving lethal.

The risk assessment of chemicals is usually divided into similar, but separate practices, depending on whether the evaluated chemical causes cancer (is a carcinogen) or not (is non-carcinogen) [7]. The major difference in the calculations of carcinogenic and non-carcinogenic risks involves the method by which risks from low level exposures are determined (Winter, 1992). For non-carcinogenic effects, it is assumed that a toxicity threshold exists and exposures at levels below this threshold should not cause any effects. This measured quantity is identified as the no-observed-adverse-effect level (NOAEL). The existence of a NOAEL suggests that a toxicity threshold exists and this concept of a threshold provides the basis for non-carcinogenic risk assessment [8].

The Joint FAO/WHO Expert Committee on Food Additives (JECFA) proposed for the first time in 1958 the 'acceptable daily intake' (ADI) concept to assess the pesticide residue in food [9] with irrelevant modifications in 1962 [10,11], 1974 and 1987 [12]. Later hundreds of food additives and pesticide residues have been evaluated and reevaluated by these two international expert groups [13]. The ADIs, used nationally and internationally in the development of food standards, have proved adequate in allowing the careful use of agrochemicals and in protecting the health of the consumer [14].

ADI represents an estimate of the amount of a food additive, expressed on a bodyweight basis that can be ingested daily over a lifetime without significant risk to health [11]. The World Health Organization (WHO) and the United States Environmental Protection Agency (U.S. EPA) have determined an ADI for an actual risk management decision in the regulatory process of pesticide safety standards.

The determination of acceptable daily intake (ADI) for the toxicological assessment implies collecting all significant data, and to establish the no-effect level using the most sensitive indicator of the toxicity, and to apply an appropriate safety factor for man [13]. The ADI is determined based on known data at one time. Therefore it is impossible to be certain about the safety of a chemical and the ADI may be revised for the new toxicological data.

ADI (considered as health-based control) values of some pesticides were modeled previously by Kim [14] using the multiple linear regression (MLR) approach. He concluded that a robust QSAR approach would be helpful to identify significant information about the uncertainty of ADI values, as preliminary human health risk assessment for certain pesticides.

This paper presents the application of the partial least squares (PLS) method to evaluate the accessible daily intake (pADIs) values of a series of 46 diverse organophosphorous pesticides (http://www.inchem.org/pages/pims.html) based on their molecular structure. Molecular mechanics calculations based on the MMFF94s force field were employed to model the pesticide structures. Structural features were computed from the minimum energy structures and were related to the pADI values. Several criteria were checked to establish the model robustness and outliers in the X and Y space.

### 2. Methods

## 2.1. Definition of Target Property and Structural Descriptors

The pesticide residues in food expressed as the acceptable daily intake (ADI) (mg/kg bodyweight), molar converted to pADI (http://www.inchem.org/pages/pims.html) was considered as the dependent variable for 46 organophosphorus pesticides (Table 1).

The OP structures were pre-optimized using the MMFF94 molecular mechanics force field included in the Omega (Omega v.2.5.1.4, OpenEye Scientific Software, Santa Fe, NM. http://www.eyesopen.com) software [15,16]. For conformer generation, the maximum number of conformers per compound set of 400 and an RMSD value of 0.5 Å were used during the conformer ensemble generation.

The conformers of minimum energy were further employed to derive the structural parameters, using the DRAGON (Dragon Professional 5.5, 2007, Talete S.R.L., Milano, Italy) and Instant JChem (2020) version 20.15.0, Chemaxon, http://www.chemaxon.com) software.

**Table 1.** The organophosphorous pesticide structures, the experimental  $(pADI_{exp})$  pesticide residues in food, and PmodXPS+[2] values derived from the PLS model.

No	Structure		PmodXPS+[2]	No	Structure	_	PmodXPS+[2]
1*	SIIIII PIIIIIO	6.79	0.424	24	~s~s~s~	7.11	0.593
2*		7.8	0.292	25		8.54	0.657
3	s - s - o - / o -	6.96	0.729	26*	N O P S	7.22	0.183
4*		6.99	0.138	27		8.22	0.903
5*		8.84	0.084	28		7.6	0.607

6	~s~~\$	8.25	0.181	29 *		7.55	0.055
7*		7.54	0.676	30 *		8.48	0.739
8		7.51	0.962	31	Br a s	8.17	0.093
9 *,**	S OH	6.46	0.039	32		8.57	0.894
10		8.89	0.129	33 *		8.85	0.368
11		8.18	0.245	34		7.77	0.658
12		7.74	0.519	35		7.12	0.999
13		7.36	0.139	36		8.03	0.853
14 *,**	HI	8.96	0.016	37		9.11	0.764

15 \*

0.561

0.625

0.808

8.28

7.2

8.91

7.99

8.78

7.51

7.74

8.78

0.400

0.981

18

0.162

0.001

7.01

7.91

8.5

0.112

0.006

0.409

0.122

19 \*,\*\*

20

0.882

0.996

21

22

9.01

45

0.000

\* Test compounds included in the PLS model. \*\* Outliers in the X space.

