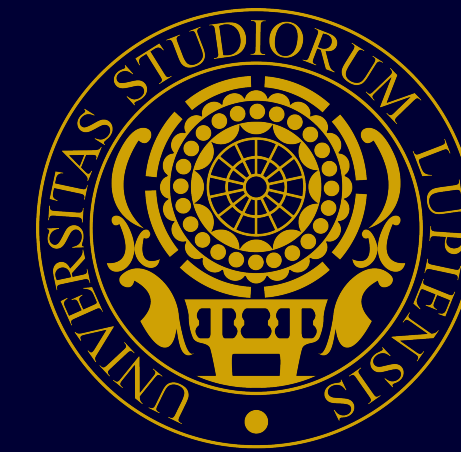


# The 2nd International Electronic Conference on Antibiotics

Drugs for Superbugs: Antibiotic Discovery, Modes of Action And Mechanisms of Resistance

Session: Repurposing & Antimicrobial Adjuvants (S4)

**ECA**  
**2022**



**UNIVERSITÀ  
DEL SALENTO**

L'Ateneo tra i due mari

---

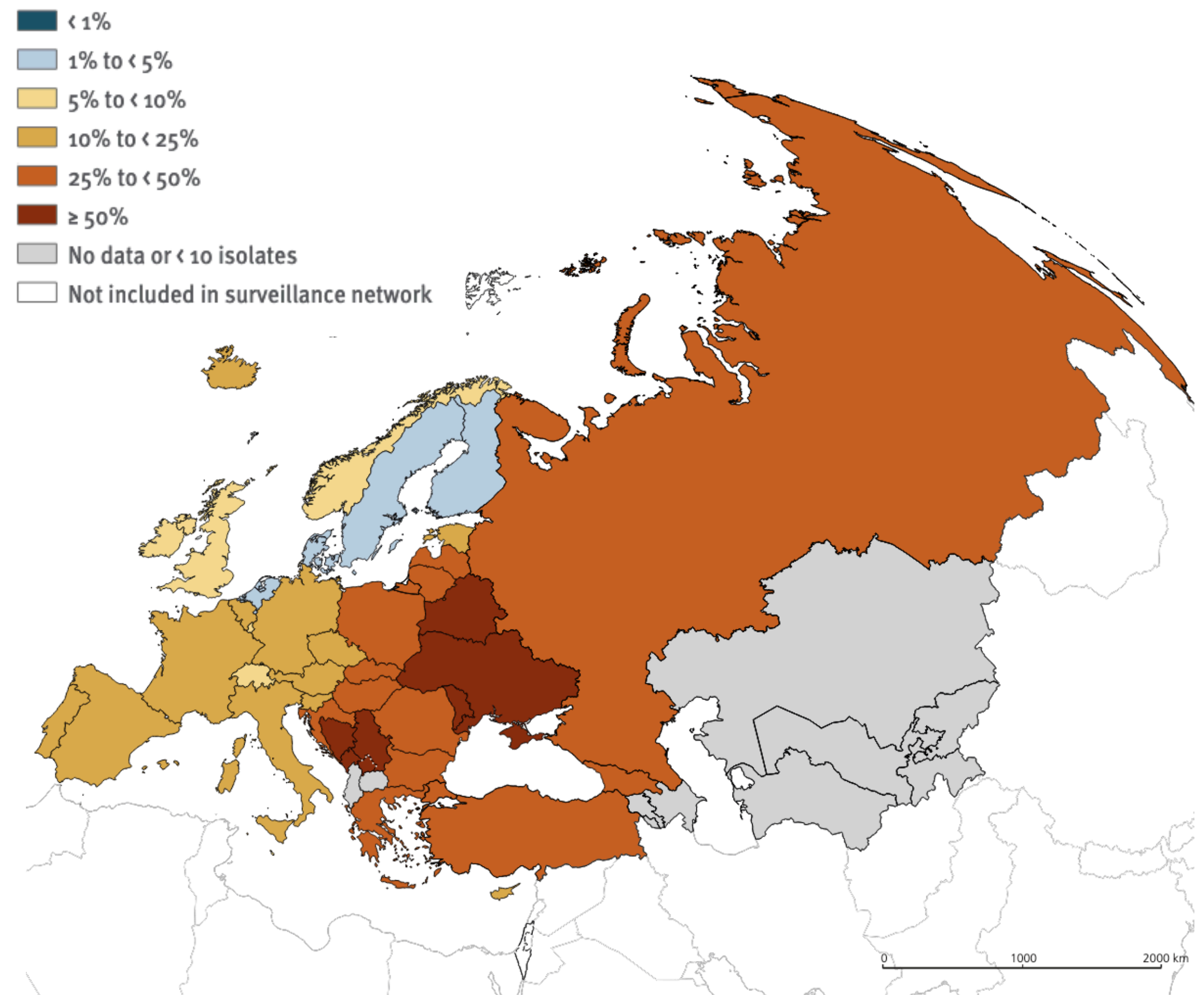
Off-target activity of spiramycin disarms *Pseudomonas aeruginosa* by  
inhibition of biofilm formation, pigment production and phenotypic  
differentiation

---

**MATTEO CALCAGNILE & PIETRO ALIFANO**

# Identikit of *Pseudomonas aeruginosa*

- Gram-negative bacterium
- one of the most common pathogens in chronic lung infection
- the third most common pathogen associated with nosocomial urinary tract infections (UTIs)
- multi-drug resistance (acquired resistance)



*P. aeruginosa*: percentage of invasive isolates with **resistance to carbapenems**, by country/area, WHO European Region, 2020 (Antimicrobial resistance surveillance in Europe 2022 - 2020 data- ECDC)

---

*Pseudomonas aeruginosa*

*in vitro* evidence:

