chemistry proceedings

MDPI

Proceedings

Tropomyosin Analysis in Foods Using An Electrochemical Immunosensing Approach †

Ricarda Torre [1], Maria Freitas [1], Estefanía Costa-Rama [2], Henri P.A. Nouws [1,*] and Cristina Delerue-Matos [1]

[1] REQUIMTE/LAQV, Instituto Superior de Engenharia do Porto, Instituto Politécnico do Porto, Dr. António Bernardino de Almeida 431, 4200-072 Porto, Porto; rdvdt@isep.ipp.pt (R.T.); maria.freitas@graq.isep.ipp.pt (M.F.); cmm@isep.ipp.pt (C.D.-M.)

[2] Departamento de Química Física y Analitica, Universidad de Oviedo, Av.Julián Clavería 8, 33006 Oviedo, Spain; costaestefania@uniovi.es (E.C.-R.)

* Correspondence: han@isep.ipp.pt

† 1st International Electronic Conference on Chemical Sensors and Analytical Chemistry, online, 01-15 July 2021.

Abstract: A screen-printed carbon electrode was used as the transducer for the development of an electrochemical immunosensor for the determination of tropomyosin (a major shrimp allergen) in food samples. Monoclonal and polyclonal antibodies were used in a sandwich-type immunoassay. The analytical signal was electrochemically obtained using an alkaline phosphatase-labelled secondary antibody and a 3-indoxyl phosphate/silver nitrate substrate. The total assay time was 2h50 min and allowed the quantification of tropomyosin between 2.5 – 20 ng mL^{-1}, with a limit of detection of 1.7 ng mL^{-1} The immunosensor was successfully applied to the analysis of commercial food products.

Keywords: Seafood allergy; tropomyosin; shrimp; food allergy; screen-printed electrodes; electrochemical biosensor

Citation: Torre, R.; Freitas, M.; Costa-Rama, E; Nouws, H.P.A.; Delerue-Matos, C. Tropomyosin analysis in foods using an electrochemical immunosensing approach. *Chem. Proc.* **2021**, *3*, x. https://doi.org/10.3390/xxxxx

Published: date

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

1. Introduction

Over the past few years food allergies have increasingly been regarded as a significant worldwide public health problem. Among shellfish allergies, shrimp is the predominant crustacean causing over 80% of allergic reactions that can result in severe hypersensitivity such as urticaria, asthma and it is a major cause of anaphylaxis [1,2].

Tropomyosin (TPM), a major common allergenic protein found in seafood, is relatively resistant to peptic acidic digestion, which causes a continuous effect of the protein on the immune system. To protect the consumer for harmful allergens and potentially life-threatening reactions, food manufacturers are required to label and highlight shellfish allergenic ingredients on food packages [3].

Currently, multiple technical approaches have been developed to identify the presence of shrimp tropomyosin in food, including enzyme-linked immunosorbent assays (ELISA), DNA detection, polymerase chain reaction (PCR), microarray and qualitative/semi-quantitative lateral flow assays. Although ELISA is the most commonly used method for TPM detection and quantification, it presents some disadvantages such as the long and tedious steps in the analysis procedure, long analysis times and high costs [4,5]. An alternative way to determine TPM in foods is through the use of electrochemical immunosensors. These sensors provide highly selective, sensitive, fast and cheap analysis and are suitable for *in situ* applications. Therefore, in this work a simple voltametric immunosensor for the determination of TPM in commercial food products was developed. The immunoassay was based on a sandwich-type assay using screen-printed carbon

electrodes (SPCE) as transducer. Monoclonal and polyclonal antibodies were used to capture and detect TPM. To obtain the analytical signal an alkaline phosphatase-labelled secondary antibody and 3-indoxyl phosphate / silver nitrate (enzymatic substrate) were employed; the enzymatically deposited silver was analysed by linear sweep voltammetry.

The applicability of the immunosensor was accessed by analyzing different food samples.

2. Materials and Methods

2.1. Instrumentation

Linear sweep voltametric analyses were performed using an Autolab PGSTAT204 potentiost/galvanostat from Methrohm Autolab. Disposable screen-printed carbon electrodes (DRP-110) with a carbon working electrode (WE, d = 4 mm), a carbon counter electrode, and a silver pseudoreference electrode were purchased from Methrohm DropSens.

2.2. Reagents and Solutions

Tris(hydroxymethyl)aminomethane (Tris, ≥ 99.8%), magnesium nitrate hexahydrate ($Mg(NO_3)_2$, 99%), nitric acid (HNO_3 ≥ 65%), 3-indoxyl phosphate (3-IP, ≥ 98%), silver nitrate ($AgNO_3$, ≥ 99.9995%), β-casein from bovine milk (≥ 98%), and bovine serum albumin (BSA) were obtained from Sigma-Aldrich.