## 2.2. Partial Least Squares (PLS) Method

The Partial Least Squares (PLS) approach [17] was employed to relate the pADI values to the calculated OP structural descriptors, using the SIMCA (SIMCA P+12 12.0.0.0 2008, Umetrics, Sweeden, www.umetrics.com) program. Stable, correct, and highly predictive models can be obtained by the PLS approach. The model quality was verified using the squared correlation regression coefficient R²(CUM), and the squared cross-validated correlation coefficient, Q²(CUM). The Variables Importance in the Projection (VIP) values and the sign of the variables' coefficients were used to explain the descriptor influence on the pADIs. The leave-7-out crossvalidation procedure was employed to select the most significant principal components and to check the internal model validation.

The Y-randomization test was employed to test the model robustness and overfitting. In this procedure, the Y-variable is randomly shuffled using the same structural descriptors. The obtained PLS models (after 999 randomizations) must have minimal  $r^2$  and  $q^2$  values [18].

Several criteria to check the potential outliers in the X and Y space were employed in the training and prediction sets: the score scatter plot, at the significance level of 0.05, the distance to the model in X space (for the selected dimension), for the observations used to fit the model (DmodX, with a significance level of 0.05), the probability of belonging to the model in the X space, for new observations in the prediction set combined with Hotelling's T<sup>2</sup> when the latter is outside the critical limit (PmodXPS+), The Hotelling's T<sup>2</sup> Range plot (which displays the distance from the origin in the score space for each selected observation, with a significance limit of 0.01).

#### 3. Results and Discussion

The X matrix of OP descriptors was analyzed using the PCA approach. A model with 6 significant components (N = 46 and X = 1733) was obtained; the first three components explain 51.5% of the information content.

The following PLS statistical results:  $R^2x(CUM) = 0.21$ ,  $R^2y(CUM) = 0.365$  and  $Q^2(CUM) = 0.141$  were obtained for one principal component for the entire set of compounds. They demonstrated the model low fitting results ( $R^2_{X(CUM)}$  and  $R^2_{Y(CUM)}$  are the cumulative sum of squares of all the X and Y values). Therefore the noise variables (e.g., coefficient values insignificantly different from 0) were excluded from this model.

The dataset was divided randomly into training and validation (43% of the total number of compounds) sets. Following compounds: 1, 2, 4, 5, 7, 9, 14, 15, 19, 23, 26, 29, 30, 33, 38, 39, 43, 46, 42, and 45 were included in the prediction set (Table 1).

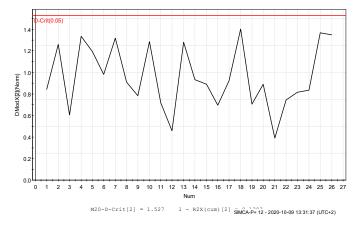
A robust and stable model with two significant principal components, which explains 88% of the information content of the descriptor matrix (for 16 structural descriptors), with R2Y(CUM) = 0.81 and Q2(CUM) = 0.77 was obtained. The descriptor coefficients and the VIP values included in the final PLS model are presented in Table 2.

The normal distribution pattern of descriptors [19] of the training and prediction sets was checked with a probability of 90% to find the X-outliers (for the training set) and the prediction compounds residing outside the AD, using the descriptor pool of the training and prediction set (included in the best PLS model). According to this criterion, compound **25** was found as a potential outlier for the training set. This assumption was not confirmed by the PModXPS+ criterion (Table 3), according to which compounds: 9, 14, 19, 42, and 45 do not belong to the prediction X space.