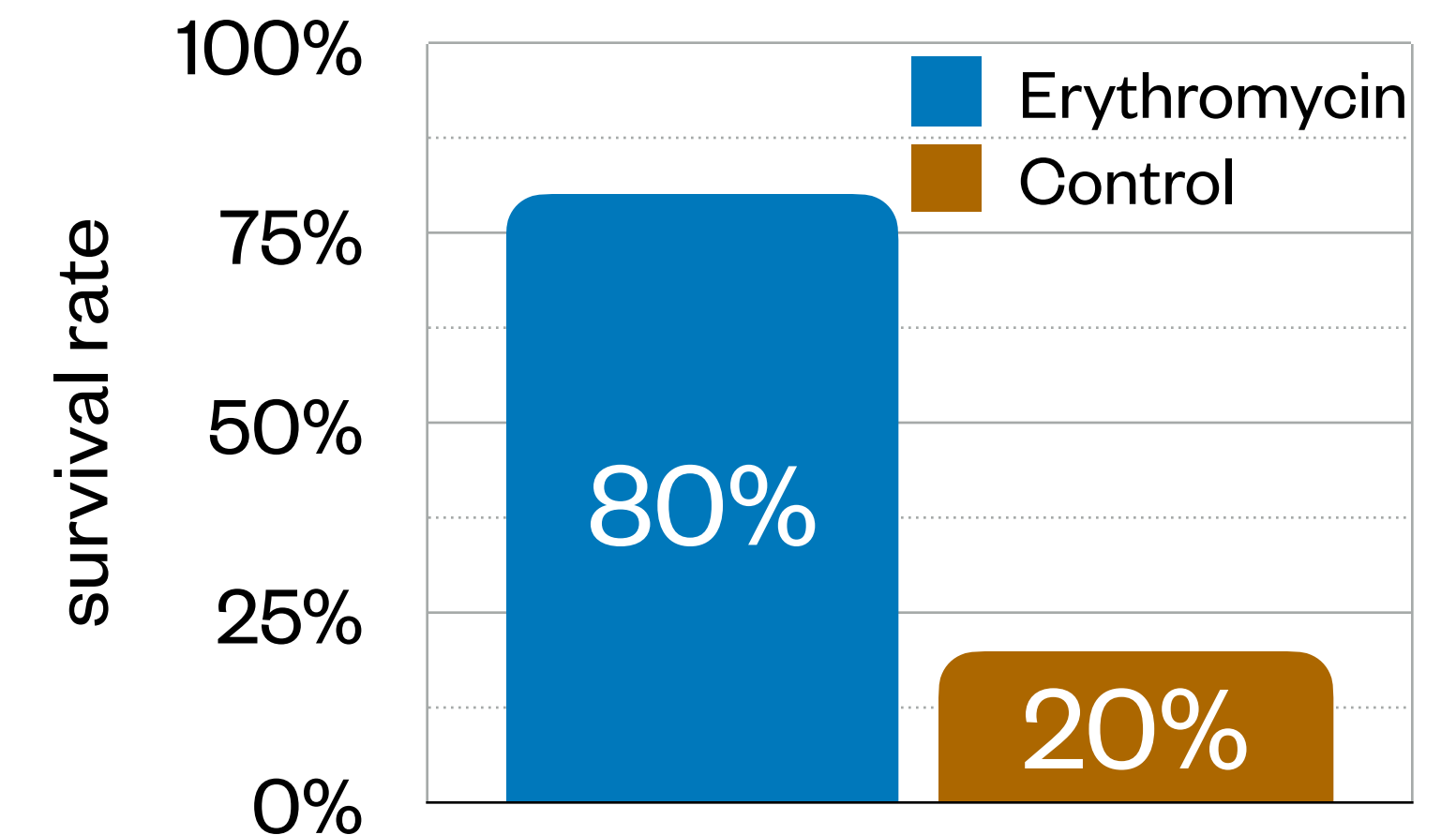
spontaneously resistance to macrolides

---

# Treatment with Macrolides of *P. aeruginosa* infection

**1** mouse model of *P. aeruginosa* bacteraemia, treatment with erythromycin

**INCREASE SURVIVAL RATE  
(HIRAKATA ET AL., 1992).**



**2** “A 4–6-month trial of azithromycin is justified in children with cystic fibrosis who do not respond to conventional treatment. The mechanism of action remains unknown.”  
(Equi et al., 2002)

Speculation on macrolides:

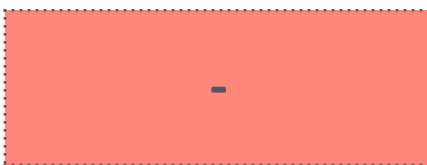
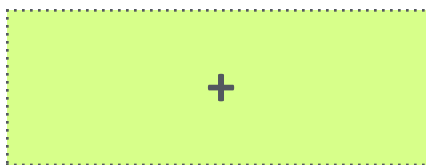
- down-modulation of the inflammatory response (host)
- down-modulation of the *P. aeruginosa* virulence.

