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Standardization of extraction of DNA from silica-gel dried leaf sample of *Garcinia indica* (Thouras) Choisy



Anila M Sunny¹, Sreedevi C. N¹, Vilas Kumar Patil², Tresa Hamalton¹ and N. Ravi¹

¹Institute of Wood science and Technology Bengaluru, India.

²College of Forestry, University of Agricultural Sciences, Dharwad, India.

Corresponding author: nravi@icfre.org

INTRODUCTION

- *Garcinia indica* is a fruit bearing tree with multifarious uses both traditionally and commercially.
- The leaves of *Garcinia indica* contains high quantity of secondary metabolites such as, phenolic acids, flavonoids, bioflavonoids, xanthones, benzophenones and terpenoids (Pandey *et al.*,2015).
- Quality and quantity of DNA will reduce if the leaf samples were brought without proper preservation.
- Transporting leaf sample in silica – gel is an inexpensive method of preservation.

MATERIALS AND METHODS

- Fresh mature leaf were collected from naturally distributed locations of Western ghats viz., Karwar (n = 6) and Kukke Subramanya (n =4).
- Leaves were cleaned with distilled water and air dried for 30 minutes. The samples were placed in zip lock bag containing dried silica gel and transported to laboratory in Bengaluru, Karnataka, India.
- Silica gel were removed periodically when the colour turns to white from blue.
- In laboratory leaves were dried till the whole moisture losses and stored in -20°C until further use.

Extraction of DNA by modified Thatte et al., 2012

- **Suspension buffer**

0.5M sucrose, 120mM Tris HCl, 50mM EDTA and 1.7 M NaCl.

- **Extraction buffer**

20 mM EDTA, 100 mM Tris HCl (pH 8), 1.4 M NaCl, 4% CTAB, 4% β -met.

Extraction of DNA by Thatte et al., 2012

- **Extraction buffer**

20 mM EDTA, 100 mM Tris HCl (pH 8), 1.4 M NaCl, 2 % CTAB, 0.2 % β -met

Genomic DNA extraction

Thatte *et al.*, 2012

Silica-gel dried leaf sample of 1.0 g were pulverised in liquid nitrogen in pestle and mortar

To the powdered leaf sample 9 ml of extraction buffer was added

The reaction mixture was homogenised thoroughly and incubated in water bath for 60° C for 1 hour.

An equal amount of C:I was added and centrifuged at 10,000rpm for 25 minutes.

Aqueous layer was collected and equal amount of CI was added. Mixture was centrifuged at 10,000 rpm for 10 min. This step was repeated twice.

Supernatant collected was added with 0.5 volume of 5 M NaCl and 2 volume of chilled ethanol.

The samples were centrifuged at 5000 rpm for 5 min and the pellets were washed with 80% ethanol.

The pellet were allowed to dry overnight and was dissolved in 0.1 X TE buffer. The DNA were stored in 20°C .

Modified extraction method

Silica -gel dried leaf samples of 1.0 g were pulverized in liquid nitrogen in ceramic pestle and mortar with 3% PVPP

Powdered leaf samples were transferred to centrifuge tube with 4 ml of suspension buffer and 100µl of β – met

Tubes were incubated at 65 ° C for 1 hour, centrifuged at 10,000 rpm for 10 min. The supernatant was transferred to another tube

Extraction buffer of 7 ml was added to supernatant and incubated in water bath at 65 ° C for 60 minutes

Equal amount of C:I was added and centrifuged at 15, 000 rpm for 10 minutes

Aqueous layer was collected and added 20 µl of RNase (20 mg/ml) and incubated at 37 ° C on dry bath for 2 hours. Followed by this 25 µl of proteinase (20 mg/ml) treatment was given for 1 hour in dry bath at 37 ° C