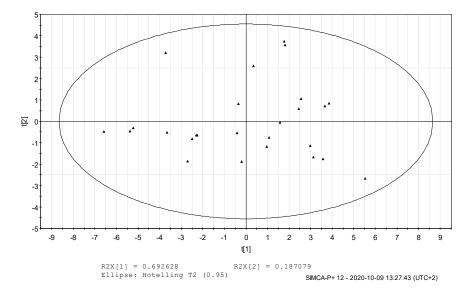
No	Variable ID*	CoefCS[2]	VIP[2]
1	CIC0	0.052	0.976
2	GATS1m	0.133	0.860
3	MATS2e	0.131	0.872
4	Mor04p	0.125	0.818
5	Mor19e	0.123	1.205
6	Mor19m	0.139	1.057
7	Mor19p	0.101	1.139
8	Mor19u	0.107	1.156
9	Mor19v	0.089	1.093
10	nΗ	0.036	0.961
11	R2u	0.075	0.985
12	RDF010e	0.031	0.951
13	RDF010m	0.031	0.953
14	RDF010p	0.036	0.963
15	RDF010u	0.033	0.957
16	RDF010v	0.036	0.963

\* CIC0-complementary information content (neighborhood symmetry of 0-order) (topological descriptors), GATS1m-Geary autocorrelation-lag 1/weighted by atomic masses (2D autocorrelations), MATS2e-Moran autocorrelation-lag 2/weighted by atomic Sanderson electronegativities (2D autocorrelations), Mor04p-3D-MoRSE-signal 04/weighted by atomic polarizabilities (3D-MoRSE descriptors), Mor19e-D-MoRSE-signal 19/weighted by atomic Sanderson electronegativities (3D-MoRSE descriptors), Mor19m-3D-MoRSE-signal 19/weighted by atomic masses (3D-MoRSE descriptors), Mor19p-3D-MoRSE-signal 19/weighted by atomic polarizabilities (3D-MoRSE descriptors), Mor19u-3D-MoRSE-signal 19/unweighted (3D-MoRSE descriptors), Mor19v-3D-MoRSE-signal 19/weighted by atomic van der Waals volumes (3D-MoRSE descriptors), nH-number of Hydrogen atoms (constitutional descriptors), R2u-R autocorrelation of lag 2/unweighted (GETAWAY descriptors), RDF010e-Radial Distribution descriptors), Function—1.0/weighted by atomic Sanderson electronegativities (RDF RDF010m—Radial Distribution Function—1.0/weighted by atomic masses (RDF descriptors), RDF010p-Radial Distribution Function-1.0/weighted by atomic polarizabilities (RDF descriptors), RDF010u-Radial Distribution Function-1.0/unweighted (RDF descriptors), RDF010v-Radial Distribution Function—1.0/weighted by atomic van der Waals volumes (RDF descriptors).

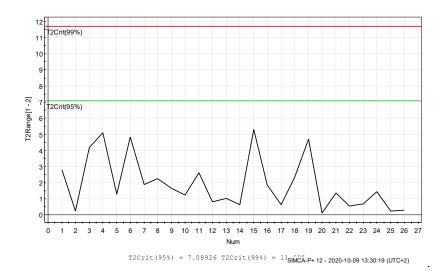
The distance to the X model plot is presented in Figure 1, the score scatter plot for the best PLS model in Figure 2 and The Hotelling's T2 range plot of the best PLS model is presented in Figure 3.



**Figure 1.** DmodX plot of the final PLS model.



**Figure 2.** Score scatter plot of the final PLS model.



**Figure 3.** The Hotelling's T2 range plot of the final PLS model.

The Hotelling's T2 range plot confirms the absence of leverage compounds and outliers. The coefficient and VIP plots are presented in Figures 4 and 5, respectively.

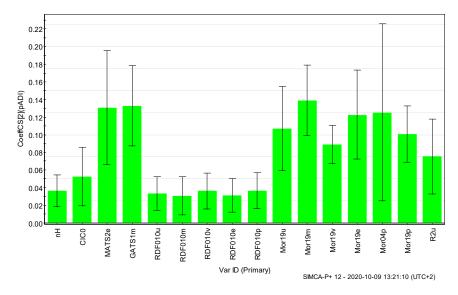


Figure 4. The coefficient plot of the final PLS model.

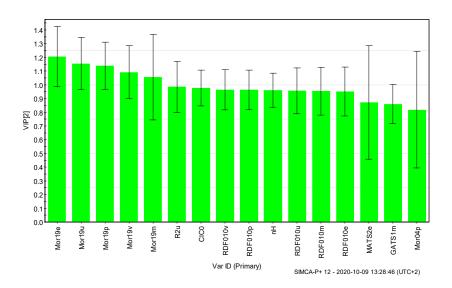


Figure 5. VIP plot for the final PLS model.