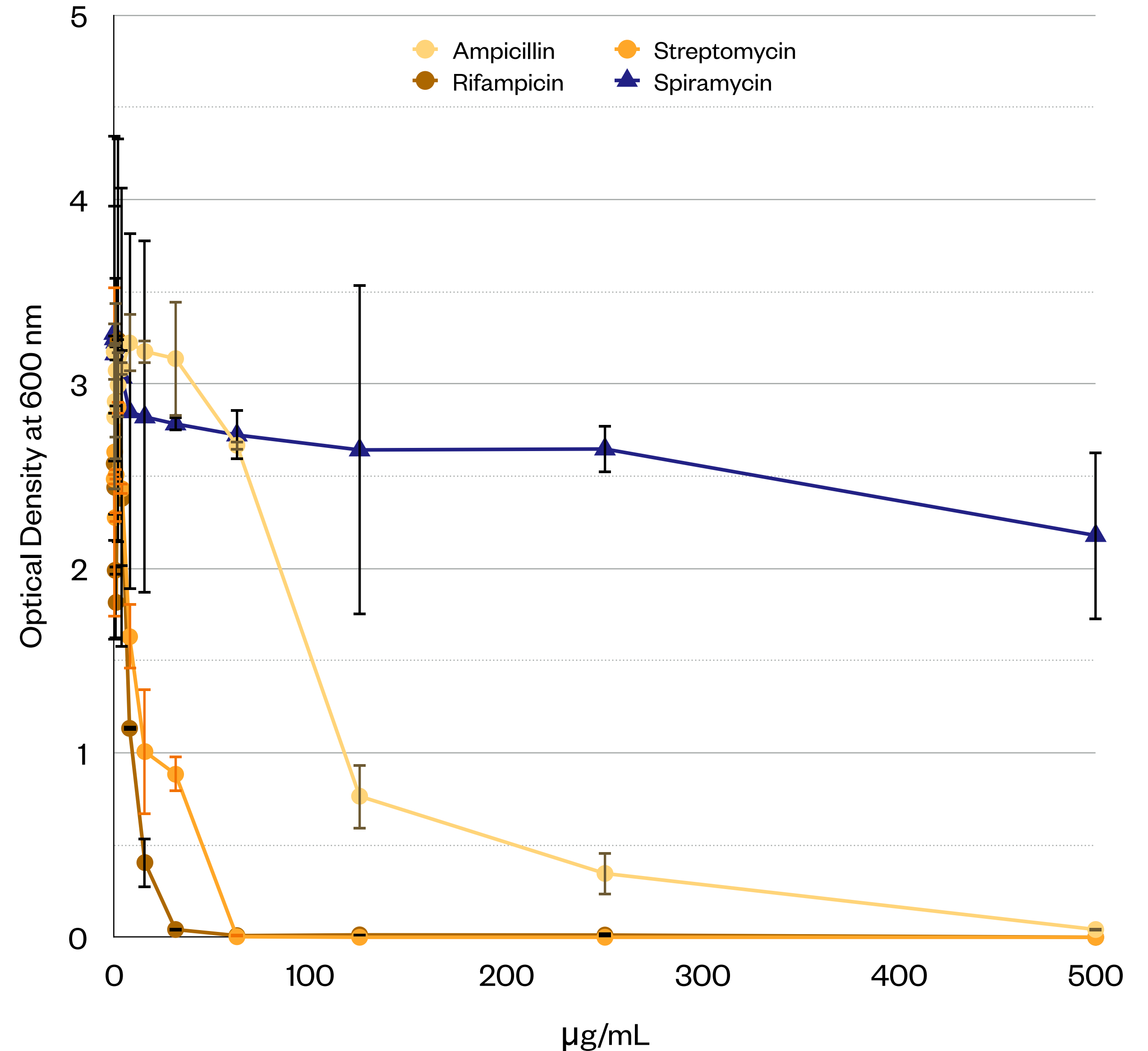
# Serendipity in drug repurposing

We studied the sensitivity of *P. aeruginosa* to antibiotics. We observed that this bacterium was resistant to spiramycin.

Usually, the bacterium is green due to the pigments (e.g., pyocyanin). With spiramycin (from 15  $\mu\text{g}/\text{mL}$ ), the strain was white but continued to grow.

$\mu\text{g}/\text{mL}$	Ampicillin	Streptomycin	Rifampicin	Spiramycin
500	-	-	-	+
250	+	-	-	+
125	+	-	-	+
62.5	+	-	-	+
31.25	+	+	-	+
15.625	+	+	+	+

 Inhibited Bacteria       Active growing Bacteria



---

# Spiramycin

- 16-membered macrolide
- produced by *Streptomyces ambofaciens*
- Effective against Gram-positive pathogens (*Staphylococcus aureus*, streptococci of groups A, B, C and D, and pneumococcus)
- It is also effective against Gram-negative pathogens (*Neisseria*, *Legionella*)
- It is an antiparasitic agent (active against *Toxoplasma spp.*)
- The antibacterial activity was associated with its ability to bind the 50S ribosomal subunit and to block the translation

