

## Our Ongoing Studies Initial Considerations

Difficult-to-treat bacterial infections involving resistant human and plant pathogens, severely afflict hospitals and concern the agri-food sector. Bacteria of the genus *Pseudomonas* can quickly become resistant to antibiotics and spread such resistance to other bacteria. Species such as *P. aeruginosa*, *P. putida* and *P. fluorescens*, which normally produce antibiotics or help reduce some forms of pollution, can trigger serious nosocomial infections in humans. *P. fragi* is a major cause of dairy and meat spoilage, while *P. syringae* can infect a wide range of economically important plant species, including tobacco, kiwi and tomato. Therefore, new strategies and antibacterial agents capable of stopping these bacteria, regardless of their resistance to antibiotics, are urgently needed to limit serious human infections, food waste, plantation extermination and economic losses.

### The Promising Candidate Molecule for Our Project

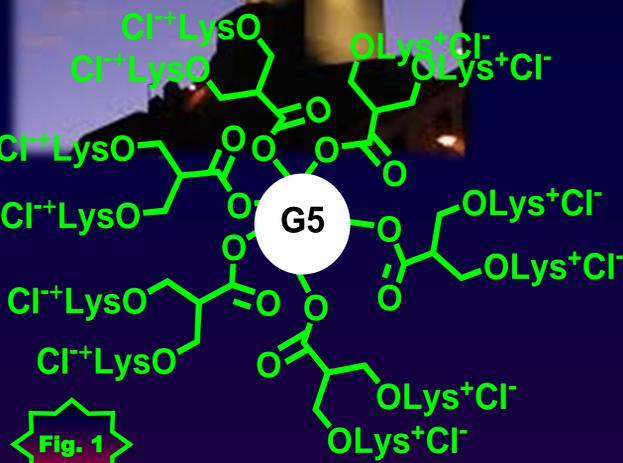


Fig. 1

Lys = lysine; G5 = generations number

We recently published the auto-biodegradability and rapid bactericidal activity of a fifth-generation dendrimer (**G5-PDK**) (Fig. 1) against *A. baumannii*, *A. johnsonii*, *A. junii*, *A. pittii* and *A. ursingii* (MIC = 3.2-12.7  $\mu$ M) [1]. Being active against different species of the genus *Acinetobacter*, we have evaluated the antibacterial effects of **G5-PDK** also against representatives of other non-fermenting Gram-negative species, such as *Pseudomonads*. Interestingly, while **G5-PDK** was practically inactive against the *P. aeruginosa* isolate tested in these first investigations (MIC  $\Rightarrow$  6.4  $\mu$ M), it showed very low MICs against other *Pseudomonas* species belonging to the *Fluorescens* group (MIC = 0.8  $\mu$ M).

Fig.1 shows the simplified structure of **G5-PDK**, which is a cationic dendrimer consisting of two uncharged fifth-generation polyester-based umbrellas, attached to a propanediol (PD) core decorated with 64 lysine hydrochloride salts, which possess a total of 128 cationic groups.

To pursue our aim, in the present study **G5-PDK** was tested on several species of the genus *Pseudomonas*. MICs for **G5-PDK** were obtained by testing a total of 48 strains, including 32 strains of *P. aeruginosa*, 5 of *P. fluorescens*, 5 of *P. putida*, 3 of *P. straminea* and 1 representative of *P. oleovorans*, *P. fragi* and *P. syringae* species. The MICs observed for **G5-PDK** against all isolates tested were reported in Table 1, which also collects the selectivity indices (SIs) of **G5-PDK** for each strain, determined using the  $LD_{50}$  of **G5-PDK** obtained from experiments of cytotoxicity performed on human keratinocytes (*Ha-CaT*) (Fig 3). Time-kill experiments were reported in Fig. 4.

Table 4. MIC values of **G5-PDK** obtained on clinical and environmental isolates of the genus *Pseudomonas* from experiments conducted in triplicate and selectivity indexes (SI) of **G5-PDK** for each strain.

