

A System-on-Chip Assay for Bilirubin Levels Measurement in Whole Blood †

Jean Pierre Ndabakuranye ^{1,2,*}, Steven Prawer ¹ and Arman Ahnood ^{1,2}

¹ School of Physics, University of Melbourne, Parkville, VIC 3010, Australia; s.prawer@unimelb.edu.au (S.P.); arman.ahnood@rmit.edu.au (A.A.)

² School of Engineering, RMIT University, Bundoora, VIC 3083, Australia

* Correspondence: jndabakuran@student.unimelb.edu.au; Tel.: +61-413-203-406

† Presented at the 8th International Electronic Conference on Sensors and Applications, 1–15 November 2021; Available online: <https://ecsa-8.sciforum.net>.

Abstract: Bilirubin (BR) is clinically confirmed as a biomarker for liver health and is used to assess the prognosis of cirrhosis. Optical and chemical methods have been utilized for blood BR biosensing. While optical methods offer real-time monitoring and are handy and immune to infection, measurements may not be practical due to the instrument complexity and space requirements. This study investigated the dual-wavelength (DWL) technique for BR estimation using a system-on-chip (SoC). The SoC includes an optical module with blue (455 nm) and green (530 nm) LEDs which were used for DWL measurement. Porcine blood was used as a surrogate of human blood and BR levels were kept within the pathophysiological ranges projected from healthy individuals (<1.2 mg/dL) to a cirrhotic patient (up to 50 mg/dL). Our findings show a high BR sensitivity in blood and this lays the groundwork for point-of-care testing for BR levels primarily for hyperbilirubinemia infants and cirrhotic adults out in homes or in-community settings.

Keywords: liver cirrhosis; biosensing; bilirubin; biomarker; blood; point-of-care testing

Citation: Ndabakuranye, J.P.;

Prawer, S.; Ahnood, A. A system-on-chip assay for bilirubin levels measurement in whole blood. *Eng. Proc.* **2021**, *3*, x. <https://doi.org/10.3390/xxxxx>

Academic Editor: Jean Pierre Ndabakuranye

Published: 1 November 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Bilirubin is a by-product of heme catabolism [1], where less than 1.2 mg/dL is always present in the blood of healthy individuals [2]. It exists as conjugated and unconjugated bilirubin (UCB) in the bloodstream. Unconjugated bilirubin (UCB) is toxic and water-insoluble; hence it should be excreted. For excretion, UCB binds with albumin and is transported to the liver through the bloodstream. In the liver, UCB is enzymatically converted into conjugated bilirubin (CB). The latter is less toxic and water-soluble, allowing excretion into the urine or poop [3]. However, in the case of pathophysiological events, bilirubin levels are elevated, and the liver fails to live up to its conjugation ability leading to irreversible neurological damage or death [4]. High bilirubin levels have been correlated with hepatic and hemolytic disorders [5,6].

Several studies on BR levels estimation have been reported [7,8], but these techniques suffer from the instrument complexity and cost. This mini-paper investigated the dual-wavelength (DWL) technique for BR estimation using a system-on-chip (SoC).

2. Materials and Methods

Materials: Bilirubin (PN: 14370), Dimethyl Sulfoxide (DMSO) and sodium citrate (NaCHO) commercial standards were supplied from Sigma-Aldrich, Australia.

Blood sample preparation: Porcine blood was procured from the abattoir immediately after sacrifice and was preferred as a prominent replacement for human blood due to its biochemical resemblance [9] and low cost. Solutions were prepared by mixing BR at varied concentrations and blood with anticoagulant (4% *w/v*)-to-blood ratio of 1:9 (*v/v*)

[10]. BR-blood samples were stored at $\sim 5^\circ\text{C}$ in the dark to prevent photodegradation. For extended preparation steps, refer to the study done by Ndabakuranye, et al. [11].

Optical measurement: A SoC platform was used for BR measurement. The setup consisted of a MAX86916 optical module, a MAX32630 host and an optical stage designed to operate in transfection mode (Figure 1). Major features of the system-on-chip are summarized in Table 1.

Table 1. Major features of MAX86916 optical module.

Feature	Value
ADC resolution	19-bit
Size	$3.5 \times 7.0 \times 1.5 \text{ mm}$
Spectral sensitivity	400–1100 nm
Data communication protocol	I2C
Radiant sensitive area	1.51 mm^2

Feature selection: Several features were optimally selected using theoretical investigations and simulations to ensure the accuracy, reliability, and safety of bilirubin measurements. These features are summarized in Table 2.

Table 2. Optimal features for DWL measurement.