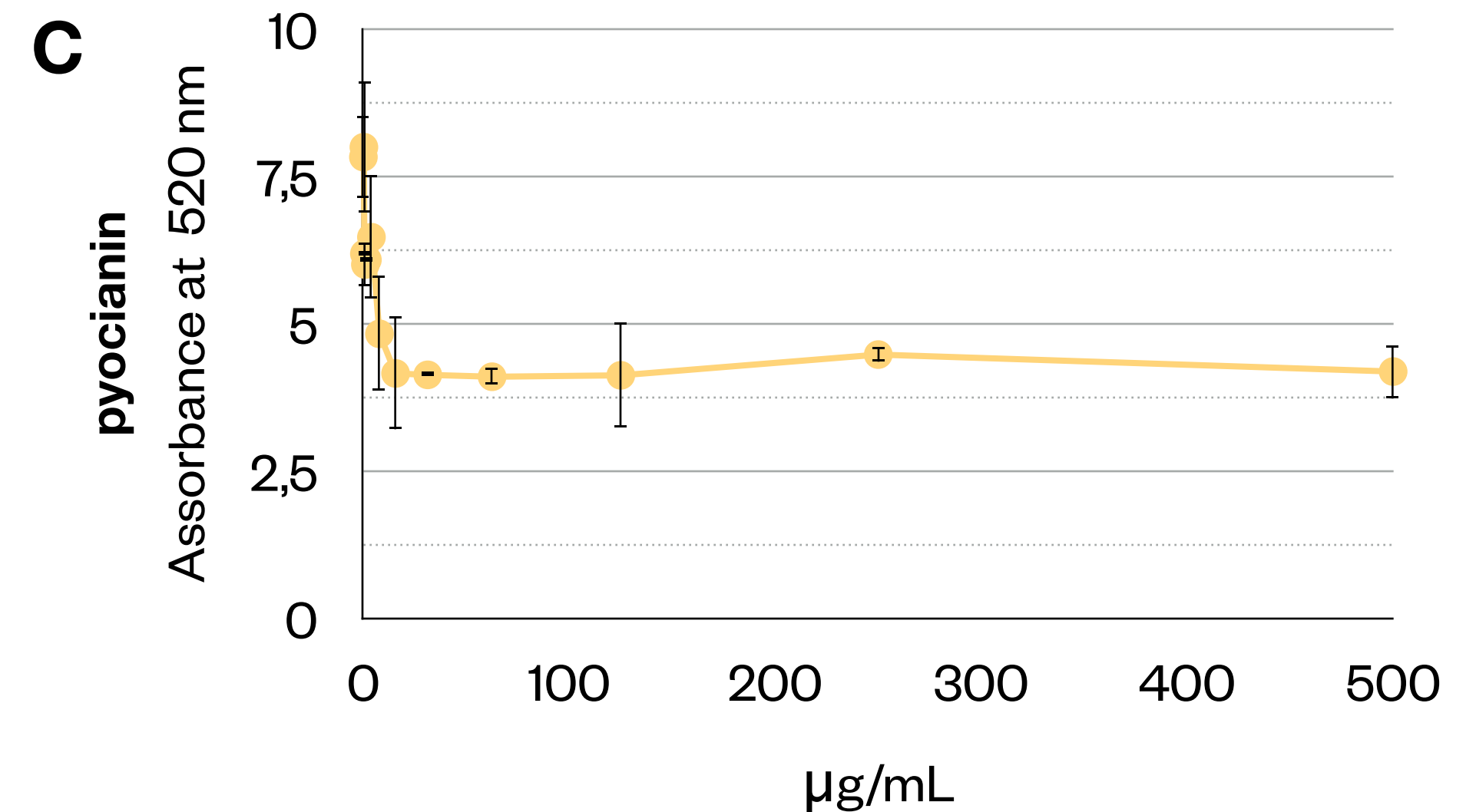
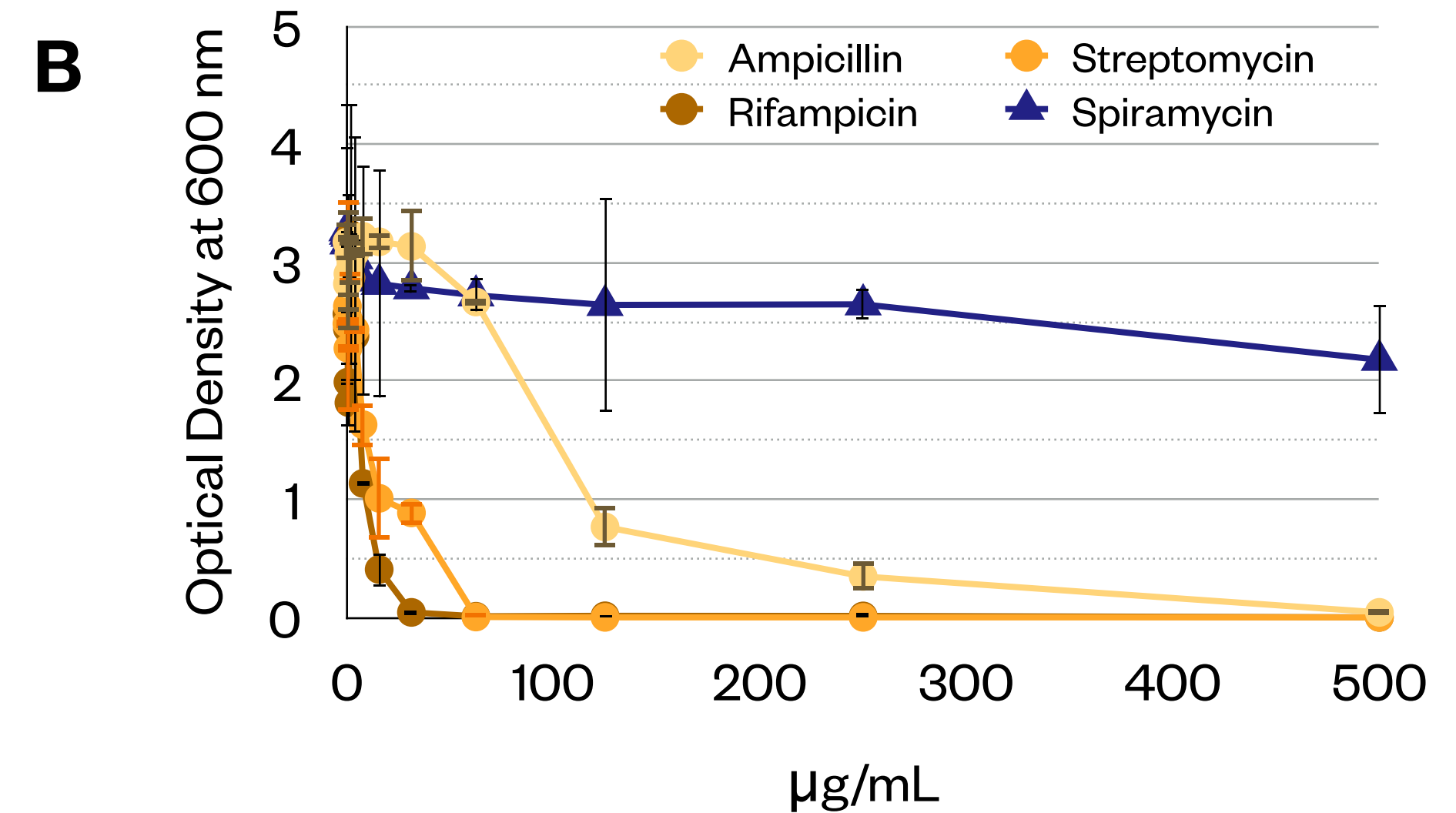
# Effect of spiramycin on Pyocyanin production

Spiramycin (from 500  $\mu\text{g}/\text{mL}$  to 1  $\mu\text{g}/\text{mL}$ ) does not inhibit the growth of *P. aeruginosa* in LB medium in both liquid and solid (Fig. A and Fig. B) cultures

## MIC (minimal inhibitory concentration)

Estimation of biomass using optical density (O.D.) show as *P. aeruginosa* is sensitive to ampicillin, Rifampicin and Streptomycin (Fig. B).

Spiramycin (from 500  $\mu\text{g}/\text{mL}$  to 30  $\mu\text{g}/\text{mL}$ ) affects the production of pigments in *P. aeruginosa* cultures (Fig. C) or solid (Fig. A).



