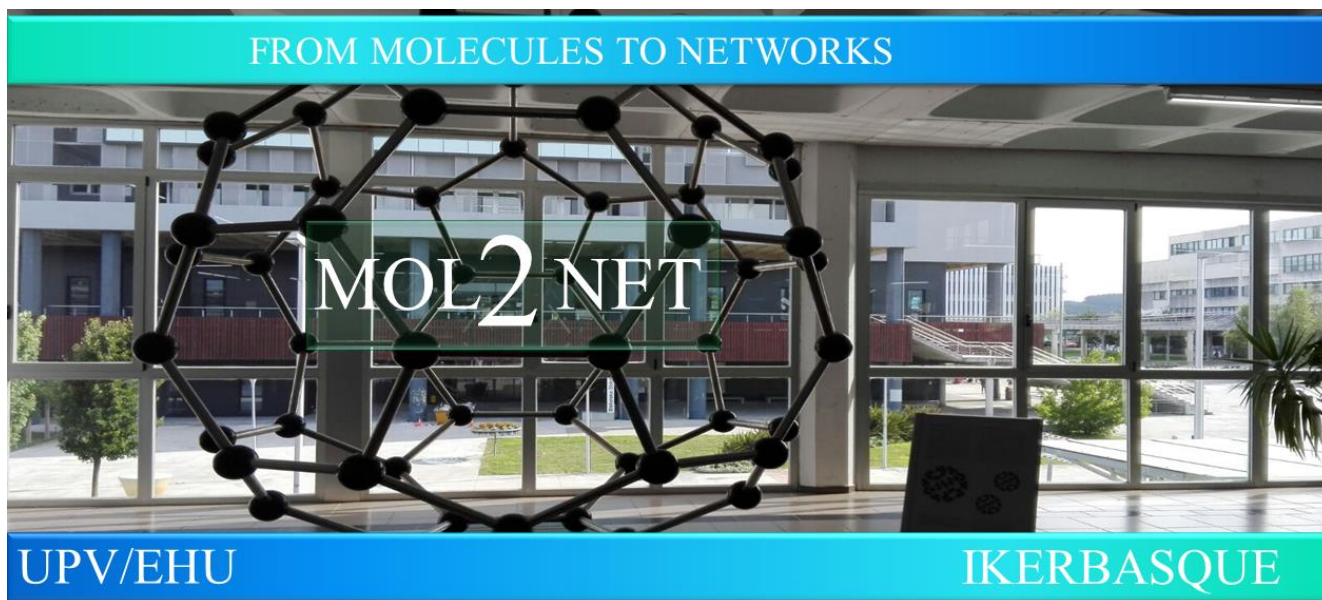




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PD-L1/PD-1 Blockade Immunotherapy: Mechanism of response and resistance

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Abstract.

Cancer immunotherapies targeting the PD-L1/PD-1 axis are revolutionizing cancer treatment and transforming the practice of medical oncology. Despite the recent successes, most patients are refractory and present intrinsic or acquired resistance. This is one of the most significant challenges in oncology. However, the mechanisms leading to resistance are still poorly understood. Here, we discuss some of the major molecular mechanisms of resistance to PD-L1/PD-1 blockade. We also argue whether tumor intrinsic or extrinsic factors constitute main determinants of response and resistance. Primary and acquired resistances are important barriers in terms of benefit to the patient. Among others, tumor-intrinsic factors include poor tumor antigenicity, DNA repair mutational load, alterations in the regulation of oncogenic pathways, defects in IFN signal transduction and PD-L1 expression. On the other hand, tumor-extrinsic factors include T cell exhaustion, expression of additional immune checkpoint molecules, differentiation and expansion of immunosuppressive cell populations, and release of immunosuppressive cytokines and metabolites. The relative contribution of tumor cell intrinsic and extrinsic factors to primary, adaptive, and acquired resistance is yet unclear. Our group recently found that NSCLC patients that failed to respond to PD-1/PD-L1 inhibitors had systemic CD4 dysfunctionality, characterized by increased PD-1/LAG-3 co-expression in T cells. These results highlighted dysfunctional CD4 T cells as main contributors to treatment failure. A deeper understanding of the basic mechanisms underlying resistance will provide insight for further development of better therapeutic strategies by overcoming treatment failure. The development of new approaches to improve CD4 responses before immunotherapy could be a solution to overcome resistance (Zuazo et al, EMBO Mol Med 2019; Zuazo et al, EMBO Mol Med 2020; Chocarro de Erauso L et al, Front. Pharmacol. 2020; Zuazo et al, Front Immunol 2020; Bocanegra et al, Int J Mol Sci 2020; Hernández et al, Int J Mol Sci 2020; Arasanz et al, Cancers 2020; Chocarro et al, Int J Mol Sci 2021)

Keywords: PD-1, PD-L1, immunotherapy, treatment response, resistance mechanisms

References

1. Huard, B.; Tournier, M.; Hercend, T.; Triebel, F.; Faure, F. Lymphocyte-Activation Gene 3/Major Histocompatibility Complex Class II Interaction Modulates the Antigenic Response of CD4+ T Lymphocytes. *Eur. J. Immunol.* **1994**, *24*, 3216–3221. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
2. Huard, B.; Prigent, P.; Pagès, F.; Bruniquel, D.; Triebel, F. T Cell Major Histocompatibility Complex Class II Molecules Down-Regulate CD4+ T Cell Clone Responses Following LAG-3 Binding. *Eur. J. Immunol.* **1996**, *26*, 1180–1186. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
3. Workman, C.J.; Dugger, K.J.; Vignali, D.A.A. Cutting Edge: Molecular Analysis of the Negative Regulatory Function of Lymphocyte Activation Gene-3. *J. Immunol.* **2002**, *169*, 5392–5395. [[Google Scholar](#)] [[CrossRef](#)]
4. Workman, C.J.; Rice, D.S.; Dugger, K.J.; Kurschner, C.; Vignali, D.A.A. Phenotypic Analysis of the Murine CD4-Related Glycoprotein, CD223 (LAG-3). *Eur. J. Immunol.* **2002**, *32*, 2255–2263. [[Google Scholar](#)] [[CrossRef](#)]