Mouse IgG_1 monoclonal antibody (capture antibody, C-Ab) purified natural shrimp Tropomyosin standard (antigen), rabbit polyclonal antiserum shrimp tropomyosin (detection antibody, D-Ab) were purchased from by Indoor Biotechnologies. An alkaline phosphatase goat anti-rabbit IgG antibody (AP-Ab) was supplied by Invitrogen. Throughout the work ultra-pure water (resistivity = 18.2 MΩ cm), obtained from a Millipore (Simplicity 185) water purification system, was used. Working solutions of BSA, the antibodies and antigen were prepared in 0.1 M Tris-HNO_3 pH 7.4 buffer (Buffer 1, B1). A second buffer (B2, 0.1 M Tris-HNO_3 pH 9.8 containing $Mg(NO_3)_2$ ($2x10^{-2}$ M)) was used to prepare the solution containing 3-IP ($1x10^{-3}$ M) and $AgNO_3$ ($4x10^{-4}$ M).

2.3. Sample Preparation

Shrimp, shrimp sauce and crab and chicken paste were used to evaluate the immunosensor's applicability to food analysis. Samples were prepared as follows: (a) 1 g of sample was mixed with 10 mL of Tris-HNO_3 (pH 8.2, 1% NaCl) at 60°C during 15 min in a water bath; (b) the resulting suspension was then centrifuged at 2500 rpm for 20 min and (c) the supernatant was divided in aliquots and stored at −20 °C until use.

2.4. Immunosensor Assay & Electrochemical Measurements

The representative scheme of the immunosensor assay and detection strategy is presented in Figure 1. The WE of the SPCE was coated with C-Ab (10 µL, 20 µg mL^{-1}) and left to incubate overnight at 4°C. After rinsing the sensor with buffer B1, surface blocking was carried out using 40 µL of a 2-% (m/V) BSA solution during 30 min. After this, the sensor was washed with buffer B1 and incubated with 40 µL of a previously mixed (10 min before use) solution containing the antigen, the detection antibody (1:2000) and BSA (1% (m/V)) during 60 min. After rinsing with buffer B1, 40 µL of an AP-Ab solution (1:40 000) was placed on the sensor for 60 min. The sensor was then rinsed with buffer B2, and the enzymatic reaction was carried out by depositing 40 µL of a mixed solution containing 3-IP and silver nitrate on the SPCE for 20 min. LSV was used to record the analytical signal (potential range: -0.03 V to +0.4 V, scan rate: 50 mV/s). All analyses were performed in triplicate and carried out at room temperature (20 ± 1°C).

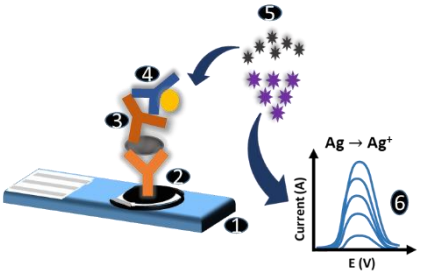

Figure 1. Schematic representation of the developed immunoassay. (1) Screen-printed carbon electrode; (2) C-Ab immobilization; (3) addition of a mixture containing standard/sample and D-Ab; (4) addition of AP-Ab; (5) addition of the enzymatic substrate (3-IP) and silver ions; and (6) voltametric detection of Ag^0.

3. Results and Discussion

3.1. Optimization Studies

The immunosensing strategy was based on a sandwich-type assay performed on bare SPCEs as transducers. In the first phase of the immunosensor development, two different surface blockers were tested: β-casein (2% (m/V)) and BSA (2% (m/V)). As can be observed in Figure 2, when BSA was used the highest peak current intensity (i_P) and signal-to-blank ratio (S/B) was obtained.

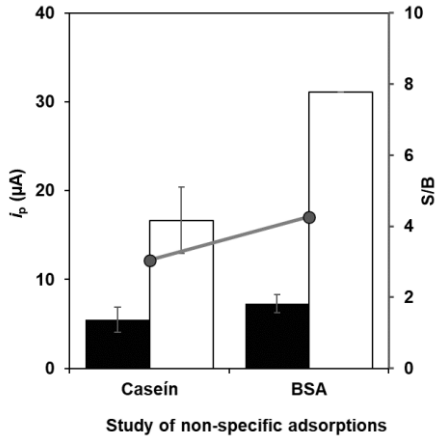

Figure 2. Peak current intensities (i_P) obtained for the study of the surface blocker (casein and BSA, both at 2% (m/V)). Black bars: blank assay. White bars: TPM (10 ng mL^{-1}). Results are presented as average ± standard deviation (n=3). Experimental conditions: C-Ab - 10 μg mL^{-1}; D-Ab - 1:250 dilution; AP-Ab - 1:20 000 dilution; 3-IP - 1.0 x 10^{-3} M; and $AgNO_3$ - 4.0 x 10^{-4} M.