# Effect of spiramycin on biofilm formation

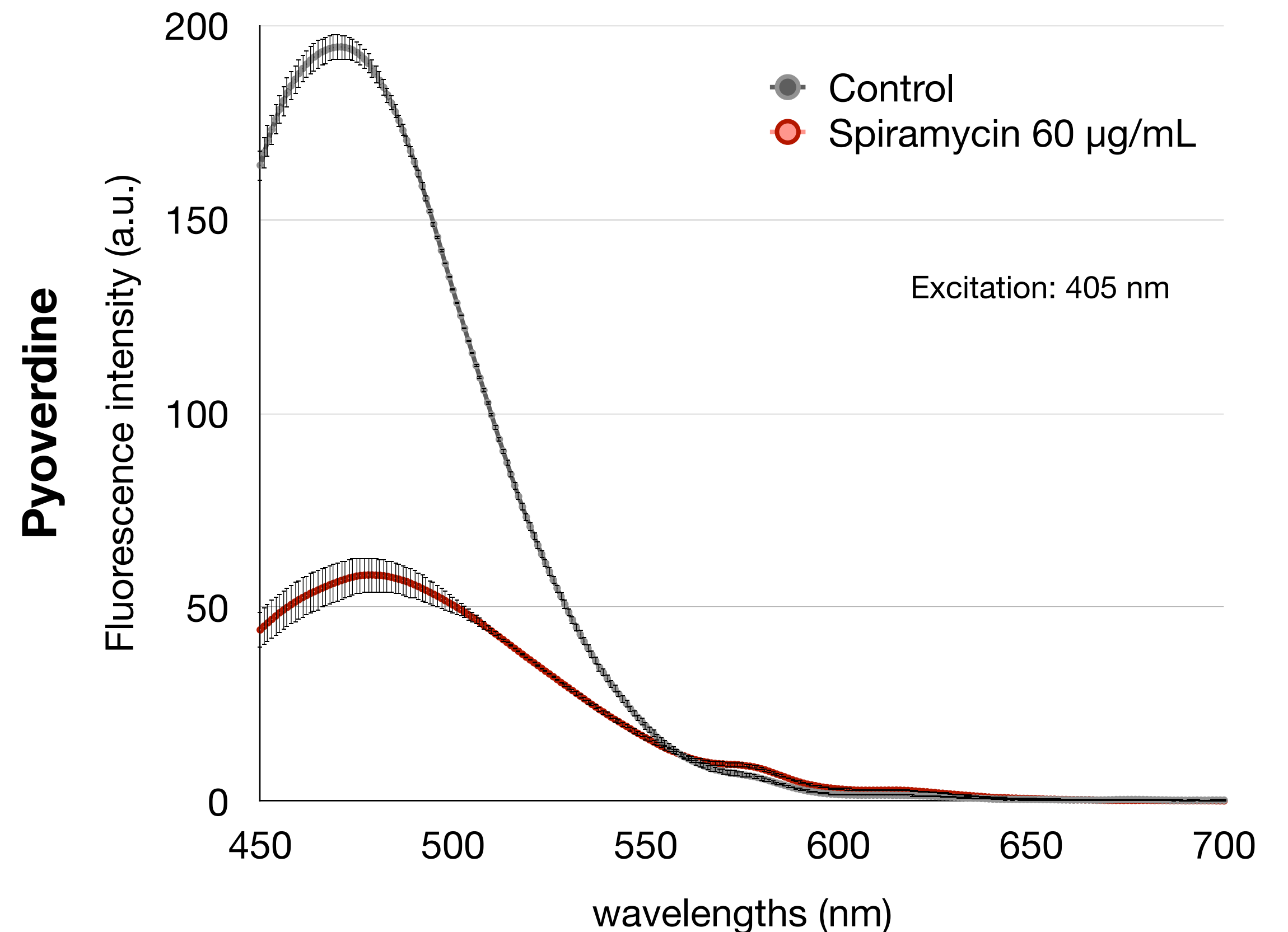
We cultured *P. aeruginosa* using hydroxyapatite dishes as supports to form a biofilm.

After 72 h we detected:

- Pigments production (Fig. A, B e C)
- Biofilm formation (Fig. D and E)
- Planktonic cell (Fig. F)

Spiramycin (60  $\mu\text{g}/\text{mL}$ ) leads to the inhibition of both biofilm and pigments (Pyocyanin & Pyoverdine)

	Units	Control	Spiramycin (60 $\mu\text{g}/\text{ml}$ )
<b>Biofilm</b>	CFU/support	$1 \times 10^6$	$5 \times 10^3$
<b>Planktonic cells</b>	Total protein ( $\mu\text{g}/\mu\text{L}$ )	1.5	2
<b>Pyocyanin</b>	Absorbance at 520 nm	4.2	9.22



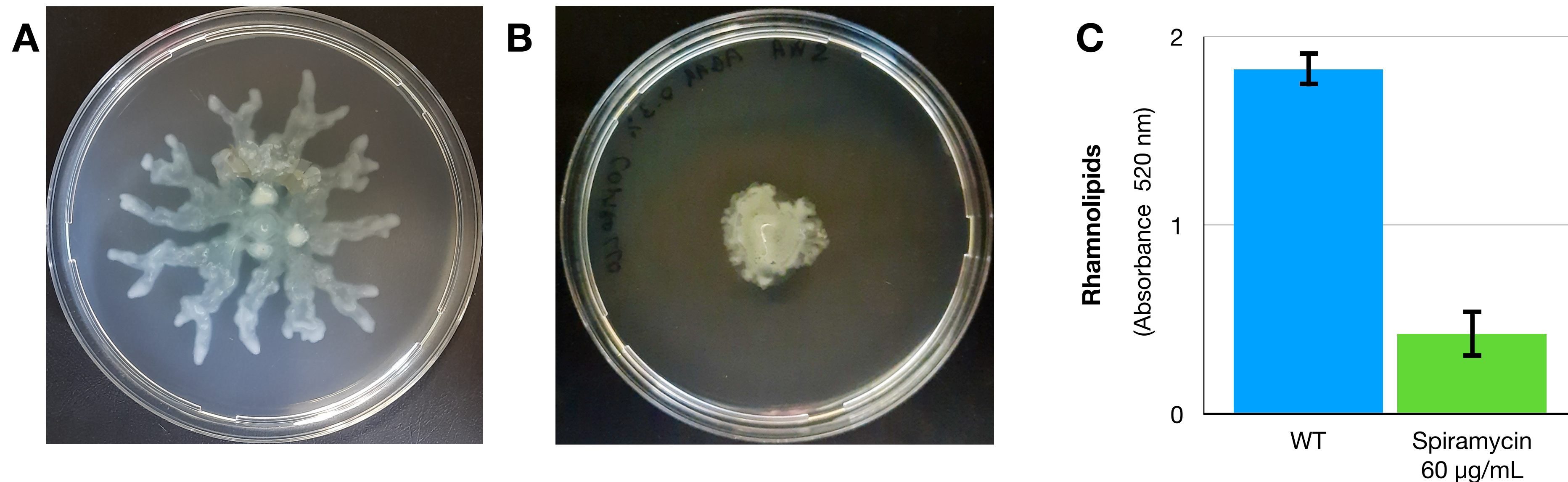


# Effect of spiramycin on motility and rhamnolipids production

We tested the ability of spiramycin to inhibit the motility of the bacterium using an appropriate minimal medium (Fig. A, control; Fig. B, spiramycin 60  $\mu\text{g}/\text{mL}$ ).