5. Workman, C.J.; Cauley, L.S.; Kim, I.-J.; Blackman, M.A.; Woodland, D.L.; Vignali, D.A.A. Lymphocyte Activation Gene-3 (CD223) Regulates the Size of the Expanding T Cell Population Following Antigen Activation in vivo. *J. Immunol.* **2004**, *172*, 5450–5455. [[Google Scholar](#)] [[CrossRef](#)]
6. Maçon-Lemaître, L.; Triebel, F. The Negative Regulatory Function of the Lymphocyte-Activation Gene-3 Co-Receptor (CD223) on Human T Cells. *Immunology* **2005**, *115*, 170–178. [[Google Scholar](#)] [[CrossRef](#)]
7. Andrews, L.P.; Marciscano, A.E.; Drake, C.G.; Vignali, D.A.A. LAG3 (CD223) as a Cancer Immunotherapy Target. *Immunol. Rev.* **2017**, *276*, 80–96. [[Google Scholar](#)] [[CrossRef](#)]
8. Triebel, F.; Jitsukawa, S.; Baixeras, E.; Roman-Roman, S.; Genevee, C.; Viegas-Pequignot, E.; Hercend, T. LAG-3, a Novel Lymphocyte Activation Gene Closely Related to CD4. *J. Exp. Med.* **1990**, *171*, 1393–1405. [[Google Scholar](#)] [[CrossRef](#)]
9. Baixeras, B.E.; Huard, B.; Miossec, C.; Jitsukawa, S.; Martin, M.; Hercend, T.; Auffray, C.; Triebel, F.; Tonneau-Piatier, D. Characterization of the Lymphocyte Activation Gene 3-Encoded Protein. A New Ligand for Human Leukoc3~ Antigen Class H Antigens. *Pharm. Res.* **1992**, *176*, 327–337. [[Google Scholar](#)]
10. Annunziato, F.; Manetti, R.; Tomasévic, I.; Giudizi, M.; Biagiotti, R.; Giannò, V.; Germano, P.; Mavilia, C.; Maggi, E.; Romagnani, S. Expression and Release of LAG-3-Encoded Protein by Human CD4 + T Cells Are Associated with IFN- γ Production. *FASEB J.* **1996**, *10*, 769–776. [[Google Scholar](#)] [[CrossRef](#)]
11. Avice, M.N.; Sarfati, M.; Triebel, F.; Delespesse, G.; Demeure, C.E. Lymphocyte Activation Gene-3, a MHC Class II Ligand Expressed on Activated T Cells, Stimulates TNF-Alpha and IL-12 Production by Monocytes and Dendritic Cells. *J. Immunol.* **1999**, *162*, 2748–2753. [[Google Scholar](#)]
12. Slevin, S.M.; Garner, L.C.; Lahiff, C.; Tan, M.; Wang, L.M.; Ferry, H.; Greenaway, B.; Lynch, K.; Geremia, A.; Hughes, S.; et al. Lymphocyte Activation Gene (LAG)-3 Is Associated with Mucosal Inflammation and Disease Activity in Ulcerative Colitis. *J. Crohn's Colitis.* **2020**, *14*, 1446–1461. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
13. Hannier, S.; Tournier, M.; Bismuth, G.; Triebel, F. CD3/TCR Complex-Associated Lymphocyte Activation Gene-3 Molecules Inhibit CD3/TCR Signaling. *J. Immunol.* **1998**, *161*, 4058–4065. [[Google Scholar](#)] [[PubMed](#)]
14. Blackburn, S.D.; Shin, H.; Haining, W.N.; Zou, T.; Workman, C.J.; Polley, A.; Betts, M.R.; Freeman, G.J.; Vignali, D.A.A.; Wherry, E.J. Coregulation of CD8+ T Cell Exhaustion by Multiple Inhibitory Receptors During Chronic Viral Infection. *Nat. Immunol.* **2009**, *10*, 29–37. [[Google Scholar](#)] [[CrossRef](#)]
15. Chihara, N.; Madi, A.; Kondo, T.; Zhang, H.; Acharya, N.; Singer, M.; Nyman, J.; Marjanovic, N.D.; Kowalczyk, M.S.; Wang, C.; et al. Induction and Transcriptional Regulation of the Co-Inhibitory Gene Module in T Cells. *Nature* **2018**, *558*, 454–459. [[Google Scholar](#)] [[CrossRef](#)]
16. Grosso, J.F.; Kelleher, C.C.; Harris, T.J.; Maris, C.H.; Hipkiss, E.L.; De Marzo, A.; Anders, R.; Netto, G.; Derese, G.; Bruno, T.C.; et al. LAG-3 Regulates CD8 + T Cell Accumulation and Effector Function in Murine Self—and Tumor-Tolerance Systems. *J. Clin. Invest.* **2007**, *117*, 3383–3392. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
17. Grosso, J.F.; Goldberg, M.V.; Getnet, D.; Bruno, T.C.; Yen, H.-R.; Pyle, K.J.; Hipkiss, E.; Vignali, D.A.A.; Pardoll, D.M.; Drake, C.G. Functionally Distinct LAG-3 and PD-1 Subsets on Activated and Chronically Stimulated CD8 T Cells. *J. Immunol.* **2009**, *182*, 6659–6669. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
18. Huang, R.-Y.; Eppolito, C.; Lele, S.; Shrikant, P.; Matsuzaki, J.; Odunsi, K. LAG3 And PD1 Co-Inhibitory Molecules Collaborate to Limit CD8+ T Cell Signaling and Dampen Antitumor Immunity in a Murine Ovarian Cancer Model. *Oncotarget* **2015**, *6*, 27359. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
19. Williams, J.B.; Horton, B.L.; Zheng, Y.; Duan, Y.; Powell, J.D.; Gajewski, T.F. The EGR2 Targets LAG-3 and 4-1BB Describe and Regulate Dysfunctional Antigen-Specific CD8 + T Cells in the Tumor Microenvironment. *J. Exp. Med.* **2017**, *214*, 381–400. [[Google Scholar](#)] [[CrossRef](#)]