In the y-scrambling test performed for the PLS model, a significant low scrambled  $r^2$  ( $r_{\rm scr}^2$ ) and cross-validated  $q^2$  ( $q_{\rm scr}^2$ ) values were obtained for 999 trials. Figure 6 shows that in the case of all the randomized models, the values of  $r_{\rm scr}^2$  and  $q_{\rm scr}^2$  for the PLS model were < 0.5 ( $r_{\rm scr}^2/q_{\rm scr}^2$  of 0.113/–0.428). The low calculated  $r_{\rm scr}^2$  and  $q_{\rm scr}^2$  values indicate no chance correlation for the PLS chosen model.

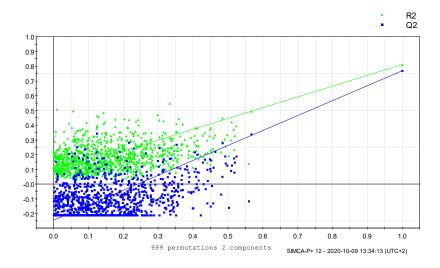


Figure 6. Y-scramble plots for the PLS model.

The experimental versus calculated pADIs plot is presented in Figure 7.

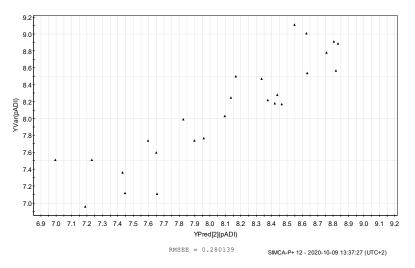


Figure 7. Experimental versus calculated pADIs plot for the final PLS model.

The final PLS model is robust and has good fitting results. All the criteria of this model used to check the presence of outliers in the X and Y space indicate that compounds: 9, 14, 19, 42, and 45 do not belong to the prediction X space. For these compounds, new experimental toxicological data would be needed, to revise their known ADI values, for qualitative and quantitative dietary long-term risk assessments.

### 4. Conclusions

The acceptable daily intake (ADI), considered to be a measure of qualitative and quantitative dietary long-term risk assessments, was modeled for a series of 46 organophosphourus (OP) pesticides using the partial least squares approach. Molecular mechanics calculations using the MMFF94s force field gave pesticide conformer ensembles. The calculated descriptors of the resulted structures of minimum energy were related to the pADIs using the PLS method. Several criteria to verify the model stability and the potential outliers in the X and Y space were applied to establish if

new experimental toxicological data would be needed for this dataset. Five OPs were found as potential outliers in the X and Y space and new ADIs would be needed to be established for these compounds.

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**Author Contributions:** G.I. analyzed the data; S.F.T. performed molecular modeling calculations, the statistical analysis, and wrote the paper.

Conflicts of Interest: The authors declare no conflict of interest.