We cultured *P. aeruginosa* in 250 ml flasks filled with 50 ml of LB (72 h, 37 ° C, 180 rpm). We measured rhamnolipids using orcinol.

Spiramycin inhibits motility and reduces the amount of rhamnolipids (biosurfactants) produced



# Infection of *G. mellonella* larvae - Protocol

We used *Galleria mellonella* larvae as an in vivo infection model.

1. **Inoculum:** a bacterial suspension formed by an injection solution (10 mM MgSO<sub>4</sub>) and bacteria (O.D. at 600 nm of 0.1)
2. **Dilution:** we diluted the solution obtained 7 times (dilution 1:10).
3. **Injection:** we injected 5 µl aliquots into the larvae through the last left leg using a 10 µl Hamilton syringe.

We prepared two different samples:

- 1) control (without spiramycin);
- 2) treated. In the second case, both LB (medium of growth) and injection solution contained spiramycin (60 mg/mL).

**We incubated the larvae at 37°C for 24 hours.**

# Infection of *G. mellonella* larvae - Results



The results show that infecting the larvae with only *P. aeruginosa* mortality is about 92%.

Mortality drops to 30% when the bacterium is inoculated with spiramycin.

These results are preliminary and will be replicated in the future.

**P. AERUGINOSA INFECTION**

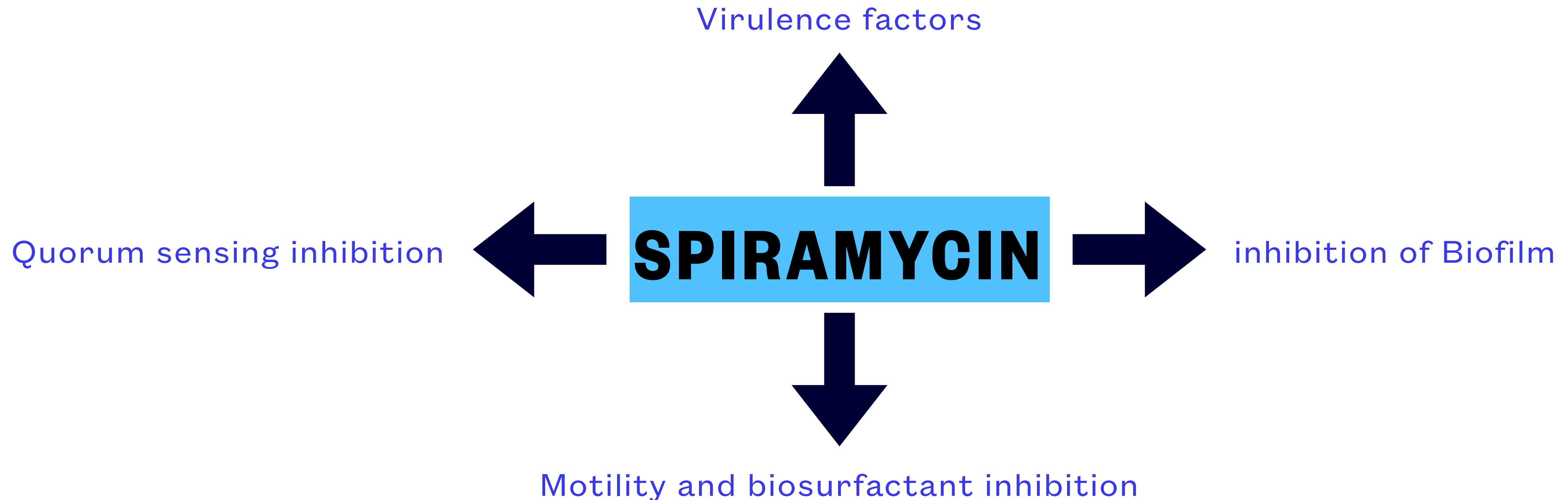


**P. AERUGINOSA INFECTION AND SPIRAMYCIN (60 mg/mL)**



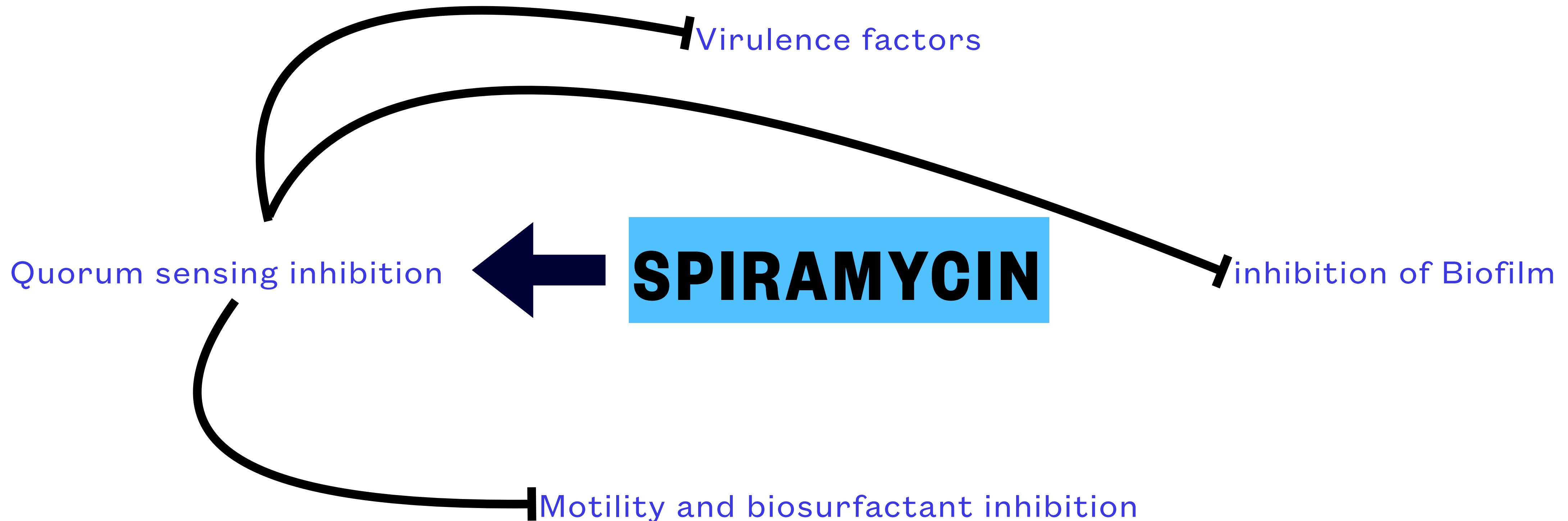
# One Hypothesis

Spiramycin binds the quorum-sensing regulators. This phenomenon leads to an inhibition of biofilm formation, motility and more.