20. Huard, B.; Mastrangeli, R.; Prigent, P.; Bruniquel, D.; Donini, S.; El-Tayar, N.; Maigret, B.; Dreano, M.; Triebel, F. Characterization of the Major Histocompatibility Complex Class II Binding Site on LAG-3 Protein. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 5744–5749. [[Google Scholar](#)] [[CrossRef](#)]
21. Li, N.; Workman, C.J.; Martin, S.M.; Vignali, D.A.A. Biochemical Analysis of the Regulatory T Cell Protein Lymphocyte Activation Gene-3 (LAG-3; CD223). *J. Immunol.* **2004**, *173*, 6806–6812. [[Google Scholar](#)] [[CrossRef](#)]
22. Li, N.; Wang, Y.; Forbes, K.; Vignali, K.M.; Heale, B.S.; Saftig, P.; Hartmann, D.; Black, R.A.; Rossi, J.J.; Blobel, C.P.; et al. Metalloproteases Regulate T-Cell Proliferation and Effector Function Via LAG-3. *EMBO J.* **2007**, *26*, 494–504. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
23. Mastrangeli, R.; Micangeli, E.; Donini, S. Cloning of Murine LAG-3 by Magnetic Bead Bound Homologous Probes and PCR (GENE-CAPTURE PCR). *Anal. Biochem.* **1996**, *241*, 93–102. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
24. Workman, C.J.; Vignali, D.A.A. The CD4-Related Molecule, LAG-3 (CD223), Regulates the Expansion of Activated T Cells. *Eur. J. Immunol.* **2003**, *3*, 970–979. [[Google Scholar](#)] [[CrossRef](#)]
25. Maeda, T.K.; Sugiura, D.; Okazaki, M.T.; Okazaki, T. Atypical Motifs in the Cytoplasmic Region of the Inhibitory Immune Co-Receptor LAG-3 Inhibit T Cell Activation. *J. Biol. Chem.* **2019**, *294*, 6017–6026. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
26. Iouzalén, N.; Andrae, S.; Hannier, S.; Triebel, F. LAP, A Lymphocyte Activation Gene-3 (LAG-3)-Associated Protein that Binds to a Repeated EP Motif in the Intracellular Region of LAG-3, May Participate in the Down-Regulation of the CD3/TCR Activation Pathway. *Eur. J. Immunol.* **2001**, *31*, 2885–2891. [[Google Scholar](#)] [[CrossRef](#)]
27. Bae, J.; Lee, S.J.; Park, C.-G.; Lee, Y.S.; Chun, T. Trafficking of LAG-3 to the Surface on Activated T Cells via Its Cytoplasmic Domain and Protein Kinase C Signaling. *J. Immunol.* **2014**, *193*, 3101–3112. [[Google Scholar](#)] [[CrossRef](#)]
28. Long, L.; Zhang, X.; Chen, F.; Pan, Q.; Phiphatwatchara, P. The Promising Immune Checkpoint LAG-3: From Tumor Microenvironment to Cancer Immunotherapy. *Genes Cancer* **2018**, *9*, 176. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
29. Huard, B.; Prigent, P.; Tournier, M.; Bruniquel, D.; Triebel, F. CD4/Major Histocompatibility Complex Class II Interaction Analyzed with CD4—and Lymphocyte Activation Gene-3 (LAG-3)-Ig Fusion Proteins. *Eur. J. Immunol.* **1995**, *25*, 2718–2721. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
30. Hemon, P.; Jean-Louis, F.; Ramgolam, K.; Brignone, C.; Viguier, M.; Bachelez, H.; Triebel, F.; Charron, D.; Aoudjit, F.; Al-Daccak, R.; et al. MHC Class II Engagement by Its Ligand LAG-3 (CD223) Contributes to Melanoma Resistance to Apoptosis. *J. Immunol.* **2011**, *186*, 5173–5183. [[Google Scholar](#)] [[CrossRef](#)]
31. Donia, M.; Andersen, R.; Kjeldsen, J.W.; Fagone, P.; Munir, S.; Nicoletti, F.; Andersen, M.H.; Straten, P.T.; Svane, I.M. Aberrant Expression of MHC Class II in Melanoma Attracts Inflammatory Tumor-Specific CD4+T-Cells, which Dampen CD8+T-Cell Antitumor Reactivity. *Cancer Res.* **2015**, *75*, 3747–3759. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
32. Chung, L.Y.; Tang, S.J.; Wu, Y.C.; Sun, G.H.; Liu, H.Y.; Sun, K.H. Galectin-3 Augments Tumor Initiating Property and Tumorigenicity of Lung Cancer through Interaction with B-Catenin. *Oncotarget* **2015**, *6*, 4936–4952. [[Google Scholar](#)] [[CrossRef](#)]
33. Lu, W.; Wang, J.; Yang, G.; Yu, N.; Huang, Z.; Xu, H.; Li, J.; Qiu, J.; Zeng, X.; Chen, S.; et al. Posttranscriptional Regulation of Galectin-3 by miR-128 Contributes to Colorectal Cancer Progression. *Oncotarget* **2017**, *8*, 15242–15251. [[Google Scholar](#)] [[CrossRef](#)]

