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Effects of some new antioxidants on apoptosis and ROS production in AFB1 treated chickens

Consiglia Longobardi [1*], Emanuela Andretta [2], Vincenzo Romano[2], Chiara Lauritano [3], Giuseppina Avantaggiato [4], Achille Schiavone [5], Watanya Jarriyawattanachaikul [5], Salvatore Florio [2], Roberto Ciarcia [2] and Sara Damiano [2].

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[1] Department of Mental, Physical Health and Preventive Medicine, University of Campania "Luigi Vanvitelli", Largo Madonna delle Grazie n.1, 80138 Naples, Italy;consiglia.longobardi@unicampania.it (C.L.1).
[2] Department of Veterinary Medicine and Animal Productions, University of Naples "Federico II", Via Federico Delpino n.1, 80137 Naples, Italy; emauelaandretta94@gmail.com (E.A.); romano.vincenzo.594@gmail.com (V.R.); florio@unina.it (S.F.);rciarcia@unina.it (R.C.);sara.damiano@unina.it (S.D.)
[3] Marine Biotechnology Department, Zoological Station Anton Dohrn, Villa Comunale, 80121 Naples, Italy;chiara.lauritano@szn.it (C.L.2).
[4] Institute of Sciences of Food Production (ISPA), National Research Council (CNR), Via Amendola, 70126 Bari, Italy; giuseppina.avantaggiato@ispa.cnr.it (G.A.)
[5] Department of Veterinary Sciences, University of Torino, Largo Braccini n. 2, 10095 Grugliasco, Turin, Italy; achille.schiavone@unito.it (A.S.); watanya.jarriyawattanachaikul@unito.it (W.J.).
* Correspondence:consiglia.longobardi@unicampania.it; Tel.: +39-331-5346768

Abstract: Aflatoxin B1 (AFB1), the mainly *Aspergillus* fungi derived mycotoxin, is well known for its carcinogenic effects on liver and frequently occurs in food supplies, leading to fatal consequences in both farm animals and humans. Poultry, one of the most important segment of agro-industry, has demonstrated to be extremely sensitive to AFB1 intake, which results in chickens' low performance, decreased quality of both eggs and meat and a negative economic feedback. Oxidative stress caused by AFB1 plays a crucial role in chickens' kidney damage by generating lipid peroxidation accompanied by a concomitant increase in the antioxidant enzymes involved in ROS metabolism [NADPH oxidase isoform 4 (NOX4) and its regulatory subunit p47-phox]. The aim of the present work was to investigate the benefits of dietary supplementation, in chickens affected by AFB1 mycotoxicosis, using a new Feed additive (FA) containing a mixture of a tri-octahedral Na-smectite with a ligno-cellulose based materialas an antioxidant adjuvant. Exposure of AFB1 treated chickens with the feed additive induced a significant down-regulation of both NOX4 and p47-phox genes expression levels. This trend was confirmed by their protein expression, demonstrating the great potential of the FA to counteract oxidative stress. To conclude, these results could open new perspectives in the way to feed chickens using eco-friendly dietary supplements able to reduce AFB1-induced mycotoxicosis and to ameliorate poultry performances.

Keywords: Aflatoxin B1; chickens; kidney; ROS; oxidative stress; Feed Additive.

1. Introduction

Foodstuffs, grains and feed for animals are the ideal substrates for the growth of fungi and molds producing mycotoxins. The buildup of mycotoxins, the secondary metabolites produced

during fungal replication, causes an accumulation in these sources of nourishment, which lead to economic losses as well as to problems for livestock, poultry and human health [1].

Owing to climate changings, mycotoxigenic *Aspergillus* (A.) species have spread, putting on risk feed and food production chain [2], shifting also in Mediterranean zones because of average temperature rise, CO_2 levels and rainfall increase, promoting a worldwide contamination [3].

Aspergillus- derived mycotoxins are named Aflatoxins (AFs) andamong them AFB1, produced by A. *flavus*, is well known for its carcinogenic effects, in fact is counted in group I of human carcinogenic compounds [4] and may cause hepatotoxicity [5], kidney and heart damage [6], immunotoxicity [7] and could also lead to fatal consequences in both farm animals and humans [8].Therefore, AFs intake is legislated by European Community which has established its maximum quantity in foodstuffs, by placing the safe limit in a range between 2 μg/kg and 4 μg/kg [9].

Farm animals, especially poultry, one of the most important segment of agro-industry, has demonstrated to be extremely sensitive to AFB1 intake, withconsequences on the quality of both eggs and meat, and with impact onthe food chain andits economic side [10; 11].