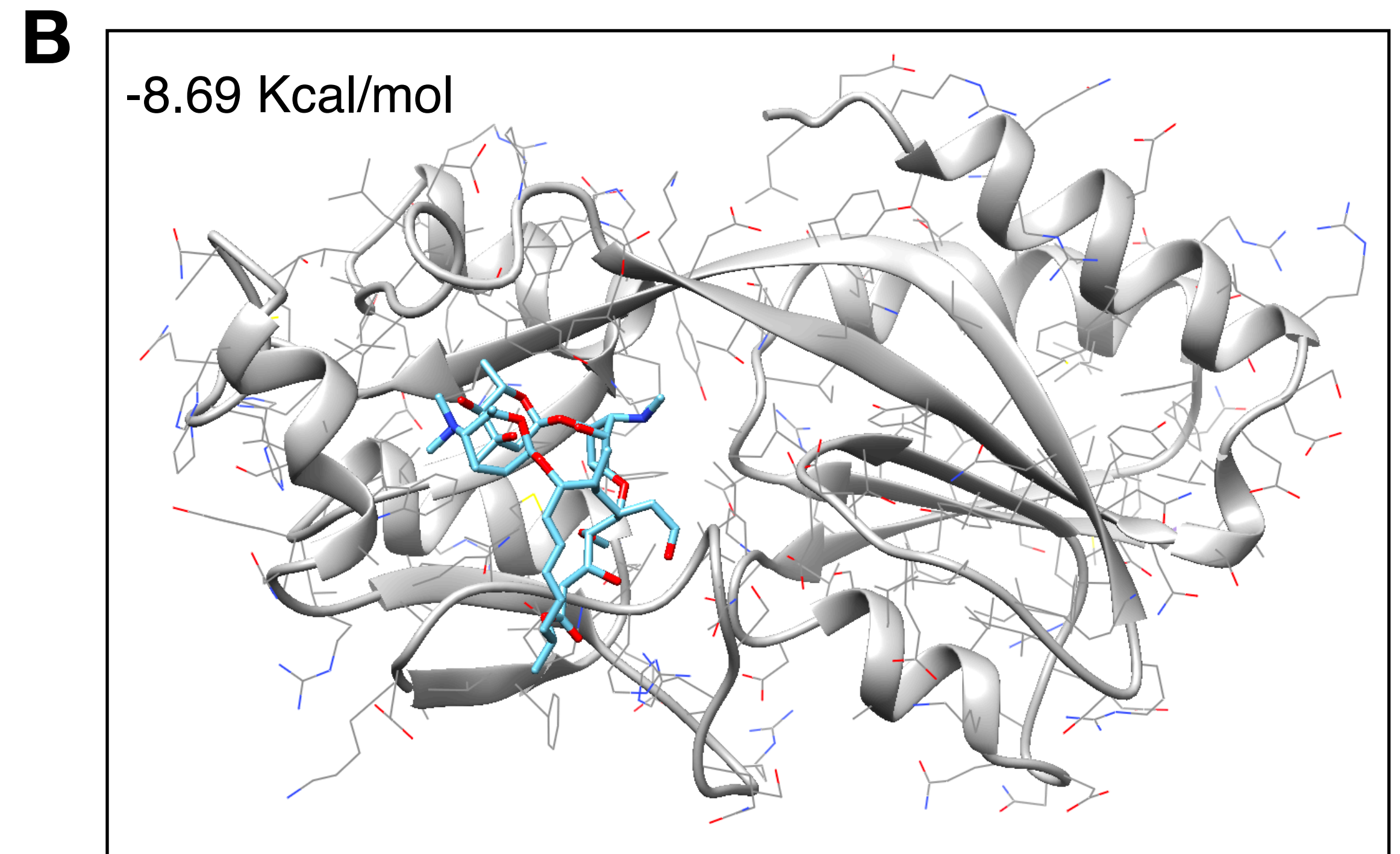
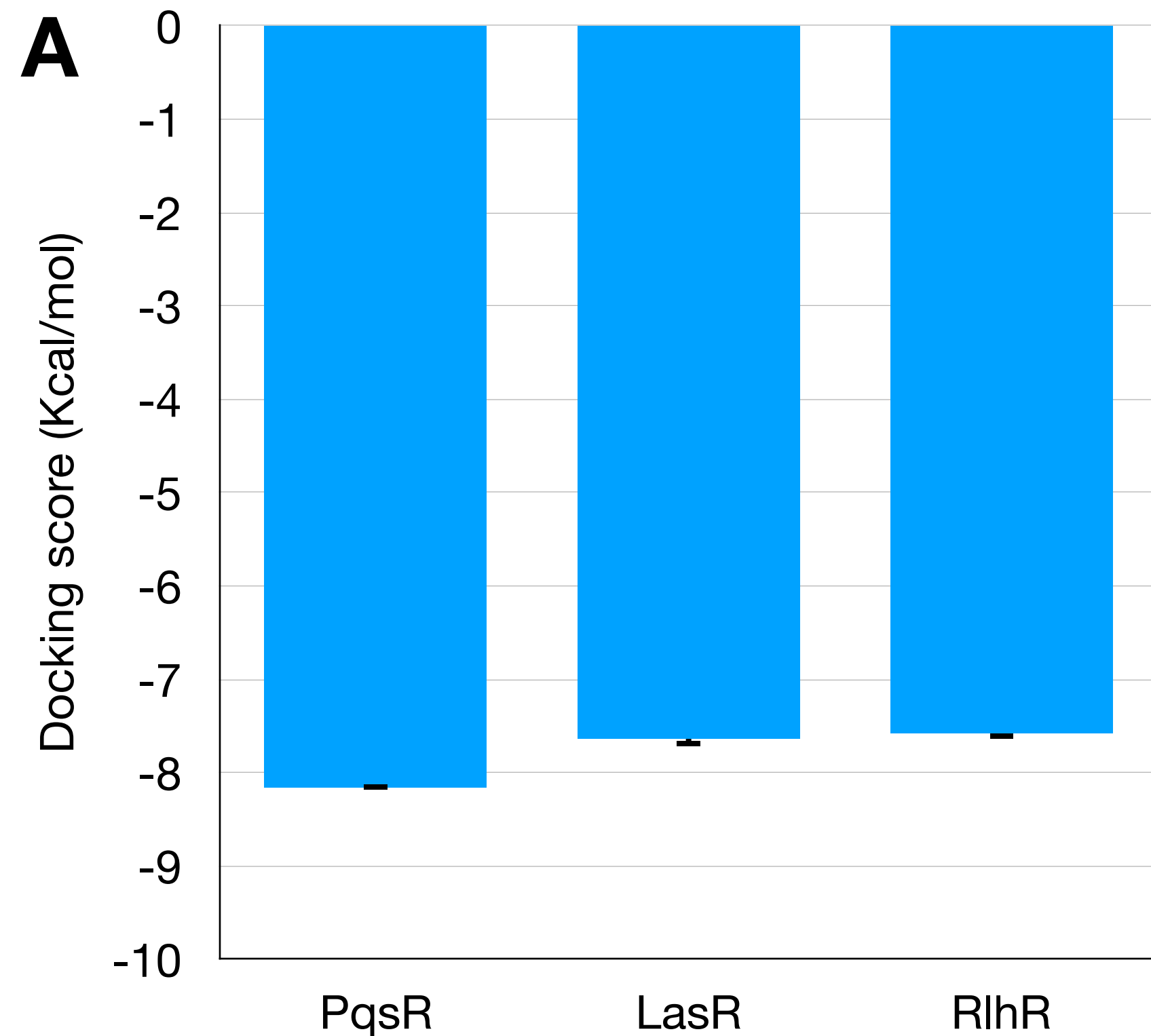
# One Hypothesis