34. Li, M.; Feng, Y.M.; Fang, S.Q. Overexpression of Ezrin and Galectin-3 as Predictors of Poor Prognosis of Cervical Cancer. *Braz. J. Med. Biol. Res.* **2017**, *50*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
35. Kouo, T.; Huang, L.; Pucsek, A.B.; Cao, M.; Solt, S.; Armstrong, T.; Jaffee, E. Galectin-3 Shapes Antitumor Immune Responses by Suppressing CD8+ T Cells via LAG-3 and Inhibiting Expansion of Plasmacytoid Dendritic Cells. *Cancer Immunol. Res.* **2015**, *3*, 412–423. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
36. Wang, J.; Sanmamed, M.F.; Datar, I.; Su, T.T.; Ji, L.; Sun, J.; Chen, L.; Chen, Y.; Zhu, G.; Yin, W.; et al. Fibrinogen-like Protein 1 Is a Major Immune Inhibitory Ligand of LAG-3. *Cell* **2019**, *176*, 334–347.e12. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
37. Huard, B.; Gaulard, P.; Faure, F.; Hercend, T.; Triebel, F. Cellular Expression and Tissue Distribution of the Human LAG-3-Encoded Protein, An MHC Class II Ligand. *Immunogenetics* **1994**, *39*, 213–217. [[Google Scholar](#)] [[CrossRef](#)]
38. Annunziato, F.; Manetti, R.; Cosmi, L.; Galli, G.; Heusser, C.H.; Romagnani, S.; Maggi, E. Opposite Role for Interleukin-4 and Interferon- γ on CD30 and Lymphocyte Activation Gene-3 (LAG-3) Expression by Activated Naive T Cells. *Eur. J. Immunol.* **1997**, *27*, 2239–2244. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
39. Bruniquel, D.; Borie, N.; Hannier, S.; Triebel, F. Regulation of Expression of the Human Lymphocyte Activation Gene-3 (LAG-3) Molecule, a Ligand for MHC Class II. *Immunogenetics* **1998**, *48*, 116–124. [[Google Scholar](#)] [[CrossRef](#)]
40. Burton, B.R.; Britton, G.J.; Fang, H.; Verhagen, J.; Smithers, B.; Sabatos-Peyton, C.A.; Carney, L.J.; Gough, J.; Strobel, S.; Wraith, D.C. Sequential Transcriptional Changes Dictate Safe and Effective Antigen-Specific Immunotherapy. *Nat. Commun.* **2014**. [[Google Scholar](#)] [[CrossRef](#)]
41. Matsuzaki, J.; Gnjatic, S.; Mhawech-Fauceglia, P.; Beck, A.; Miller, A.; Tsuji, T.; Eppolito, C.; Qian, F.; Lele, S.; Shrikant, P.; et al. Tumor-Infiltrating NY-ESO-1-Specific CD8 + T Cells are Negatively Regulated by LAG-3 and PD-1 in Human Ovarian Cancer. *Proc. Natl. Acad. Sci USA* **2010**, *107*, 7875–7880. [[Google Scholar](#)] [[CrossRef](#)]
42. Zuazo, M.; Arasanz, H.; Fernández-Hinojal, G.; García-Granda, M.J.; Gato, M.; Bocanegra, A.; Martínez, M.; Hernández, B.; Teijeira, L.; Morilla, I.; et al. Functional Systemic CD4 Immunity Is Required for Clinical Responses to PD-L1/PD-1 Blockade Therapy. *EMBO Mol. Med.* **2019**, *11*, e10293. [[Google Scholar](#)] [[CrossRef](#)]
43. Huard, B.; Tournier, M.; Triebel, F. LAG-3 Does Not Define a Specific Mode of Natural Killing in Human. *Immunol. Lett.* **1998**, *61*, 109–112. [[Google Scholar](#)] [[CrossRef](#)]
44. Huang, C.T.; Workman, C.J.; Flies, D.; Pan, X.; Marson, A.L.; Zhou, G.; Hipkiss, E.L.; Ravi, S.; Kowalski, J.; Levitsky, H.I.; et al. Role of LAG-3 in Regulatory T cells. *Immunity* **2004**, *21*, 503–513. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
45. Gagliani, N.; Magnani, C.F.; Huber, S.; Gianolini, M.E.; Pala, M.; Licona-Limon, P.; Guo, B.; Herbert, D.B.R.; Bulfone, A.; Trentini, F.; et al. Coexpression of CD49b and LAG-3 Identifies Human and Mouse T Regulatory Type 1 Cells. *Nat. Med.* **2013**, *19*, 739–746. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
46. Alfen, J.S.; Larghi, P.; Facciotti, F.; Gagliani, N.; Bosotti, R.; Paroni, M.; Maglie, S.; Gruarin, P.; Vasco, C.M.; Ranzani, V.; et al. Intestinal IFN- γ -Producing Type 1 Regulatory T Cells Coexpress CCR5 and Programmed Cell Death Protein 1 and Downregulate IL-10 in the Inflamed Guts of Patients with Inflammatory Bowel Disease. *J. Allergy Clin. Immunol.* **2018**, *142*, 1537–1547.e8. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
47. White, A.M.; Wraith, D.C. Tr1-Like T Cells—An Enigmatic Regulatory T Cell Lineage. *Front. Immunol.* **2016**, *7*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
48. Do, J.S.; Visperas, A.; Sanogo, Y.O.; Bechtel, J.J.; Dvorina, N.; Kim, S.; Jang, E.; Stohlman, S.A.; Shen, B.; Fairchild, R.L.; et al. An IL-27/Lag3 Axis Enhances Foxp3+ Regulatory T Cell-Suppressive Function and Therapeutic Efficacy. *Mucosal. Immunol.* **2016**, *9*, 137–145. [[Google Scholar](#)] [[CrossRef](#)]

49. Workman, C.J.; Vignali, D.A.A. Negative Regulation of T Cell Homeostasis by Lymphocyte Activation Gene-3 (CD223). *J. Immunol.* **2005**, *174*, 688–695. [[Google Scholar](#)] [[CrossRef](#)]
50. Camisaschi, C.; Casati, C.; Rini, F.; Perego, M.; De Filippo, A.; Triebel, F.; Parmiani, G.; Belli, F.; Rivoltini, L.; Castelli, C. LAG-3 Expression Defines a Subset of CD4 (+) CD25(High)Foxp3 (+) Regulatory T Cells that Are Expanded at Tumor Sites. *J. Immunol.* **2010**, *184*, 6545–6551. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
51. Gandhi, M.K.; Lambley, E.; Duraiswamy, J.; Dua, U.; Smith, C.; Elliott, S.; Gill, D.; Marlton, P.; Seymour, J.; Khanna, R. Expression of LAG-3 by Tumor-Infiltrating Lymphocytes Is Coincident with the Suppression of Latent Membrane Antigen-Specific CD8+ T-Cell Function in Hodgkin Lymphoma Patients. *Blood* **2006**, *108*, 2280–2289. [[Google Scholar](#)] [[CrossRef](#)]
52. Hald, S.M.; Rakaee, M.; Martinez, I.; Richardsen, E.; Al-Saad, S.; Paulsen, E.E.; Blix, E.S.; Kilvaer, T.; Andersen, S.; Busund, L.T.; et al. LAG-3 in Non-Small-cell Lung Cancer: Expression in Primary Tumors and Metastatic Lymph Nodes Is Associated with Improved Survival. *Clin. Lung Cancer* **2018**, *19*, 249–259.e2. [[Google Scholar](#)] [[CrossRef](#)]
53. Lee, S.J.; Jun, S.Y.; Lee, I.H.; Kang, B.W.; Park, S.Y.; Kim, H.J.; Park, J.S.; Choi, G.-S.; Yoon, G.; Kim, J.G. CD274, LAG3, and IDO1 Expressions in Tumor-Infiltrating Immune Cells as Prognostic Biomarker for Patients with MSI-High Colon Cancer. *J. Cancer Res. Clin. Oncol.* **2018**, *144*, 1005–1014. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
54. Zhang, Y.; Yongdong, L.; Yiling, L.; Binliu, L.; Qitao, H.; Wang, F.; Zhong, Q. Prognostic Value of Lymphocyte Activation Gene-3 (LAG-3) Expression in Esophageal Squamous Cell Carcinoma. *J. Cancer* **2018**, *9*, 4287–4293. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
55. Kisielow, M.; Kisielow, J.; Capoferri-Sollami, G.; Karjalainen, K. Expression of Lymphocyte Activation Gene 3 (LAG-3) on B Cells Is Induced by T Cells. *Eur. J. Immunol.* **2005**, *35*, 2081–2088. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
56. Lino, A.C.; Dang, V.D.; Lampropoulou, V.; Welle, A.; Joedicke, J.; Pohar, J.; Simon, Q.; Thalmensi, J.; Baures, A.; Fluhler, V.; et al. LAG-3 Inhibitory Receptor Expression Identifies Immunosuppressive Natural Regulatory Plasma Cells. *Immunity* **2018**, *49*, 120–133.e9. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
57. Khsheibun, R.; Paperna, T.; Volkowich, A.; Lejbkovicz, I.; Avidan, N.; Miller, A. Gene Expression Profiling of the Response to Interferon Beta in Epstein-Barr-Transformed and Primary B Cells of Patients with Multiple Sclerosis. *PLoS ONE* **2014**, *9*, 102331. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
58. Andrae, S.; Piras, F.; Burdin, N.; Triebel, F. Maturation and Activation of Dendritic Cells Induced by Lymphocyte Activation Gene-3 (CD223). *J. Immunol.* **2002**, *168*, 3874–3880. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
59. Guo, W.; Zhou, M.; Qiu, J.; Lin, Y.; Chen, X.; Huang, S.; Mo, M.; Liu, H.; Peng, G.; Zhu, X.; et al. Association of LAG3 Genetic Variation with an Increased Risk of PD in Chinese Female Population. *J. Neuroinflammation* **2019**, *16*. [[Google Scholar](#)] [[CrossRef](#)]
60. Angelopoulou, E.; Paudel, Y.N.; Villa, C.; Shaikh, M.F.; Piperi, C. Lymphocyte-Activation Gene 3 (LAG3) Protein as a Possible Therapeutic Target for Parkinson's Disease: Molecular Mechanisms Connecting Neuroinflammation to α -Synuclein Spreading Pathology. *Biology* **2020**, *9*, 86. [[Google Scholar](#)] [[CrossRef](#)]
61. Mao, X.; Ou, M.T.; Karuppagounder, S.S.; Kam, T.I.; Yin, X.; Xiong, Y.; Ge, P.; Umanah, G.E.; Brahmachari, S.; Shin, J.; et al. Pathological α -Synuclein Transmission Initiated by Binding Lymphocyte-Activation Gene 3. *Science*. **2016**, *353*, 6307. [[Google Scholar](#)] [[CrossRef](#)]
62. Cunningham, F.; Achuthan, P.; Akanni, W.; Allen, J.; Amode, M.R.; Armean, I.M.; Bennett, R.; Bhai, J.; Billis, K.; Boddu, S.; et al. Ensembl 2019. *Nucleic Acids Res.* **2019**, *47*. [[Google Scholar](#)] [[CrossRef](#)]