Method Parameter	Optimized Parameter
Operating wavelengths	Blue and green
Path length	$200 \mu\text{m}$
Solvent	DMSO
Bilirubin concentration range	1.2–50 mg/dL
Anticoagulant	Sodium citrate

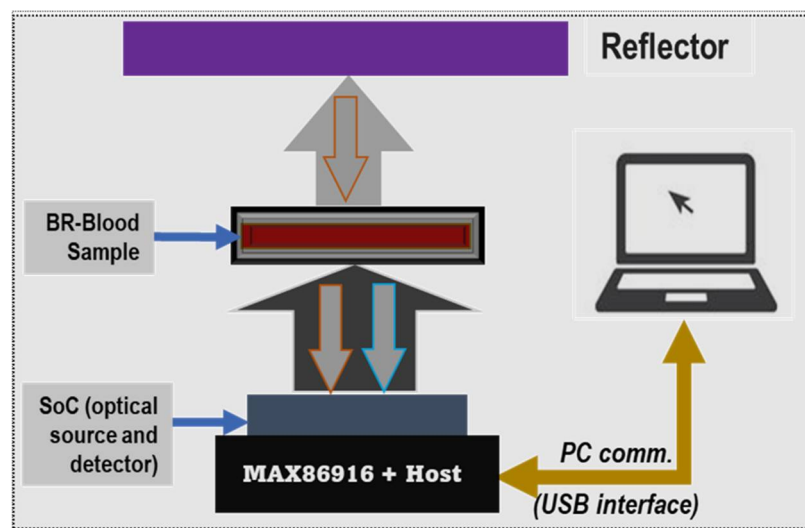


Figure 1. Illustration of the DWL measurement in transfection mode.

3. Results and Discussions

The analytical (470 nm) and reference (525 nm) wavelengths were obtained by analyzing the distinct optical signatures of blood and BR. It does not significantly absorb green light but strongly absorbs blue light ($\epsilon_{525} = 214$, $\epsilon_{460} = 53869 \text{ [cm}^{-1} \text{ M}^{-1}]$) [12]. However, although our SoC has 455 and 530 nm as the maximum power outputs, they still include 470 and 525 nm respectively since LEDs are not ideal monochromatic sources.

The DWL measurement was performed using a revised 2-dimensional Beer's law as shown in Equation (1) which suggests that BR concentration can be correlated with its absorbance ratio at 470 and 525 nm (t : path length [cm], ϵ : extinction coefficient, C : concentration).

$$\begin{pmatrix} A_{470nm} \\ A_{525nm} \end{pmatrix} = \begin{bmatrix} \epsilon_{BR, 470} & \epsilon_{Hb, 470} \\ \epsilon_{BR, 525} & \epsilon_{Hb, 525} \end{bmatrix} \begin{pmatrix} C_{BR} \\ C_{Hb} \end{pmatrix} \times t \quad (1)$$

$$\tilde{R} = \frac{\log[ADC\ count_{Blue}]}{\log[ADC\ count_{Green}]} \quad (2)$$

To investigate BR's sensitivity in blood, R-parameters were deduced by collecting the MAX86916 ADC counts data and calculated using Equation (2). Figure 1 shows the SoC platform used to measure the ADC data used to compute R-parameters. The plot of R-parameters vs. BR concentrations is shown in Figure 2, and results showed a strong linear relationship with R-square greater than 0.991.

Although this technique provides a robust and simple way of measuring BR, it may be susceptible to errors due to LEDs' spectral and spatial distribution inadequacies, residual bilirubin, other hemoglobin forms (COHb or MetHb), SaO2 and Hb levels variability.

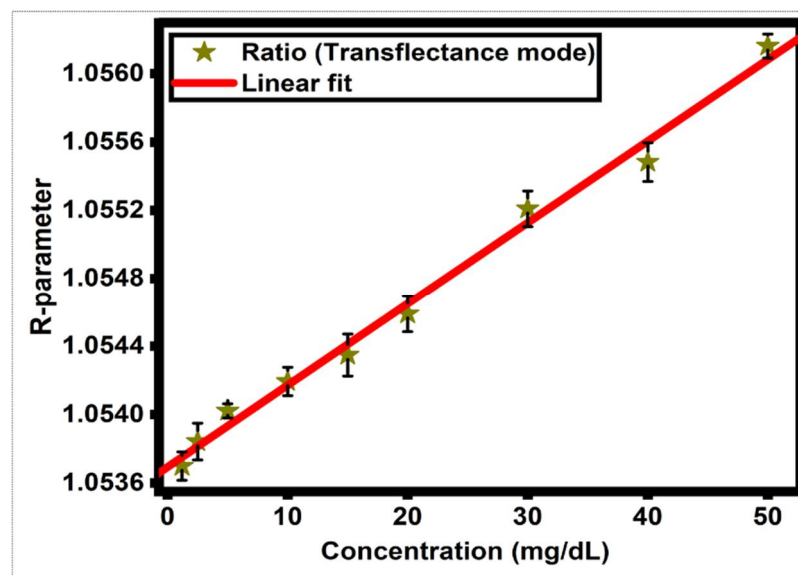


Figure 2. The graph of R-parameters as a function of bilirubin concentrations as obtained from DWL measurements on whole blood using the SoC platform.