Spiramycin binds the quorum-sensing regulators. This phenomenon leads to an inhibition of biofilm formation, motility and more.



# Docking: to test this hypothesis we performed *in-silico* analysis

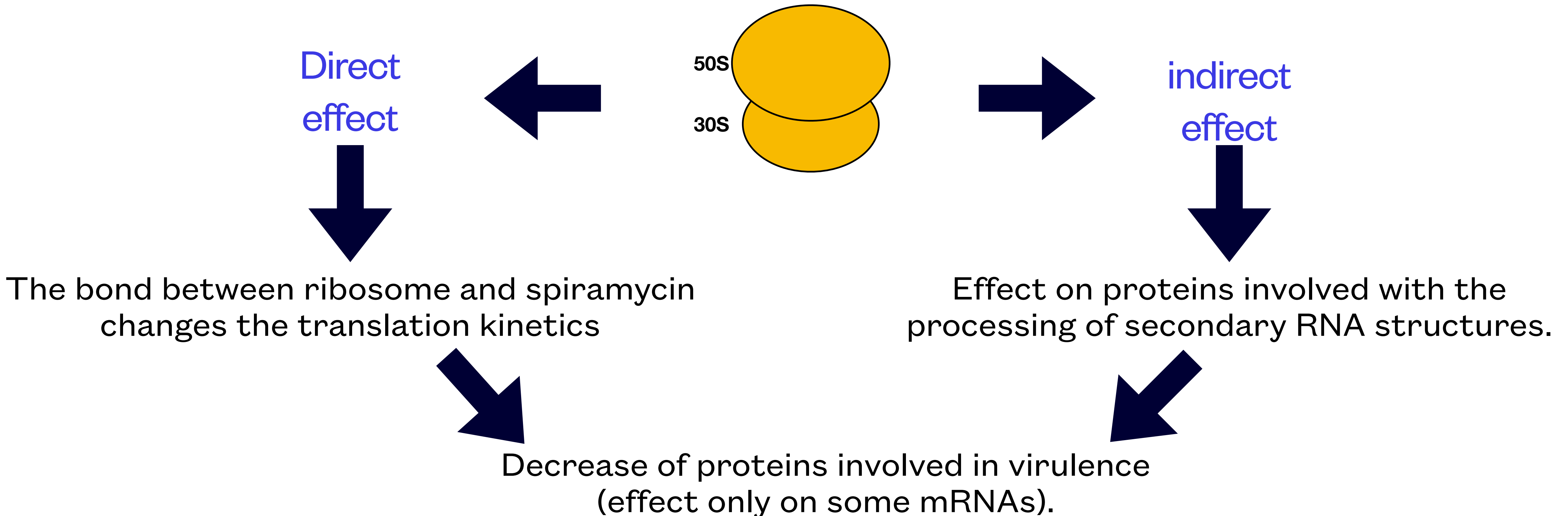
We included the three known QS regulators in *P. aeruginosa* in this analysis: LasR, PqsR and RhlR.



According to the results obtained with SwissDock, PqsR could be the target of spiramycin.

# Another Hypothesis ...

Another hypothesis of the mechanism of spiramycin against *P. aeruginosa* could concern ribosomes:



# Conclusions & Perspective

- Our in vitro results show as spiramycin inhibits motility, biofilm formation, rhamnolipids production and pigment secretion by *P. aeruginosa*.
- Our in vivo model show as spiramycin inhibit the virulence of *P. aeruginosa* injected in *G. mellonella* larvae.
- We have elaborated two hypotheses on the mechanism including:
  1. QS inhibition (supported by docking analysis)
  2. Direct or indirect interaction with the ribosome

We expect to obtain data on the mechanism of action in the future.

The data presented could be the basis for the design of clinical trials.



---

Thanks!

---

# References

- Antimicrobial resistance surveillance in Europe 2022 - 2020 data- ECDC
- Hirakata Y, Kaku M, Tomono K, Tateda K, Furuya N, Matsumoto T, Araki R, Yamaguchi K. Efficacy of erythromycin lactobionate for treating *Pseudomonas aeruginosa* bacteremia in mice. *Antimicrob Agents Chemother*. 1992 Jun;36(6):1198-203. doi: 10.1128/AAC.36.6.1198. PMID: 1416819; PMCID: PMC190317.
- Equi, A., Balfour-Lynn, I. M., Bush, A., & Rosenthal, M. (2002). Long term azithromycin in children with cystic fibrosis: a randomised, placebo-controlled crossover trial. *The Lancet*, 360(9338), 978-984.
- Pinnert-Sindico, S. (1955). A new antibiotic-spiramycin. *Antibiot. Ann.*, 1954, 724-727.