63. Saleh, R.; Toor, S.M.; Nair, V.S.; Elkord, E. Role of Epigenetic Modifications in Inhibitory Immune Checkpoints in Cancer Development and Progression. *Front. Immunol.* **2020**, *11*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
64. Goltz, D.; Gevensleben, H.; Grünen, S.; Dietrich, J.; Kristiansen, G.; Landsberg, J.; Dietrich, D. PD-L1 (CD274) Promoter Methylation Predicts Survival in Patients with Acute Myeloid Leukemia. *Leukemia* **2017**, *31*, 738–743. [[Google Scholar](#)] [[CrossRef](#)]
65. Goltz, D.; Gevensleben, H.; Dietrich, J.; Ellinger, J.; Landsberg, J.; Kristiansen, G.; Dietrich, D. Promoter Methylation of the Immune Checkpoint Receptor PD-1 (PDCD1) Is an Independent Prognostic Biomarker for Biochemical Recurrence-Free Survival in Prostate Cancer Patients Following Radical Prostatectomy. *Oncoimmunology* **2016**, *5*, e1221555. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
66. Goltz, D.; Gevensleben, H.; Dietrich, J.; Schroeck, F.; de Vos, L.; Droege, F.; Kristiansen, G.; Schroeck, A.; Landsberg, J.; Bootz, F.; et al. PDCD1 (PD-1) Promoter Methylation Predicts Outcome in Head and Neck Squamous Cell Carcinoma Patients. *Oncotarget* **2017**, *8*, 41011–41020. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
67. Goltz, D.; Gevensleben, H.; Vogt, T.J.; Dietrich, J.; Golletz, C.; Bootz, F.; Kristiansen, G.; Landsberg, J.; Dietrich, D. CTLA4 Methylation Predicts Response to Anti-PD-1 and Anti-CTLA-4 Immunotherapy in Melanoma Patients. *JCI Insight.* **2018**, *3*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
68. Wu, X.; Lv, D.; Cai, C.; Zhao, Z.; Wang, M.; Chen, W.; Liu, Y. A TP53-Associated Immune Prognostic Signature for the Prediction of Overall Survival and Therapeutic Responses in Muscle-Invasive Bladder Cancer. *Front. Immunol.* **2020**, *11*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
69. Clifford, R.L.; Fishbane, N.; Patel, J.; MacIsaac, J.L.; McEwen, L.M.; Fisher, A.J.; Brandsma, C.; Nair, P.; Kobor, M.S.; Hackett, T.L.; et al. Altered DNA Methylation is Associated with Aberrant Gene Expression in Parenchymal but not Airway Fibroblasts Isolated from Individuals with COPD. *Clin. Epigenetics* **2018**, *10*. [[Google Scholar](#)] [[CrossRef](#)]
70. Klümper, N.; Ralsler, D.J.; Bawden, E.G.; Landsberg, J.; Zarbl, R.; Kristiansen, G.; Toma, M.; Ritter, M.; Hölzel, M.; Ellinger, J.; et al. LAG3 (LAG-3, CD223) DNA Methylation Correlates with LAG3 Expression by Tumor and Immune Cells, Immune Cell Infiltration, and Overall Survival in Clear Cell Renal Cell Carcinoma. *J. Immunother. Cancer* **2020**, *8*. [[Google Scholar](#)] [[CrossRef](#)]
71. Querfeld, C.; Wu, X.; Sanchez, J.F.; Palmer, J.M.; Motevalli, A.; Zain, J.; Rosen, S.T. The miRNA Profile of Cutaneous T Cell Lymphoma Correlates with the Dysfunctional Immunophenotype of the Disease. *Blood* **2016**, *128*, 4132. [[Google Scholar](#)] [[CrossRef](#)]
72. Laino, A.S.; Betts, B.C.; Veerapathran, A.; Dolgalev, I.; Sarnaik, A.; Quayle, S.N.; Jones, S.S.; Weber, J.S.; Woods, D.M. HDAC6 Selective Inhibition of Melanoma Patient T-Cells Augments Anti-Tumor Characteristics. *J. Immunother. Cancer* **2019**, *7*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
73. Okamura, T.; Fujio, K.; Shibuya, M.; Sumitomo, S.; Shoda, H.; Sakaguchi, S.; Yamamoto, K. CD4+CD25-LAG3+ Regulatory T Cells Controlled by the Transcription Factor Egr-2. *Proc. Natl. Acad. Sci USA* **2009**, *106*, 13974–13979. [[Google Scholar](#)] [[CrossRef](#)]
74. Huang, K.; Pang, T.; Tong, C.; Chen, H.; Nie, Y.; Wu, J.; Zhang, Y.; Chen, G.; Zhou, W.; Yang, D.; et al. Integrative Expression and Prognosis Analysis of DHX37 in Human Cancers by Data Mining. *Biomed. Res. Int.* **2021**, *2021*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
75. Zeng, Z.; Wei, F.; Ren, X. Exhausted T Cells and Epigenetic Status. *Cancer Biology and Medicine. Cancer Biol. Med.* **2020**, *17*, 923–936. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
76. Deng, W.W.; Mao, L.; Yu, G.T.; Bu, L.L.; Ma, S.R.; Liu, B.; Gutkind, J.S.; Kulkarni, A.B.; Zhang, W.; Sun, Z.; et al. LAG-3 Confers Poor Prognosis and Its Blockade Reshapes Antitumor Response in Head And Neck Squamous Cell Carcinoma. *Oncoimmunology* **2016**, *5*, 1–14. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]