AFB1 plays a crucial role in chickens kidney damage due to oxidative stress it induces in this organ [12]. So far there are many detoxification methods described in literature but none is able to completely remove mycotoxins in foodstuffs [13]. In the last few years, many studies have engaged in the search for eco-friendly dietary supplements, which could prevent or reduce the oxidative stress, e.g. supplementation of Vitamins A, E and C have showed antioxidative effects in poultry birds [14].

Oxidative stress in the kidneys is correlated to NOX4, the most aboundant NOX isoform at renal level. In fact, NOX4 has demonstrated to be the most important contributor to ROS generation in the kidney in several pathological conditions [15]. Physiologically, NOXs are implicated in homeostasis because of their antioxidant defence, but in pathologicalstates, their levels increase, inducingROSaccumulation [16].For this reason, the inhibition of NOX4, together with its p47-phox subunit, could lead to a promising new nutraceutical strategy in feeding not only poultry, but also other farm animals [17].

In the present work we investigate the benefits of a supplementation dietary with a Feed additive (FA)in chickens affected by AFB1 mycotoxicosis. In particular, we evaluate the role of this additive as antioxidant binder against kidney oxidative stress that affects kidneys of chickens poisoned by AFB1.

2. Experiments

2.1 Ethics Statement

The use and care of animals in this work was approved by the Bioethic Commettee of the University of Turin (Italy) (Approval number: 319508/2017-PR).

2.2 Animals and diet

Twenty-fourfemale broilers(ROSS 308) chickens 21-days-oldand 860.25 ±25.2 g of weight were housed in a cage (according to Directive 2007/43) and received a standard basal diet (190-210 g/kg of crude protein; 12,6 – 13,6 MJ/kg of Metabolizable Energy; Aviagen) *ad libitum*. After 4 days of adaptation period, they were randomly divided into 4 experimental groups: CONTROL group (*n*=6, basal diet); AFB1 group (*n*=6, AFB1 = 0,02 mg/kg feed)); FA group (*n*=6, FA = 5g/kg feed) and AFB1 plus FA group (*n*=6, AFB1 = 0,02 mg/kg feed, FA = 5g/kg feed).The treatment lasted from 25 to 35 days of age, after which animals were sacrificed and the kidneys removed to perform the following experiments.

2.3Reverse Transcription-Quantitative Polymerase Chain Reaction (RT-qPCR)

2.3.1 RNA extraction and complementary DNA (cDNA) synthesis

Three replicate chicken kidney tissues for each animal group (CONTROL, AFB1, FA, AFB1 + FA) were used for RNA extraction. Tissues were homogenized in 0,5 mL of TRIZOL Reagent (Invitrogen, Thermo Fisher Scientific) using the Tissue lyser (MM300, Retsch, Conquer Scientific) and Tungsten Carbide Beads (3 mm) (Qiagen) for 5 min at 20.1 Hz until all samples were completely homogenized (as in Lauritano et al., 2013). After centrifuging at 12.000 rpm for 10 min at 4 °C to remove debris, the supernatant was passed about 5 times through a 0.1 mm syringe-needle (as in Asai et al., 2014). Total RNA was extracted by following Trizol manufacturer's protocol and treated with DNase I (Merck KGaA, Darmstadt, Germany). RNA quantity was assessed by Nano-Drop (ND-1000 UV–Vis spectrophotometer; NanoDrop Technologies) monitoring the absorbance at 260 nm, while purity by monitoring the 260/280 nm and 260/230 nm ratios (Both ratios were approximately 2.0). For RT-qPCR, 1 μg for each sample was retrotranscribed into complementary DNA (cDNA) with the iScript™ cDNA Synthesis Kit (BIORAD, Hercules, CA, USA), following the manufacturer's instructions using the GeneAmp PCR System 9700 (Perkin Elmer, Waltham, MA, USA).

2.3.2Selection of gene of interest and RT-qPCR

Five genes of interest (GOI) were selected: the anti-apoptotic protein BCL-2, NOX4 and regulatory subunitp47-phox. 18S was used as reference gene.In order to analyse the selected GOI, the primers in Table 2 were used (Table 1).

Table 1. Gene names, primer forward (F) and reverse (R), amplicon size, oligo efficiencies (E) and correlation factors (R^2), and GenBank accession numbers..

