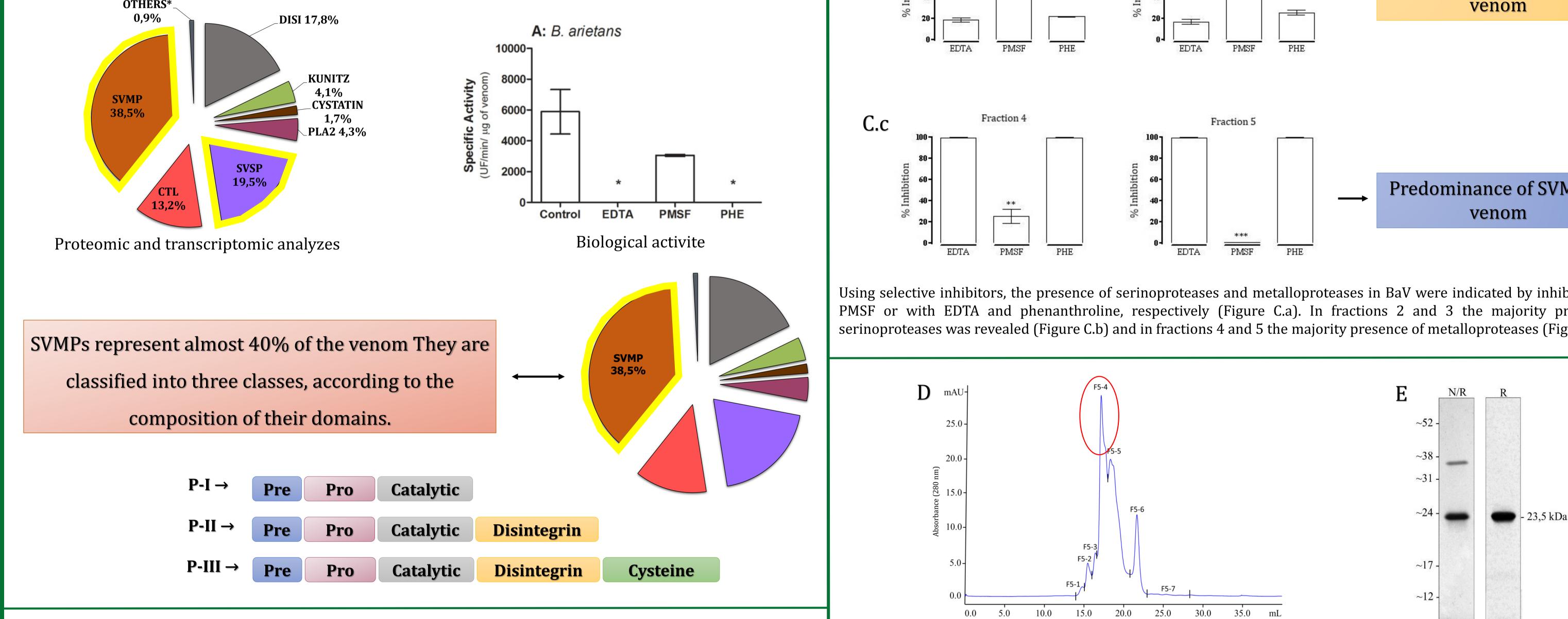