#### References

- 1. Renwick, A.G. Pesticide residue analysis and its relationship to hazard characterisation (ADI/ARfD) and intake estimations (NEDI/NESTI). *Pest. Manag. Sci.* **2002**, *58*, 1073–1082.
- 2. Bhanti, M.; Taneja, A. Contamination of vegetables of different seasons with organophosphorous pesticides and related health risk assessment in northern India. *Chemosphere* **2007**, *69*, *63*–68.
- Galloway, T.; Handy, R. Immunotoxicity of organophosphorous pesticides. *Ecotoxicology*. 2003, 12, 345–363.
- Guodong, D.; Pei, W.; Ying, T.; Jun, Z.; Yu, G.; Xiaojin, W.; Rong, S.; Guoquan, W.; Xiaoming, S. Organophosphate pesticide exposure and neurodevelopment in young Shanghai children. *Environ. Sci. Technol.* 2012, 46, 2911–2917.
- Casarett and Doull's Toxicology. The Basic Science of Poisons, 9th ed.; Klaassen, C.D., Amdur, M.O., Doull, J., Eds.; McGraw-Hill Education: New York, NY, USA; Chicago, IL, USA; San Francisco, CA, USA.; Athens, Greece; London, UK; Madrid, Spanish; Mexico City, Mexico; Milan, Italy; New Delhi, Idian; Singapore; Sydney, Australia; Toronto, Japan, 2019.
- 6. Casida, J.E.; Quistad, G.B. Serine hydrolase targets of organophosphorus toxicants. *Chem. Biol. Interact.* **2005**, *157*, 277–283.
- Chun, O.K.; Kang, H.G. Estimation of risks of pesticide exposure, by food intake, to Koreans. Food Chem. Toxicol. 2003, 41, 1063–1076.
- Joint FAO/WHO Expert Consultation on the Application of Risk Analysis to Food Standards Issues (1995: Geneva, Switzerland), World Health Organization. Food Safety Team & Food and Agriculture Organization of the United Nations. Application of Risk Analysis to Food Standards Issues: Report of the Joint FAO/WHO Expert Consultation, Geneva, Switzerland, 13–17 March 1995. World Health Organization. Available online: https://apps.who.int/iris/handle/10665/58913.
- 9. Joint FAO/WHO Expert Committee on Food Additives. World Health Organization & Food and Agriculture Organization of the United Nations: Procedures for the Testing of Intentional Food Additives to Establish Their Safety for Use: Second Report of the Joint FAO/WHO Expert Committee on Food Additives [Meeting Held in Geneva from 17 to 24 June 1957]. WHO Techn. Rep. Ser., No. 144, FAO Nutrition Meetings. Report Series, No. 17. 1958. Available online: https://apps.who.int/iris/handle/10665/40403.
- 10. WHO Expert Committee on Pesticide Residues. World Health Organization & Food and Agriculture Organization of the United Nations: Principles Governing Consumer Safety in Relation to Pesticide Residues: Report of a Meeting of a WHO Expert Committee on Pesticide Residues Held Jointly with the FAO Panel of Experts on the Use of Pesticides in Agriculture [Meeting held in Rome from 9 to 16 October 1961]. WHO Tech. Rep. Ser., No. 240. 1962. Available online: https://apps.who.int/iris/handle/10665/40536.
- 11. Joint FAO/WHO Expert Committee on Food Additives; World Health Organization & Food and Agriculture Organization of the United Nations. Evaluation of the Toxicity of a Number of Antimicrobials and Antioxidants: Sixth Report of the Joint FAO/WHO Expert Committee on Food Additives [Meeting Held in Geneva from 5 to 12 June 1961]. WHO Tech. Rep. Ser. No. 228. FAO Nutrition Meetings. Report Series No. 31. 1962. Available online: https://apps.who.int/iris/handle/10665/40518.
- 12. WHO International Programme on Chemical Safety; Joint FAO/WHO Expert Committee on Food Additives; WHO Task Group on Updating the Principles for the Safety Assessment of Food Additives and Contaminants in Food; International Labour Organization. Principles for the Safety Assessment of Food

Additives and Contaminants in Food, published under the joint sponsorship of the United Nations Environment Programme, the International Labour Organisation, and the World Health Organization in Collaboration with the Food and Agriculture Organization of the United Nations. WHO, Environmental Health Criteria No. 70. 1987. Available online: https://apps.who.int/iris/handle/10665/37578.

- 13. Lu, F.C. Acceptable daily intake: Inception, evolution, and application. *Regul. Toxicol. Pharmacol.* **1988**, *8*, 45–60.
- 14. Rubery, E.D.; Barlow, S.M.; Steadman, J. H. Criteria for setting quantitative estimates of acceptable intakes of chemicals in food in the U.K. *Food Addit. Contam.* **1990**, *7*, 287–302.
- 15. Hawkins, P.C.D.; Skillman, A.G.; Warren, G.L.; Ellingson, B.A.; Stahl, M.T. Conformer generation with OMEGA: Algorithm and validation using high quality structures from the Protein Databank and Cambridge Structural Database. *J. Chem. Inf. Model.* **2010**, *50*, 572–584.
- Hawkins, P.C.D.; Nicholls, A. Conformer generation with OMEGA: Learning from the data set and the analysis of failures. J. Chem. Inf. Model. 2012, 52, 2919–2936.
- 17. Wold, H.; Kotz, S.; Johnson, N.L. (Eds.) *Encyclopedia of Statistical Sciences*; Wiley: New York, NY, USA, 1985; Volume 6, p. 581.
- 18. Roy, P.P.; Paul, S.; Mitra, I.; Roy, K. On two novel parameters for validation of predictive QSAR models. *Molecules* **2009**, *14*, 1660–1701.
- 19. Roy, K.; Kar, S.; Ambure, P. On a simple approach for determining applicability domain of QSAR models. *Chemometr. Intell. Lab. Syst.* **2015**, 145, 22–29.

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