Gene name	Primer F Primer R	Amplicon size	E	R^2	Acc. Number
NOX4	TCGGGTGGCTTGTTGAAGTA-GTCTGTGGGAAATGAGCTTGG	224	90	0.99	NM_053524
p47-phox	TACGCTGCTGTTGAAGAGGA-GATGTCCCCTTTCCTGACCA	105	100	0.99	AY029167.1
BCL-2	GCCTTCTTTGAGTTCGGTGG-CTGAGCAGCGTCTTCAGAGA	221	100	0.99	L14680.1
18S	AGAAACGGCTACCACATCCA-CCCTCCAATGGATCCTCGTT	158	93	0.99	NR_046237.1

RT-qPCR experiments were carried out in a Viia7 real-time PCR system (Applied Biosystem, Thermo Fisher Scientific, Waltham, MA, USA). PCR reaction total volume was 10 μL, including 5 μL of Fast Start SYBR Green Master Mix (Roche, Basilea, Switzerland), 0.7 pmol/μL for each oligo, and 1 μL of the cDNA template (dilution of 1:10). The thermal profile used was: 95 °C for 10 min, 40 cycles of 95 °C for 1 s, and 60 °C for 20 s. To normalize GOI expression levels, 18S was used as reference gene. The Excel-applet qGene from Muller et al., 2002 was used for the expression levels analysis.

2.4 Western Blot analysis

Kidney tissues of chickens were homogenized in a lysis buffer (RIPA buffer) with a protease inhibitor mix (cOmplete™, Mini, EDTA-free Protease Inhibitor Cocktail Tablets, Roche), employing Tissue Lyser system to promote lysis. In this phase, the cold chain was manteined. The BCA Protein Assay Kit (Bio-Rad, Milan, Italy) was used to measure total protein content of each sample.

NOX4, p47-phox and BCL-2 proteins expression were analysed by Western Blot assay.Mini-PROTEAN® precast gel 4-12% (Bio-Rad) and Opti-Protein XL (abm) as molecular weight marker were used. Trans-Blot® Turbo Nitrocellulose membrane (Bio-Rad) was used to transfer proteins. The membranes were probed with primary antibodies: NOX4(Rabbit monoclonal antibody, abcam, diluition 1:1000), p47-phox (Rabbit polyclonal antibody, Elabscience, diluition 1:500),BCL-2 (Rabbit polyclonal antibody, Cell Signaling, diluition 1:1000) and GADPH (Rabbit monoclonal antibody, Genetex, diluition 1:20000), as housekeeping expression proteins. Blots were incubated with HRP conjugates secondary antibodies (Santa Cruz Biotechnology), according to the species of primary antibodies and developed using ECL substrate (Immobilon, Millipore). Signal intensity was quantified by ChemiDoc™Imaging System (Bio-Rad) with the Bio-Rad Quantity One® software version 4.6.3. The results were expressed as arbitrary units.

2.5 Statistical analysis

The GraphPad Prism Version 8.00 (GraphPad Software, San Diego, CA) was used for statistical analysis. Statistically significant differences were evaluated by one-way analysis of Variance 55 (ANOVA), followed by Turkey's post-test. The experiments were performed at least in triplicate. *P<0.05; **P<0.01; ***P<0.001 was considered statistically significant.

3. Results

3.1. Gene expression results

3.1.1. NOX4,p47-phox and BCL-2 genes expression results

Expression levels of NOX4and its regulatory subunit p47-phox were investigated. Results, expressed as mean normalized expression, showed that expression levels of NOX4 and p47-phox significantly increased in AFB1 group, compared to control (*P<0.05, Figure 1a and 1b). Exposure of AFB1 group with Feed additive induces a decreased expression of NOX4 (#P<0.05) (Figure 1a) and p47-phox (*P<0.05) (Figure 1b) compared to control. Regarding genes involved in apoptosis regulation, results showed that anti-apoptotic protein BCL-2 increased in AFB1 group respect to control, but Feed additive (Figure 1c) has not been able to restore these values.

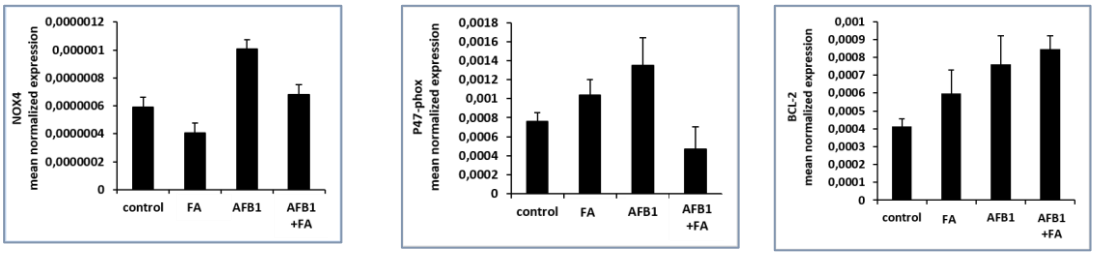

Figure 1. NOX4, p47-phox and BCL-2genesexpression in CONTROL (*n*=3), FA (*n*=3), AFB1 (*n*=3) and AFB1+ FA (*n*=5) treated groups: (**a**) mRNA levels of NOX4; (**b**) mRNA levels of p47-phox; (**c**) mRNA levels of BCL-2.Values are presented as mean normalized expression (MNE) normalized towards 18S expression (mean ± standard error).