4. Conclusions

The feasibility of BR monitoring by the DWL method was investigated. BR's sensitivity in blood was explored at pathophysiological ranges (1.2–50 mg/dL) using an Soc. The SoC includes a miniature MAX86916 optical module with integrated signal conditioning and processing capabilities. Results showed a strong correlation between R-parameters and BR concentration (R-squared > 0.99). Our findings lay the groundwork for point-of-care testing for BR levels, primarily for hyperbilirubinemia infants and cirrhotic adults out of clinical settings.

Institutional Review Board Statement:

Informed Consent Statement:

Data Availability Statement:

Acknowledgments: Jean Pierre gratefully acknowledges the Melbourne Research Scholarship (the University of Melbourne).

References

1. Ahmad, H.B.; Ahmad, S.; Shad, M.A.; Hussain, M. Kinetic measurements for photodecomposition of bilirubin. *Asian J. Chem.* **2013**, *25*, 7945–7948. <http://dx.doi.org/10.14233/ajchem.2013.14749>.
2. Dufour, D.R.; Lott, J.A.; Nolte, F.S.; Gretch, D.R.; Koff, R.S.; Seeff, L.B. Diagnosis and monitoring of hepatic injury. I. Performance characteristics of laboratory tests. *Clin. Chem.* **2000**, *46*, 2027–2049. <https://doi.org/10.1093/clinchem/46.12.2027>.
3. Berk, P.D. Bilirubin metabolism and the hereditary hyperbilirubinemias. *Semin. Liver Dis.* **1994**, *14*, 321–322. <https://doi.org/10.1055/s-2007-1007321>.
4. AAP. Management of hyperbilirubinemia in the newborn infant 35 or more weeks of gestation. *Pediatrics* **2004**, *114*, 297–316. <https://doi.org/10.1542/peds.114.1.297>.
5. Lopez-Velazquez, J.A.; Chavez-Tapia, N.C.; Ponciano-Rodriguez, G.; Sanchez-Valle, V.; Caldwell, S.H.; Uribe, M.; Mendez-Sanchez, N. Bilirubin alone as a biomarker for short-term mortality in acute-on-chronic liver failure: An important prognostic indicator. *Ann. Hepatol.* **2014**, *13*, 98–104. [https://doi.org/10.1016/S1665-2681\(19\)30910-X](https://doi.org/10.1016/S1665-2681(19)30910-X).
6. D'Amico, G.; Garcia Tsao, G.; Pagliaro, L. Natural history and prognostic indicators of survival in cirrhosis: A systematic review of 118 studies. *J. Hepatol.* **2006**, *44*, 217–231. <https://doi.org/10.1016/j.jhep.2005.10.013>.
7. Hulzebos, C.V.; Vitek, L.; Zabetta, C.D.C.; Dvořák, A.; Schenk, P.; van der Hagen, E.A.; Cobbaert, C.; Tiribelli, C. Diagnostic methods for neonatal hyperbilirubinemia: Benefits, limitations, requirements, and novel developments. *Pediatric Res.* **2021**, *90*, 277–283.
8. Rawal, R.; Kharangarh, P.R.; Dawra, S.; Tomar, M.; Gupta, V.; Pundir, C. A comprehensive review of bilirubin determination methods with special emphasis on biosensors. *Process Biochem.* **2020**, *89*, 165–174. <https://doi.org/10.1016/j.procbio.2019.10.034>.
9. Pond, W.G.; Mersmann, H.J. *Biology of the Domestic Pig*; Comstock Pub. Associates, Cornell University Press: 2001.
10. Sorapukdee, S.; Narunatsopanon, S. Comparative study on compositions and functional properties of porcine, chicken and duck blood. *Korean J. Food Sci. Anim. Resour.* **2017**, *37*, 228–241. <https://doi.org/10.5851/kosfa.2017.37.2.228>.
11. Ndabakuranye, J.P.; Rajapaksa, A.E.; Burchall, G.; Li, S.; Prawer, S.; Ahnood, A. A novel optical assay system for bilirubin concentration measurement in whole blood. *IEEE Trans. Biomed. Eng.* **2021**. <https://doi.org/10.1109/TBME.2021.3111150>.
12. Yao, J.; Wang, L.V. Photoacoustic microscopy. *Laser Photonics Rev.* **2013**, *7*, 758–778. <https://doi.org/10.1002/lpor.201200060>.