77. Marcq, E.; De Waele, J.; Van Audenaerde, J.; Lion, E.; Santermans, E.; Hens, N.; Pauwels, P.; Meerbeeck, J.P.V.; Smits, E.L.J. Abundant Expression Of TIM-3, LAG-3, PD-1 and PD-L1 As Immunotherapy Checkpoint Targets in Effusions of Mesothelioma Patients. *Oncotarget* **2017**, *8*, 89722–89735. [[Google Scholar](#)] [[CrossRef](#)]
78. Burugu, S.; Gao, D.; Leung, S.; Chia, S.K.; Nielsen, T.O. LAG-3+ Tumor Infiltrating Lymphocytes in Breast Cancer: Clinical Correlates and Association with PD-1/PD-L1 + Tumors. *Ann. Oncol.* **2017**, *28*, 2977–2984. [[Google Scholar](#)] [[CrossRef](#)]
79. Yanik, E.L.; Kaunitz, G.J.; Cottrell, T.R.; Succaria, F.; McMiller, T.L.; Ascierto, M.L.; Esandrio, J.; Xu, H.; Ogurtsova, A.; Cornish, T.; et al. Association of HIV Status with Local Immune Response to Anal Squamous Cell Carcinoma: Implications for Immunotherapy. *JAMA Oncol.* **2017**, *3*, 974–978. [[Google Scholar](#)] [[CrossRef](#)]
80. Saka, D.; Gökalp, M.; Piyade, B.; Cevik, N.C.; Sever, E.; Unutmaz, D.; Ceyhan, G.O.; Demir, I.E.; Asimgil, H. Mechanisms of T-Cell Exhaustion in Pancreatic Cancer. *Cancers* **2020**, *12*, 2274. [[Google Scholar](#)] [[CrossRef](#)]
81. Wuerdemann, N.; Pütz, K.; Eckel, H.; Jain, R.; Wittekindt, C.; Huebbers, C.U.; Sharma, S.J.; Langer, C.; Gattenlöhner, S.; Büttner, R.; et al. LAG-3, TIM-3 and Vista Expression on Tumor-Infiltrating Lymphocytes in Oropharyngeal Squamous Cell Carcinoma-Potential Biomarkers for Targeted Therapy Concepts. *Int. J. Mol. Sci.* **2021**, *22*, 1–17. [[Google Scholar](#)]
82. Shapiro, M.; Herishanu, Y.; Katz, B.Z.; Dezurella, N.; Sun, C.; Kay, S.; Polliack, A.; Avivi, I.; Wiestner, A.; Perry, C. Lymphocyte Activation Gene 3: A Novel Therapeutic Target in Chronic Lymphocytic Leukemia. *Haematol.* **2017**, *102*, 874–882. [[Google Scholar](#)] [[CrossRef](#)]
83. Chen, J.; Chen, Z. The Effect of Immune Microenvironment on the Progression and Prognosis of Colorectal Cancer. *Med. Oncol.* **2014**, *31*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
84. Li, F.J.; Zhang, Y.; Jin, G.X.; Yao, L.; Wu, D.Q. Expression Of LAG-3 Is Coincident with the Impaired Effector Function of HBV-Specific CD8 + T Cell in HCC Patients. *Immunol. Lett.* **2013**, *150*, 116–122. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
85. Giraldo, N.A.; Becht, E.; Pagès, F.; Skliris, G.; Verkarre, V.; Vano, Y.; Mejean, A.; Saint-Aubert, N.; Lacroix, L.; Natario, I.; et al. Orchestration and Prognostic Significance of Immune Checkpoints in the Microenvironment of Primary and Metastatic Renal Cell Cancer. *Clin. Cancer Res.* **2015**, *21*, 3031–3040. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
86. Takaya, S.; Saito, H.; Ikeguchi, M. Upregulation of Immune Checkpoint Molecules, PD-1 and LAG-3, On CD4 + and CD8+ T Cells After Gastric Cancer Surgery. *Yonago Acta. Med.* **2015**, *58*, 39–44. [[Google Scholar](#)] [[PubMed](#)]
87. Yang, Z.Z.; Kim, H.J.; Villasboas, J.C.; Chen, Y.P.; Price-Troska, T.P.; Jalali, S.; Wilson, M.; Novak, A.J.; Ansell, S.M. Expression of LAG-3 Defines Exhaustion of Intratumoral PD-1+ T Cells and Correlates with Poor Outcome in Follicular Lymphoma. *Oncotarget* **2017**, *8*, 61425–61439. [[Google Scholar](#)] [[CrossRef](#)]
88. Saleh, R.R.; Peinado, P.; Fuentes-Antrás, J.; Pérez-Segura, P.; Pandiella, A.; Amir, E.; Ocaña, A. Prognostic Value of Lymphocyte-Activation Gene 3 (LAG3) in Cancer: A Meta-Analysis. *Front. Oncol.* **2019**, *9*, 1040. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
89. Wang, Y.; Dong, T.; Xuan, Q.; Zhao, H.; Qin, L.; Zhang, Q. Lymphocyte-Activation Gene-3 Expression and Prognostic Value in Neoadjuvant-Treated Triple-Negative Breast Cancer. *J. Breast Cancer* **2018**, *21*, 124–133. [[Google Scholar](#)] [[CrossRef](#)]
90. Datar, I.; Sanmamed, M.F.; Wang, J.; Henick, B.S.; Choi, J.; Badri, T.; Dong, W.; Mani, N.; Toki, M.; Mejías, L.D.; et al. Expression Analysis and Significance of PD-1, LAG-3, and TIM-3 in Human Non-Small Cell Lung Cancer Using Spatially Resolved and Multiparametric Single-Cell Analysis. *Clin. Cancer Res.* **2019**, *25*, 4663–4673. [[Google Scholar](#)] [[CrossRef](#)]