3.2.Protein expression results

3.2.1. NOX4, p47-phox and BCL-2 proteins expression results

The trend showed in gene expression is also respected in protein expression. Western blot analysis confirmed that NOX4 (Figure 2a) and p47-phox (Figure 2b) proteins were significantly up-regulated in AFB1 respect to control animals (*P < 0.05 vs control). FAtreatment restored the NOX4 values (*P< 0.05 AFB1 vs AFB1+FA Figure 2a) and a similar trend is about p47-phox (Figure 2b). Western blot analysis for BCL-2 (Figure 2c) protein showed no significant increase in AFB1 respect to control animals (Figure 2c).

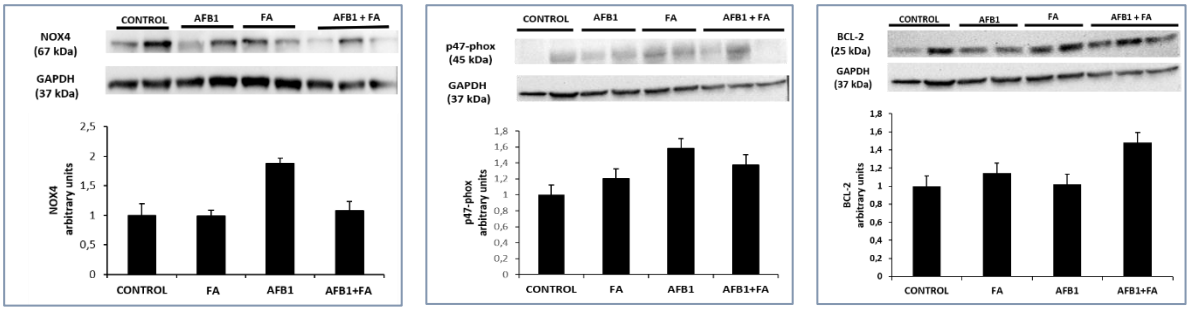

Figure 2.NOX4, p47-phox and BCL-2 proteins expression in CONTROL (*n*=3), FA (*n*=3), AFB1 (*n*=3) and AFB1+FA (*n*=5) treated groups: (**a**) protein levels of NOX4; (**b**) protein levels of p47-phox; (**c**) protein levels of BCL-2.Values are presented as arbitrary units, normalized towards GAPDH.

4. Discussion

Ubiquitary presence of mycotoxins in feed isdisadvantageous for poultry's performances,representing a critical risk for chickens farming.Nutraceuticals are progressively evaluated as valid tools in veterinary medicinebecause of their capacity to counteract the presence of mycotoxins in animals feedings [17].

The FAhas been tested and has demonstrated to bea valid binder for feed decontamination from AFB1.As a matter of fact, FAis able to down-regulate both the transcription and the expression of NOX4 (considered one of the crucial factors for oxidative stress) in chichenskidneys, together with its p47-phox subunit, suggesting its capacity to bind AFB1 according to the mechanism with which it acts, reducing bioavailability of this kind of mycotoxin.In particular, FA treatment shows an improvement of renal alterations by reverting the increased levels of ROS and activating antioxidant enzymes.

As regard anti-apoptotic action, BCL-2 is over-expressed in AFB1 plus FAtreated group, demonstrating a lack of involvement of the apoptotic process in Aflatoxicosis. This data is still incomplete because it's necessary to also investigate the role of some pro-apoptotic proteins, e.g. BAX, in order to evaluate BCL-2/BAX ratio.

Anyway, the management of chickens environmental risk by adding FAas adsorbent supplement in animal diets, could prevent the deleterious effects of poultry mycotoxicosis.

5. Conclusions

The experiments performed in this work highlight the capacity of a new Feed additive to revert nephrotoxicity induced by AFB1 in poultry.

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Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

The following abbreviations are used in this manuscript:

MDPI: Multidisciplinary Digital Publishing Institute
DOAJ: Directory of open access journals
TLA: Three letter acronym
LD: linear dichroism
AFB1: Aflatoxin B1
ROS: Reactive oxygen species
NOX4: NADPH oxidase 4
FA: Feed Additive

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