| Strains                       | MIC $\mu$ M ( $\mu$ g/mL) | Selectivity Index |
|-------------------------------|---------------------------|-------------------|
| <i>P. aeruginosa</i> 1* 1     | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 2# 1     | 6.4 (128)                 | 13                |
| <i>P. aeruginosa</i> 4# 1     | 6.4 (128)                 | 13                |
| <i>P. aeruginosa</i> 7* 1     | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 9# 1     | 6.4 (128)                 | 13                |
| <i>P. aeruginosa</i> 10* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 11* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 12# 1    | 6.4 (128)                 | 13                |
| <i>P. aeruginosa</i> 13* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 14* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 16* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 17* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 18* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 19* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 20* 1    | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 244§ 1   | 1.6 (32)                  | 51                |
| <i>P. aeruginosa</i> 247* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 248* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 256# 1   | 6.4 (128)                 | 13                |
| <i>P. aeruginosa</i> 259* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 402* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 403* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 405* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 426* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 427* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 428* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 432§ 1   | 1.6 (32)                  | 51                |
| <i>P. aeruginosa</i> 433* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 434* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 435* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> 436* 1   | >6.4 (>128)               | <13               |
| <i>P. aeruginosa</i> ATCC* 1  | >6.4 (>128)               | <13               |
| <i>P. fluorescens</i> A8# 2   | 0.8 (16)                  | 101               |
| <i>P. fluorescens</i> SMM8# 2 | 1.6 (32)                  | 51                |
| <i>P. fluorescens</i> SMI1# 2 | 0.8 (16)                  | 101               |
| <i>P. fluorescens</i> SMI2# 2 | 0.8 (16)                  | 101               |
| <i>P. fluorescens</i> SMI6# 2 | 1.6 (32)                  | 51                |
| <i>P. fragi</i> G2# 2         | 1.6 (32)                  | 51                |
| <i>P. oleovorans</i> # 1      | 0.4 (8)                   | 202               |
| <i>P. putida</i> 262* 1       | 3.2 (64)                  | 25                |
| <i>P. putida</i> 407* 1       | 3.2 (64)                  | 25                |
| <i>P. putida</i> 409* 1       | 3.2 (64)                  | 25                |
| <i>P. putida</i> 410* 1       | 6.4 (128)                 | 13                |
| <i>P. putida</i> SMA1# 2      | 0.8 (16)                  | 101               |
| <i>P. straminea</i> A5# 2     | 1.6 (32)                  | 51                |
| <i>P. straminea</i> A7# 2     | 1.6 (32)                  | 51                |
| <i>P. straminea</i> A13# 2    | 1.6 (32)                  | 51                |
| <i>P. syringae</i> # 2        | 0.2 (4)                   | 404               |

\* clinical isolates; # environmental isolates; # not pigmented colonies (bacteria not producing pigments); \* yellowish-green, green-blue, blue strains (pyoverdine and/or pyocyanin-producers); § brown strains (pyomelanin-producers).

1. Alfei, S.; Caviglia, D.; Piatti, G.; Zuccari, G.; Schito, A.M. Bactericidal Activity of a Self-Biodegradable Lysine-Containing Dendrimer against Clinical Isolates of *Acinetobacter* Genus. *Int. J. Mol. Sci.* 2021, 22, 7274. <https://doi.org/10.3390/ijms22147274>.

### Strong Correlation Between the Antibacterial Effects of **G5-PDK** and the Production of Bacterial Pigments

**G5-PDK** showed MIC > 6.4  $\mu$ M on 78.1% of the *P. aeruginosa* strains reported in Table 1 which produced yellowish, yellow-green, green-blue or blue pigmented colonies (\* strains in Table 1, Fig. 2), because they are producers of pyoverdine and/or pyocyanin (Fig. 2). Non-pigmented *P. aeruginosa* isolates (strains 2, 4, 9, 12, Table 1, Fig. 2) showed lower MICs = 6.4  $\mu$ M. Finally, much lower MICs (1.6  $\mu$ M) were observed on brown pigmented *P. aeruginosa* strains 244 (Table 1, Fig. 2) and 432 (Table 1), which produce pyomelanin (Fig. 2).