In order to select the optimum concentrations of both the capture and detection antibodies, a standard solution of tropomyosin (10 ng mL^{-1}) was used. First, for fixed dilutions of D-Ab (1:250) and AP-Ab (1:20 000), different C-Ab concentrations between 2.5 and 20 μg mL^{-1} were tested. The obtained results reveal that a concentration of 20 μg mL^{-1} resulted in the highest peak current intensity and S/B ratio. After this and maintaining the AP-Ab dilution at 1:20 000, different D-Ab dilutions (between 1:250 and 1:12 000) were tested. The selected dilution was 1:2000 because the highest i_P and lowest blank signal were obtained. After selecting the C-Ab concentration (20 μg mL^{-1}) and D-Ab dilution (1:2000), different assay formats were studied in order to reduce the number of incubation steps and, subsequently, the assay time. Different steps were combined and the most adequate combination, the previous mixing of the antigen with the D-Ab, led to a 60-min reduction of the assay time. The next studies were performed to select the optimum AP-Ab dilution by testing dilutions between 1:10 000 and 1:40 000. A 1:40 000 dilution was selected because a low blank signal and the highest S/B ratio were observed. After this, the AP-Ab

incubation time was studied between 15 and 60 min, obtaining and the best results for 60 min. A summary of the optimization studies is indicated in Table 1.

Table 1. Optimization of the different experimental variables involved in the construction of the immunosensor for TPM analysis.

Variable	Studied range	Selected value
[C-Ab), µg mL^{-1}	2.5 - 20	20
[D-Ab], dilution	1:250 – 1:12 000	1:2000
[AP-Ab], dilution	1:10 000 – 1:40 000	1:40 000
AP-Ab incubation time, min	15 - 60	60

3.2. Analytical Performance

To establish the performance characteristics of the immunosensor, standard solutions with different TPM concentrations (2.5 - 50 ng mL^{-1}) were analyzed. A linear relationship was observed between 2.5 - 20 ng mL^{-1} (i_P (µA) = 0.787 [tropomyosin] (ng mL^{-1}) + 5.45, r = 0.990, n=5). Examples of voltammograms in the linear range (Figure 2(a)) and the calibration plot (Figure 2(b)) are shown in Figure 2. The limit of detection (LOD) was calculated as 3 times the standard deviation of the blank divided by the slope and the value obtained was 1.7 ng mL^{-1}. The limit of quantification (LOQ) was calculated as 10 times the standard deviation of the blank divided by the slope, obtaining a concentration of 5.7 ng mL^{-1}. The coefficient of variation of the method was < 9%.

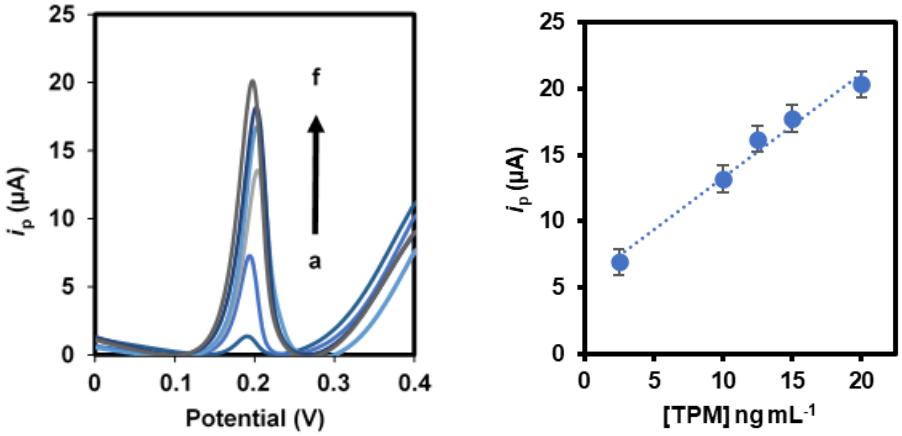

Figure 2. (**a**) Examples of voltammograms in the linear range (a - blank; b - 2.5 ng mL^{-1}; c - 10 ng mL^{-1}; d - 12.5 ng mL^{-1}; e - 15 ng mL^{-1}; and f - 20 ng mL^{-1}); (**b**) Calibration plot. Experimental conditions: C-Ab - 20 µg mL^{-1}; BSA - 2% (m/V); mixture of standard TPM solutions with D-Ab - 1:2000; AP-Ab - 1:40 000; 3-IP - 1x10^{-3} M; and $AgNO_3$ - 4x10^{-4} M.