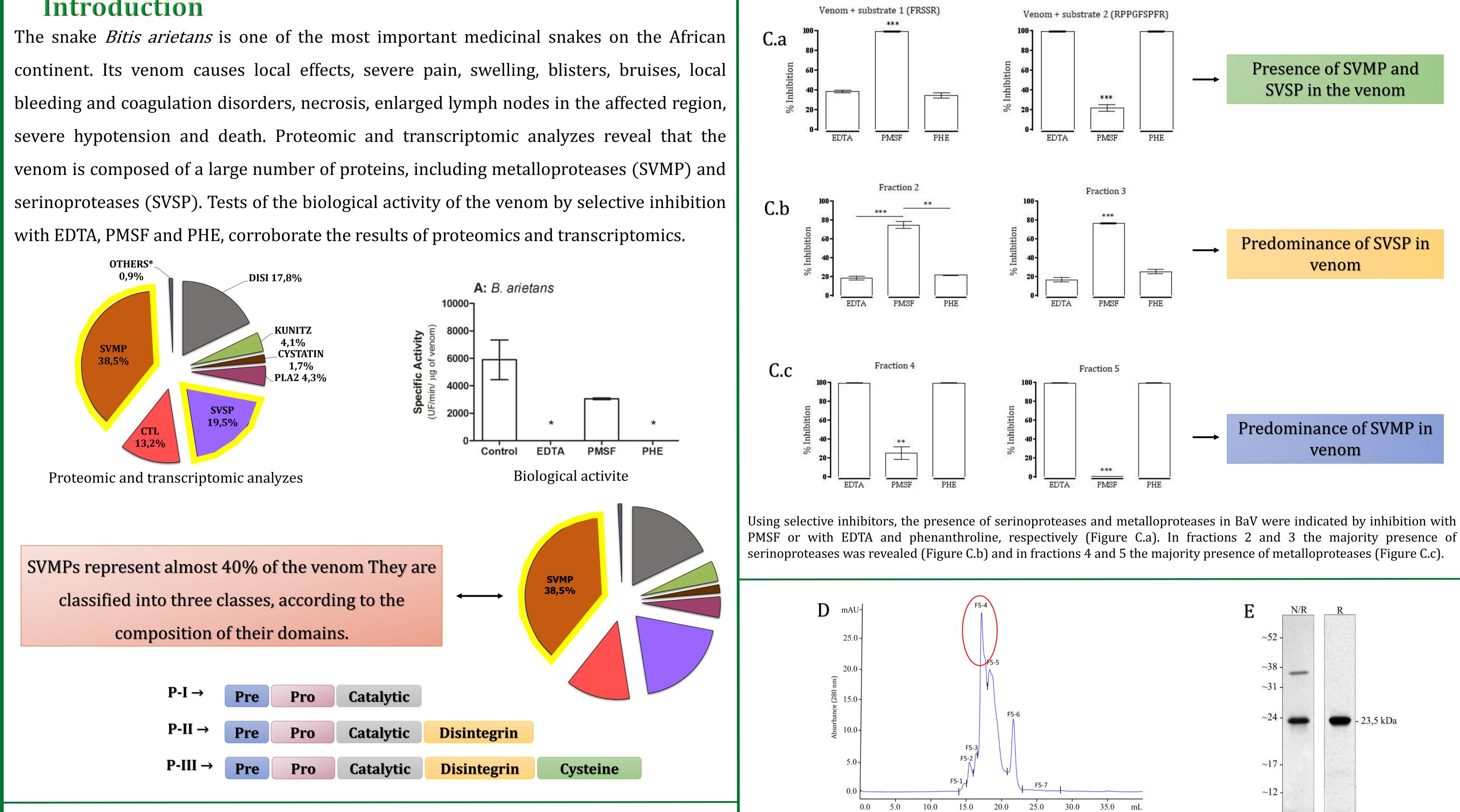