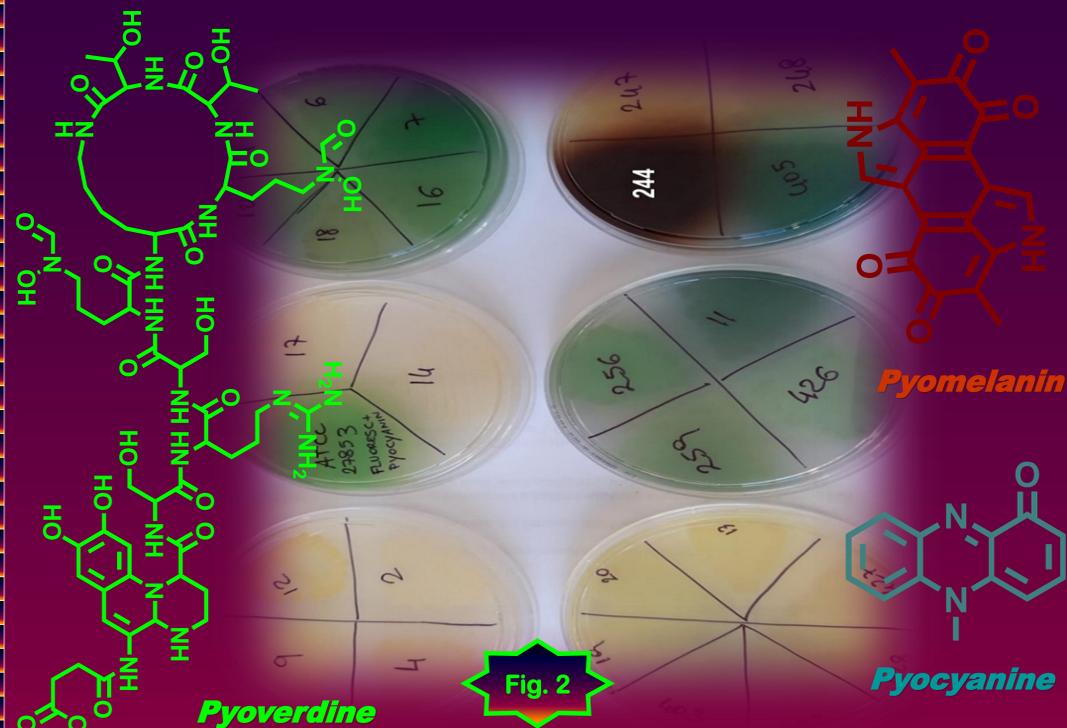


Fig. 2

Pyocyanin and pyoverdine are cationic molecules whose secretion can repel other cationic molecules, such as **G5-PDK**, thus justifying the low activity of **G5-PDK** towards the yellow to blue-green pigmented *P. aeruginosa* isolates of this study. On the contrary, pyomelanin is a negatively charged pigment which, once secreted, creates an anionic environment, favoring the adsorption of cationic antibacterial agents, such as **G5-PDK**. This may explain the high susceptibility of the pyomelanin producing strains in this study (strains 244 and 432) to **G5-PDK**. Regarding the non-pigmented strains of the *P. aeruginosa* species (strains 2,4,9 and 12), the interaction of **G5-PDK** with the bacterial surface was neither hindered nor promoted and MIC = 6.4  $\mu$ M were observed. Very low MICs (0.2,0.4,1,6  $\mu$ M) were observed on *P. straminea*, *P. fragi*, *P. oleovorans* and *P. syringae* because they were unable to produce pigments. With respect to *P. fluorescens* and *P. putida* producing pyoverdine only in iron deficiency, higher MICs have been observed for clinical isolates producing pyoverdine to steal iron from the human host.

### Conclusions

Overall, **G5-PDK** due to its low cytotoxicity and high SI, could represent a promising new antibacterial agent to limit serious human infections. Furthermore, it could also find applications to limit food waste and the destruction of economically significant plantations such as tobacco, kiwi and tomato.

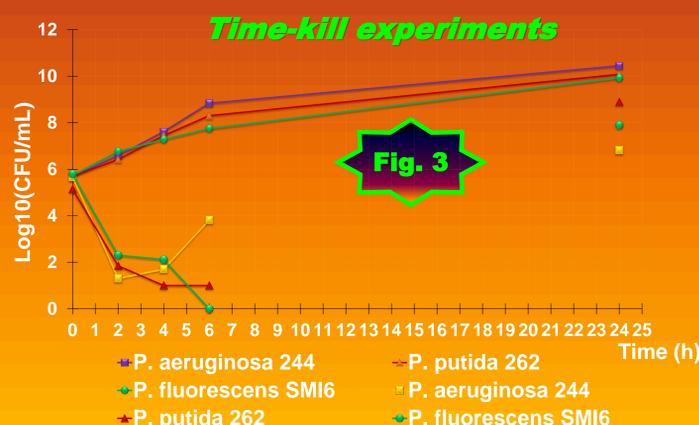


Fig. 3

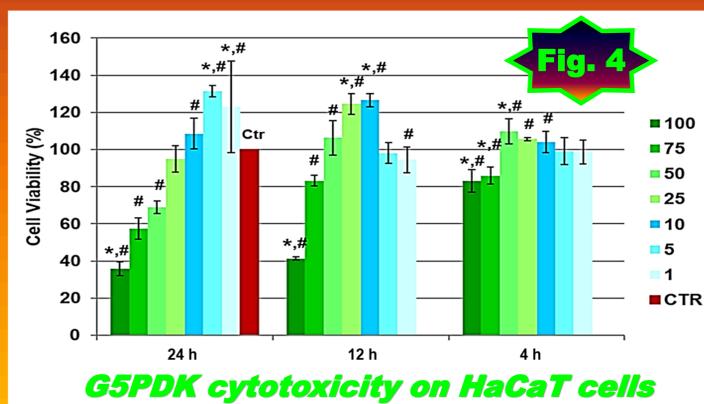


Fig. 4

G5PDK cytotoxicity on HaCaT cells