91. Sobottka, B.; Moch, H.; Varga, Z. Differential PD-1/LAG-3 Expression and Immune Phenotypes in Metastatic Sites of Breast Cancer. *Breast Cancer Res.* **2021**, *23*. [[Google Scholar](#)] [[CrossRef](#)]
92. Zhang, X.; Zhao, H.; Shi, X.; Jia, X.; Yang, Y. Identification and Validation of an Immune-Related Gene Signature Predictive of Overall Survival in Colon Cancer. *Aging* **2020**, *12*, 26095–26120. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
93. Lichtenegger, F.S.; Rothe, M.; Schnorfeil, F.M.; Deiser, K.; Krupka, C.; Augsberger, C.; Schlüter, M.; Neitz, J.; Subklewe, M. Targeting LAG-3 and PD-1 to Enhance T Cell Activation by Antigen-Presenting Cells. *Front. Immunol.* **2018**, *9*, 1–12. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
94. Jing, W.; Gershan, J.A.; Weber, J.; Tlomak, D.; McOlash, L.; Sabatos-Peyton, C.; Johnson, B.D. Combined Immune Checkpoint Protein Blockade and Low Dose Whole Body Irradiation as Immunotherapy for Myeloma. *J. Immunother. Cancer* **2015**, *3*. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
95. Haudebourg, T.; Dugast, A.S.; Coulon, F.; Usual, C.; Triebel, F.; Vanhove, B. Depletion of LAG-3 Positive Cells in Cardiac Allograft Reveals Their Role in Rejection and Tolerance. *Transplant* **2007**, *84*, 1500–1506. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
96. Zhu, Z.; Ye, J.; Ma, Y.; Hua, P.; Huang, Y.; Fu, X.; Lie, D.; Yuan, M.; Xiag, Z. Function of T Regulatory Type 1 Cells Is Down-Regulated and Is Associated with the Clinical Presentation of Coronary Artery Disease. *Hum. Immunol.* **2018**, *79*, 564–570. [[Google Scholar](#)] [[CrossRef](#)]
97. Golden, D.; Kolmakova, A.; Sura, S.; Vella, A.T.; Manichaikul, A.; Wang, X.-Q.; Bielinski, S.J.; Taylor, K.D.; Chen, Y.I.; Rich, S.S.; et al. Lymphocyte Activation Gene 3 and Coronary Artery Disease. *JCI Insight* **2016**, *1*, 88628. [[Google Scholar](#)] [[CrossRef](#)]
98. Rodriguez, A. High HDL-Cholesterol Paradox: SCARB1-LAG3-HDL Axis. *Curr. Atheroscler. Rep.* **2021**, *23*. [[Google Scholar](#)] [[CrossRef](#)]
99. Bauché, D.; Joyce-Shaikh, B.; Jain, R.; Grein, J.; Ku, K.S.; Blumenschein, W.M.; Ganal-Vonarburg, S.C.; Wilson, D.C.; McClanahan, T.K.; Malefyt, R.D.W.; et al. LAG3 + Regulatory T Cells Restrain Interleukin-23-Producing CX3CR1 + Gut-Resident Macrophages during Group 3 Innate Lymphoid Cell-Driven Colitis. *Immunity* **2018**, *49*, 342–352.e5. [[Google Scholar](#)] [[CrossRef](#)]
100. Zhang, Z.; Duvefelt, K.; Svensson, F.; Masterman, T.; Jonasdottir, G.; Salter, H.; Emahazion, T.; Hellgren, D.; Falk, G.; Olsson, T.; et al. Two Genes Encoding Immune-Regulatory Molecules (LAG3 And IL7R) Confer Susceptibility to Multiple Sclerosis. *Genes Immun.* **2005**, *6*, 145–152. [[Google Scholar](#)] [[CrossRef](#)]
101. Bettini, M.; Szymczak-Workman, A.L.; Forbes, K.; Castellaw, A.H.; Selby, M.; Pan, X.; Drake, C.G.; Korman, A.J.; Dario, A.; Vignali, A. Cutting Edge: Accelerated Autoimmune Diabetes in the Absence of LAG-3. *J. Immunol.* **2011**, *187*, 3493–3498. [[Google Scholar](#)] [[CrossRef](#)]
102. Delmastro, M.M.; Styche, A.J.; Trucco, M.M.; Workman, C.J.; Vignali, D.A.A.; Piganelli, J.D. Modulation of Redox Balance Leaves Murine Diabetogenic TH1 T Cells “LAG-3-Ing” Behind. *Diabetes* **2012**, *61*, 1760–1768. [[Google Scholar](#)] [[CrossRef](#)]
103. Doe, H.T.; Kimura, D.; Miyakoda, M.; Kimura, K.; Akbari, M.; Yui, K. Expression Of PD-1/LAG-3 and Cytokine Production by CD4 + T Cells During Infection with Plasmodium Parasites. *Microbiol. Immunol.* **2016**, *60*, 21–31. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
104. Phillips, B.L.; Mehra, S.; Ahsan, M.H.; Selman, M.; Khader, S.A.; Kaushal, D. LAG3 Expression in Active Mycobacterium Tuberculosis Infections. *Am. J. Pathol.* **2015**, *185*, 820–833. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
105. Graydon, C.G.; Balasko, A.L.; Fowke, K.R. Roles, Function and Relevance of LAG3 in HIV Infection. *PLoS Pathog.* **2019**, *15*, e1007429. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]

