# Modifications of the PAMONO-sensor help to size and quantify low number of individual biological and non-biological nano-particles.

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#### Introduction

In the series of works, we demonstrated that plasmon assisted microscopy of nano-objects (PAMONO) technique can be successfully employed for sizing and quantification of viruses, microvesicles and charged non-biological particles. The PAMONO-sensor allows for label-free and specific detection of individual biological nano-particles. In order to achieve homogeneous illumination and enlarged focused image area on the sensor slide, the PAMONO-sensor configuration was modified. These modifications are discussed in the present work. In future, applied modifications will help to analyze samples with low concentration of target nano-particles and also help to gain biochemical information about nano-vesicles of interest.

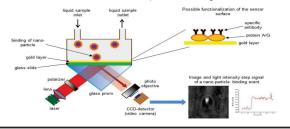
#### Aims of studies

- 1) Verify whether dependency between the value of light intensity step and size of nanoparticles remains linear for modified PAMONO-sensor.
- 2) Confirm the ability of the PAMONO-sensor to detect simultaneously 800nm and 200nm silica particles (they may serve as optical models of bacteria and viruses).
- Verify whether formation the layers of biological molecules: Cys-conjugated protein A/G and antibodies remains detectable after applied sensor modifications.
- 4) Define possible duration time for the analysis of nano-particle samples using one sensor chip without its replacement.

# Detection of single particles, not particle layers

Classical SPR: The signal appears as a result of the resonance curve shift after formation of the biomolecule layer onto sensor surface.

PAMONO-sensor: Particle binding signal appears as a bright spot on a gray backround. Secondary concentric plasmonic waves help to visualize a signal.



### Standard and modified PAMONO-sensor





Standard PAMONO-sensor

Modified PAMONO-senso

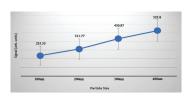
In the modified PAMONO-sensor setup, a concave lens was employed to obtain homogeneous illumination of the gold sensor chip surface. Further, the enlargement of the image focused area for the modified PAMONO - sensor was achieved via mounting of a diffraction grating.

## Results

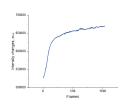
It was important to verify whether key characteristics of the PAMONO-sensor demonstrated in our previous works (Gurevich, 2011), (Shpacovitch, 2015) (Shpacovitch, 2017) remain unchanged after modifications of the instrument's optical scheme. Thus, we proved that the modified PAMONO-sensor still demonstrates linear dependency between the value of intensity step and the size of analized nanoparticles. Further, we also show that modified PAMONO-sensor retains the traits of classical SPR sensors and helps to monitor the coating of sensor surface with different proteins such as Cys-conjugated protein A/G and antibodies, for example.

Dependency between the value of intensity step and the size of nanoparticles (200nm polystyren particles were used;

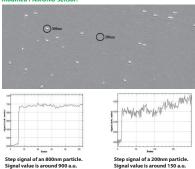
modified DAMONO correct)



2. Monitoring the adhesion of Cys-protein A/G to the gold sensor



 Simultaneous detection of 800nm and 200nm silica particles usin modified PAMONO-sensor.



Changes the number of signals appeared per recorded image during sample analysis (200nm polystyrene particles were used in experiments performed on standard and modified PAMONO-sensors).



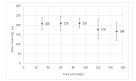


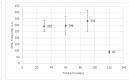
The received data were normalized according to the image area and also the concentration of the nano-particles used for analysis.

 Changes the value of intensity step signal monitored during sample analysis (200nm polystyrene particles were used in experiments performed on standard and modified PAMONO-sensors).









## Perspectives:

- . Define possible duration time for the analysis of sample containing biological nano-particles.
- Confirm the power of the modified PAMONO-sensor for sizing and quantification of nano-particles in low concentrated samples

References:

- 1) Shpacovitch V. et al., *Analytica Chimica Acta* (<u>2018</u>) 1005 pp 1-15
- 2) Shpacovitch V. et al., *Sensors* (2017) 17, 244; 10.3390/s17020244
- 3) Shpacovitch V. et al., *Analyt. Biochemistry* (<u>2015</u>) 486 pp 62-69.
- 4) Gurevich E. et al., Sensors and Actuators B (2011) 160 pp 1210-1215.