Production of Monoclonal Antibodies (mAbs) Purified Anti-metalloprotease from the Venom of the Serpent Bitis arietans

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Introduction

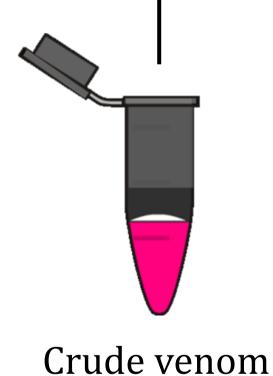




The fifth chromatographic peak (F5), obtained from the first purification step, was subjected to a second cycle of molecular exclusion chromatography on a Superdex 75 column, equilibrated and eluted with ammonium acetate (50 mM) in an airconditioned environment (22 ± 2°C) (Figure D). The fraction 5-4 (4 ug/well), corresponding to the metalloprotease isolated from fraction 5, was submitted to 12% polyacrylamide gel under reducing (R) and non-reducing (N/R) conditions (Figure E).

Methodology

1. Two steps of molecular exclusion chromatography 2. Proteolytic activity - FRET substrates **3.** Characterization of the enzymatic class - EDTA, 1.10 PHE and PMSF

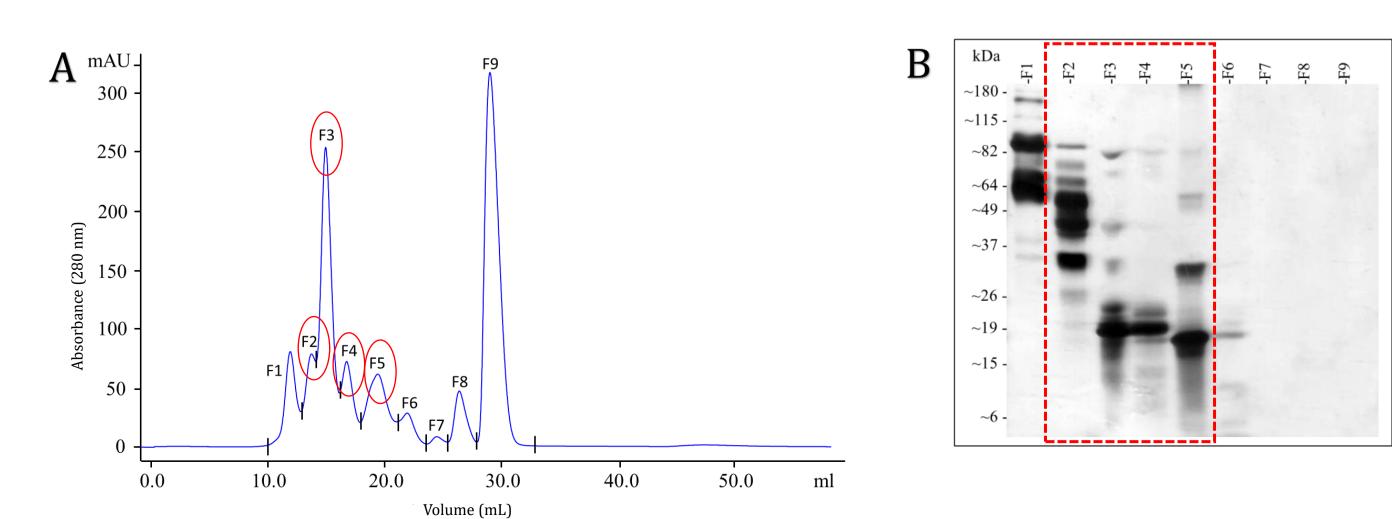


4. Mass spectrometry identification

Next steps

- Immunization
- Peritoneal cells
- Identification of anti-SVMP mAbs
- Fusion myelomas + activated B cells
- Identification and expansion
- Evaluation of antitoxic activity
- Quantification of proteins
- Titration of avidity and affinity

Results



Conclusion

- SVMPs are classified into three classes, according to the composition of their domains
- First stage of the *B. arietans* venom fractionation originated 9 subfractions where fraction 2, 3, 4 and 5 was selected
- SVMP is inhibited by EDTA and phenanthroline
- Fractions 4 and 5 have a majority presence of SVMPs

Volume (mL)

Second stage of fraction 5 molecular exclusion chromatography gave rise to 5

subfractions where the F5-4 fraction was selected to continue the experiments

References

WARRELL, D. A.; Ormerod, L. D.; Davidson, N. M. Bites by puff-adder (*Bitis arietans*) in Nigeria, and value of antivenom.Br. Med. J., v. 4, n. 5998, p. 697-700, 1975.

ANGELA, A. A. M. Doctoral Thesis. Inflammatory properties of the snake venom Bitis arietans: contribution of lipid mediators to in vivo poisoning and action of toxins isolated from the venom on human macrophages. Submitted to the Postgraduate Program in Immunology, Institute of Biomedical Sciences, University of São Paulo, 2019. Defended and approved".

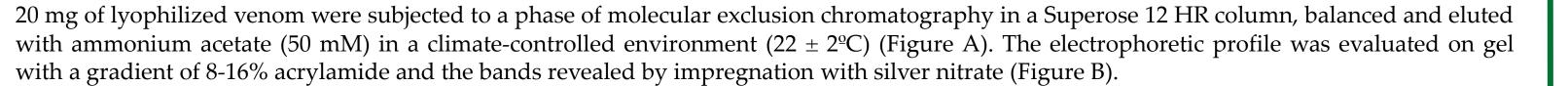
ANGELA, A. A. M.; BONETTI, M.; MONTES, A.; JANCAR, S.; PORTARO, F. C. V.; DIAS DA SILVA, W. The Inflammatory response induced by *Bitis arietans* snake venom.(Manuscript in preparation), 2019. ANGELA, A. A. M.; MAGNOLI, F. C.; KUNIYOSHI, A.K.; IWAI, L. K.; TAMBOURGI, D. V.; PORTARO, F. C. V.; DIAS

DA SILVA, W.Kn-Ba: a novel serine protease isolated from *Bitis arietans* snake venom with fibrinolytic and kinin-releasing activity. J. Venom. Anim. Toxinxs, 24:1-11. 2018.

ANGELA, A. A. M.; BONETTI, M.; MONTES, A.; JANCAR, S.; PORTARO, F. C. V.; DIAS DA SILVA, W. Purification of a metalloprotease from *Bitis arietans* Snake Venom (manuscript in preparation), 2019./

Acknowledgments







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Advisor: Wilmar Dias da Silva