106. Jochems, S.P.; Jacquelin, B.; Tchitchek, N.; Busato, F.; Pichon, F.; Huot, N.; Liu, Y.; Ploquin, M.J.; Roché, E.; Cheynier, R.; et al. DNA Methylation Changes in Metabolic and Immune-Regulatory Pathways in Blood and Lymph Node CD4 + T Cells in Response to SIV Infections. *Clin. Epigenetics*. **2020**, *12*. [[Google Scholar](#)] [[CrossRef](#)]
107. McLane, L.M.; Abdel-Hakeem, M.S.; Wherry, E.J. *CD8 T Cell Exhaustion During Chronic Viral Infection and Cancer*; Annual Review of Immunology; Annual Reviews Inc.: Palo Alto, CA, USA, 2019; Volume 37, pp. 457–495. [[Google Scholar](#)]
108. Anderson, A.C.; Joller, N.; Kuchroo, V.K. Lag-3, Tim-3, and TIGIT: Co-inhibitory Receptors with Specialized Functions in Immune Regulation. *Immun. Cell Press* **2016**, *44*, 989–1004. [[Google Scholar](#)] [[CrossRef](#)]
109. Richter, K.; Agnellini, P.; Oxenius, A. On the Role of the Inhibitory Receptor LAG-3 in Acute and Chronic LCMV Infection. *Int. Immunol.* **2009**, *22*, 13–23. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
110. Roy, S.; Coulon, P.-G.; Prakash, S.; Srivastava, R.; Geertsema, R.; Dhanushkodi, N.; Lam, C.; Nguyen, V.; Gorospe, E.; Nguyen, A.M.; et al. Blockade of PD-1 and LAG-3 Immune Checkpoints Combined with Vaccination Restores the Function of Antiviral Tissue-Resident CD8 + T RM Cells and Reduces Ocular Herpes Simplex Infection and Disease in HLA Transgenic Rabbits. *J. Virol.* **2009**, *93*. [[Google Scholar](#)] [[CrossRef](#)]
111. Roy, S.; Coulon, P.G.; Srivastava, R.; Vahed, H.; Kim, G.J.; Walia, S.S.; Yamada, T.; Fouladi, M.A.; Ly, V.T.; BenMohamed, L. Blockade of LAG-3 Immune Checkpoint Combined With Therapeutic Vaccination Restore the Function of Tissue-Resident Anti-Viral CD8 + T Cells and Protect Against Recurrent Ocular Herpes Simplex Infection and Disease. *Front. Immunol.* **2018**, *9*, 2922. [[Google Scholar](#)] [[CrossRef](#)]
112. Liu, Y.; Source, S.; Nuvolone, M.; Domange, J.; Aguzzi, A. Lymphocyte Activation Gene 3 (Lag3) Expression Is Increased in Prion Infections but Does Not Modify Disease Progression. *Sci. Rep.* **2018**, *8*. [[Google Scholar](#)] [[CrossRef](#)]
113. Martins, F.; Sofiya, L.; Sykietis, G.P.; Lamine, F.; Maillard, M.; Shabafrouz, K.; Shabafrouz, K.; Ribi, C.; Cairolì, A.; Guex-Crosier, Y.; et al. Adverse effects of immune—checkpoint inhibitors: Epidemiology, management and surveillance. *Nat. Rev. Clin. Oncol.* **2019**, *16*, 563–580. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
114. Nishino, M.; Ramaiya, N.H.; Hatabu, H.; Hodi, F.S. Monitoring Immune-Checkpoint Blockade: Response Evaluation and Biomarker Development. *Nat. Rev. Clin. Oncol.* **2018**, *14*, 655–668. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
115. Prasad, V.; De Jesús, K.; Mailankody, S. The High Price of Anticancer Drugs: Origins, Implications, Barriers, Solutions. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 381–390. [[Google Scholar](#)] [[CrossRef](#)]
116. Topalian, S.L.; Weiner, G.J.; Pardoll, D.M. Cancer Immunotherapy Comes of Age. *J. Clin. Oncol.* **2011**, *29*, 4828. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
117. Topalian, S.L.; Drake, C.G.; Pardoll, D.M. Perspective Immune Checkpoint Blockade: A Common Denominator Approach to Cancer Therapy. *Cancer Cell.* **2015**, *27*, 450–461. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
118. Chocarro de Erauso, L.; Zuazo, M.; Arasanz, H.; Bocanegra, A.; Hernandez, C.; Fernandez, G.; Garcia-Granda, M.J.; Blanco, E.; Vera, R.; Kochan, G.; et al. Resistance to PD-L1/PD-1 Blockade Immunotherapy. A Tumor-Intrinsic or Tumor-Extrinsic Phenomenon? *Front. Pharmacol.* **2020**, *11*. [[Google Scholar](#)] [[CrossRef](#)]
119. Zuazo, M.; Arasanz, H.; Bocanegra, A.; Fernandez, G.; Chocarro, L.; Vera, R.; Kochan, G.; Escors, D. Systemic CD4 Immunity as a Key Contributor to PD-L1/PD-1 Blockade Immunotherapy Efficacy. *Front. Immunol.* **2020**, *11*. [[Google Scholar](#)] [[CrossRef](#)]
120. Zuazo, M.; Arasanz, H.; Bocanegra, A.; Chocarro, L.; Vera, R.; Escors, D.; Kagamu, H.; Kochan, G. Systemic CD4 Immunity: A Powerful Clinical Biomarker for PD-L1/PD-1 Immunotherapy. *EMBO Mol. Med.* **2020**, *12*, e12706. [[Google Scholar](#)] [[CrossRef](#)]

121. Bocanegra, A.; Fernandez-Hinojal, G.; Zuazo-Ibarra, M.; Arasan, H.; Garcia-Granda, M.J.; Hernandez, C.; Ibañez, M.; Hernandez-Marin, B.; Martinez-Aguillo, M.; Lecumberri, M.J.; et al. PD-L1 Expression in Systemic Immune Cell Populations as a Potential Predictive Biomarker of Responses to PD-L1/PD-1 Blockade Therapy in Lung Cancer. *Int. J. Mol. Sci.* **2019**, *20*, 1631. [[Google Scholar](#)] [[CrossRef](#)]
122. Rotte, A.; Jin, J.Y.; Lemaire, V. Mechanistic overview of immune checkpoints to support the rational design of their combinations in cancer immunotherapy. *Ann. Oncol.* **2018**, *29*, 71–83. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
123. Legg, J.W.; McGuinness, B.; Arasan, H.; Bocanegra, A.; Bartlett, P.; Benedetti, G.; Birkett, N.; Cox, C.; De Juan, E.; Enever, C.; et al. Abstract 930: CB213: A Half-Life Extended Bispecific Humabody V H Delivering Dual Checkpoint Blockade to Reverse the Dysfunction of LAG3 + PD-1 + Double-Positive T Cells. *Am. Assoc. Cancer Res.* **2020**. [[Google Scholar](#)] [[CrossRef](#)]
124. Chocarro L, Blanco E, Arasan H, Fernández-Rubio L, Bocanegra A, Echaide M, Garnica M, Ramos P, Fernández-Hinojal G, Vera R, Kochan G, Escors D, Clinical landscape of LAG-3 targeted therapy, *Immuno-Oncology and Technology*, <https://doi.org/10.1016/j.iotech.2022.100079>
125. Chocarro, L., Blanco, E., Zuazo, M., Arasan, H., Bocanegra, A., Fernandez-Rubio, L., Morente, P., Fernandez-Hinojal, G., Echaide, M., Garnica, M., et al. (2021). Understanding LAG-3 Signaling. *International journal of molecular sciences* *22*. <https://doi.org/10.3390/ijms22105282>

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