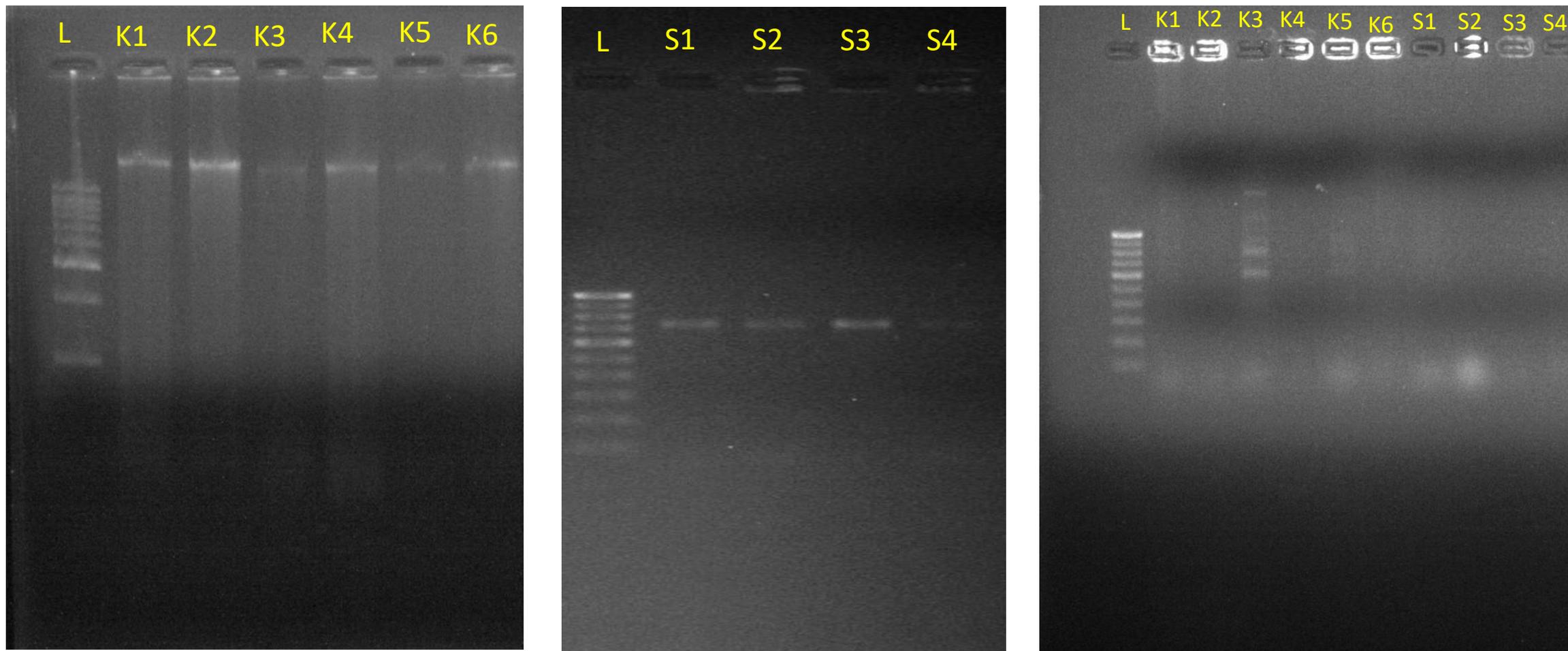
Equal amount of C:I was added, centrifuged at 10,000 rpm for 10 minutes. This step was repeated until a clear solution was obtained

Supernatant was transferred and added 100 µl of Ammonium acetate and ¾th volume of chilled ethanol and kept at -20°C for overnight

Next day, samples were centrifuged at 10,000 rpm for 10 minutes to develop DNA pellets. The pellets were then washed with 70% ethanol and air- dried at room temperature for 3 – 4 hours

The pellet was then dissolved in 30 -50 µl of TE buffer and stored at -20°C until further use

RESULTS



a

b

c

Fig 1. Genomic DNA of *Garcinia indica* silica- gel dried leaf sample extracted using modified Thatte *et al.*, 2012 method - (a) Karwar (K1-K6), (b) Kukke (S1 –S 4) and Thatte *et al.*, 2012 method – (c) Karwar (K1-K6) and Kukke (S1-S-6) resolved under 0.8 % agarose using 1Kb ladder (L)

Quality and Quantity of Genomic DNA

| Thatte <i>et al.</i> , 2012 | | |
|-----------------------------|--------------------|-----------------------------|
| SAMPLE NAME / ID | PURITY (A 260/280) | CONCENTRATION (ng/ μ l) |
| KARWAR -1/ K-1 | 1.21 | 20.6 |
| KARWAR -2/ K-2 | 1.35 | 55.6 |
| KARWAR -3/ K-3 | 1.10 | 34.2 |
| KARWAR -4/ K-4 | 1.00 | 82.3 |
| KARWAR -5/ K-5 | 1.12 | 41.8 |
| KARWAR -6/ K-6 | 1.20 | 15.1 |
| KUKKE -1/ S-1 | 1.30 | 25.3 |
| KUKKE -1/ S-1 | 1.10 | 39.3 |
| KUKKE -1/ S-1 | 1.25 | 57.4 |
| KUKKE -1/ S-1 | 0.99 | 32.9 |

| Modified extraction method | | |
|----------------------------|--------------------|-----------------------------|
| SAMPLE NAME / ID | PURITY (A 260/280) | CONCENTRATION (ng/ μ l) |
| KARWAR -1/ K-1 | 1.69 | 127.5 |
| KARWAR -2/ K-2 | 1.68 | 111.8 |
| KARWAR -3/ K-3 | 1.58 | 61.2 |
| KARWAR -4/ K-4 | 1.75 | 180.9 |
| KARWAR -5/ K-5 | 1.64 | 150.6 |
| KARWAR -6/ K-6 | 1.81 | 147.9 |
| KUKKE -1/ S-1 | 1.81 | 252.5 |
| KUKKE -1/ S-1 | 1.80 | 297.9 |
| KUKKE -1/ S-1 | 1.74 | 249.5 |
| KUKKE -1/ S-1 | 1.76 | 279.2 |

CONCLUSIONS

- The DNA of *Garcinia indica* obtained using modified extraction method was observed to have high purity and good quality.
- The purity of DNA obtained using modified method ranged between 1.58-1.81 and concentration ranging between 61 – 297 ng/μl.
- The results confirmed that modified extraction method can be used for DNA finger printing in *Garcinia indica*.

REFERENCE

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