3.3. Selectivity and Interference Studies

The selectivity of the sensor towards TPM was evaluated by analyzing other allergens such as Ara h 1 (peanut allergen, 250 ng mL^{-1}), Cyp C 1 (fish allergen, 20 ng mL^{-1}), Ovalbumin (GAL d 2, chicken egg allergen, 1% (m/V)). Besides these allergens, histamine (6.8 mg mL^{-1}), a biogenic amine and the most important fish freshness indicator, was also included in this study. The signal for all these compounds was similar to the blank signal, confirming the selectivity of the proposed sensor. Besides this, TPM was mixed with each of the compounds to evaluate their interference in the analysis. The obtained signals were nearly the same as the one obtained for a 10-ng mL^{-1} TPM solution, which indicates that the other allergens and histamine do not significantly interfere in the analysis.

3.4. Applicability to Food Analysis

The feasability of the sensor for the determination of TPM in commercial food samples was tested. Shrimp, shrimp sauce and crab paste were analysed, obtaining TPM concentrations of 80.42 ± 2.7 μg g^{-1}, 170.4 ± 1.80 ng g^{-1} and 21.6 ± 4.13 ng g^{-1}, respectively. The developed immunosensor was also used to detect the presence of TPM in chicken paste. As expected, this sample gave a negative result (no significant differences when compared with the blank signal), so the TPM concentration was below the sensor's LOD.

4. Conclusions

The current trends in analytical chemistry are focused on the development of simple and *in situ* analysis devices to ensure food safety. In this work, a simple immunosensor for tropomyosin analysis was developed. This immunoassay only takes 2h50 min, and it requires 40 μL of sample to perform the analysis. The sensor can determine tropomyosin in a concentration range between 2.5 and 20 ng mL^{-1} and a limit of detection of 1.7 ng mL^{-1} was achieved. The developed methodology fulfills the requirements of (bio)sensors construction such as small size and the use of low amounts of reagents and samples. Moreover, it allows the possibility of decentralized analysis, which could be useful for the control of tropomyosin, avoiding cases of food allergy.

Author Contributions: Conceptualization, M.F. and H.N.; methodology, M.F. and H.N.; validation, all authors; formal analysis, M.F. and R.C..; investigation, M.F. and R.C..; resources, H.N. and C.D.-M.; data curation, M.F. and R.C.; writing—original draft preparation, R.C.; writing—review and editing, M.F. and H.N..; visualization, R.C.; supervision, M.F., H.N., E.C.-R. and C.D.-M.; project administration, H.N.; funding acquisition, H.N. and C.D.-M. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Portuguese national funds (FCT/MCTES, Fundação para a Ciência e a Tecnologia and Ministério da Ciência, Tecnologia e Ensino Superior) through the project UIDB/50006/2020. This work also received financial support from the European Union (FEDER funds through COMPETE POCI-01-0145-FEDER-030735) and National Funds (FCT) through the project PTDC/QUI-QAN/30735/2017 - TracAllerSens - Electrochemical sensors for the detection and quantification of trace amounts of allergens in food products.R.Torre thanks FCT and ESF (European Social Fund) through POCH (Programa Operacional Capital Humano) for her PhD grant (SFRH/BD/143753/2019).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Reference

1. D. Jiang, J. Ji, luan, X. Sun, Y. Zhang, G. Zhang, L. Tang, Mast cell-based electrochemical biosensor for quantification of the major shrimp allergen Pen a 1 (tropomyosin), Biosens. Bioelectron. 50 (2013) 150–156. doi:10.1016/j.bios.2013.06.032.
2. A. Angulo-Ibáñez, U. Eletxigerra, X. Lasheras, S. Campuzano, S. Merino, Electrochemical tropomyosin allergen immunosensor for complex food matrix analysis, Anal. Chim. Acta. 1079 (2019) 94–102. doi:10.1016/j.aca.2019.06.030.
3. Y. Wang, Z. Rao, J. Zhou, L. Zheng, L. Fu, A chiral assembly of gold nanoparticle trimer-based biosensors for ultrasensitive detection of the major allergen tropomyosin in shellfish, Biosens. Bioelectron. 132 (2019) 84–89. doi:10.1016/j.bios.2019.02.038.
4. A. Mohamad, M. Rizwan, N.A. Keasberry, A.S. Nguyen, T.D. Lam, M.U. Ahmed, Gold-microrods/Pd-nanoparticles/polyaniline-nanocomposite-interface as a peroxidase-mimic for sensitive detection of tropomyosin, Biosens. Bioelectron. 155 (2020) 112108. doi:10.1016/j.bios.2020.112108.
5. R. Chinnappan, A.A. Rahamn, R. AlZabn, S. Kamath, A.L. Lopata, K.M. Abu-Salah, M. Zourob, Aptameric biosensor for the sensitive detection of major shrimp allergen, tropomyosin, Food Chem. 314 (2020) 126133. doi:10.1016/j.foodchem.2019.126133.