

# Halotolerant *Bacillus* spp. strains isolated from the Great Sebkhah of Oran (Algeria): a source of antimicrobial secondary metabolites

Fatima NAS \*<sup>1</sup>, Nadia AISSAOUI<sup>2</sup>, Amina Nour Elhouda SAIBI<sup>1</sup>, Nihel KLOUCHE KHELIL<sup>1, 3</sup>

<sup>1</sup>Laboratory of Applied Microbiology in Food, Biomedical and Environment (LAMAABE), Faculty of Nature and Life, Earth and Universe Sciences, Department of Biology, Aboubekr Belkaid University of Tlemcen, Tlemcen, Algeria

<sup>2</sup>Laboratory for the Sustainable Management of Natural Resources in Arid and Semi Arid Areas, University center Salhi Ahmed Naâma, Naâma, Algeria

<sup>3</sup>Laboratory of Experimental Surgery, Dental Surgery Department, Medical Faculty, Abou bekr Belkaid University of Tlemcen, Tlemcen, Algeria

## Abstract

*Bacillus* species are Gram-positive bacteria found in abundance in nature, they could colonize different habitats even under extreme conditions and their secondary metabolites were found to possess various potential activities, notably antimicrobial. In this study, three-halotolerant *Bacillus* sp. LMB3051, LMB3073 and LMB3093 were isolated from water of the Great Sebkhah of Oran, they exhibited interesting antimicrobial activities against a broad spectrum of reference bacterial and fungal strains. Their metabolites were extracted with chloroform and ethyl acetate solvents. Structural elucidation of active compounds was carried out using gas chromatography–mass spectrometry (GC-MS). Fifty-six compounds were identified; they include tert-butyl phenol compounds, fatty acid methyl esters due to the methylation procedure, hydrocarbons, aldehydes, benzoquinones, pyrroles, and terpenes. Literature reports such compounds to have wide biological and pharmaceutical applications. Partial 16S rRNA gene sequencing of the three isolates showed very high similarity with many species of *Bacillus*. The comparison with 16S rRNA EzBioCloud database revealed that the isolates LMB3051 (529 bp) and LMB3073 (429 bp) showed a similarity of 99.81% and 98.60%, respectively, with *Bacillus licheniformis* ATCC 14,580, *B. aureus* 24 K, *B. paralicheniformis* KJ-16, *B. glicinifermentans* GO-13, *B. haynesii* NRRL B-41327, and *B. piscis* 16MFT21. The isolate LMB3093 (400 bp) showed 98.75% of similarity with *Bacillus paralicheniformis* KJ-16 and *B. haynesii* NRRL B-41327. The findings suggest that the Great Sebkhah of Oran is a valuable source of strains exhibiting variety of beneficial attributes that can be utilized in the development of biological antibiotics.

## Materials and Methods

### Phenotypical characterization

morphological, cultural, biochemical, and physiological characterization was realized as described by Logan *et al.* (2009)

### Molecular identification

Genomic DNA was extracted using the standard phenol/chloroform method (Ettoumi *et al.* 2013). The amplification of the 16S rRNA gene was carried out using, the universal Primers 5'-S-D-Bact-0008-a-S-20-3' and 5'-S-D-Bact-1495-a-S-20-3' (Daffonchio *et al.* 2000). The amplified 16S rRNA fragments were sequenced and identified using EzBioCloud databases (Yoon *et al.* 2017).

### Screening of antimicrobial activities

The antagonistic properties of the isolates were screened by the agar plug method (Balouiri *et al.* 2016) against Gram positive bacteria (*Bacillus cereus* ATCC11778 and *Staphylococcus aureus* ATCC 25,923), Gram-negative bacteria (*Escherichia coli* ATCC 25,922 and *Acinetobacter baumannii* ATCC 19,606), and fungi (*Candida albicans* ATCC 444, *Candida albicans* ATCC 10,231, and *Aspergillus flavus* MNHN994294

### Extraction of secondary metabolites

Was performed from solid substrate fermentation (agar culture) with two organic solvents, ethyl acetate, and chloroform, using the method of Kim *et al.* (2016)

### GC-MS analysis of bioactive crude extract

The analysis was carried out using Agilent 7890B gas chromatography instrument coupled with an Agilent MS 240 Ion Trap with capillary column HP-5MS (5% phenyl methyl polysiloxane, 30 m, 250 µm, 0.25 µm)

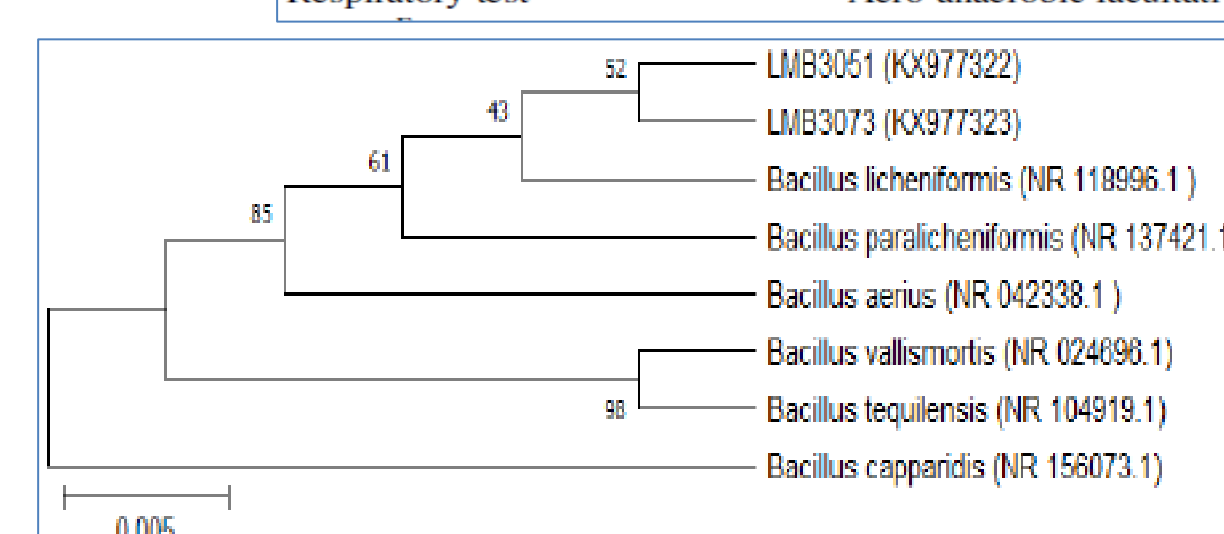
## References

- Logan NA, Berge O, Bishop AH, Busse HJ, De Vos P, Fritze D, Heyndrickx M, Kämpfer P, Rabinovitch I, Salkinoja-Salonen MS, Seldin I, Ventosa A, (2009) Proposed minimal standards for describing new taxa of aerobic, endospore-forming bacteria. *Int J Syst Evol Microbiol* 59:2114–2121. <https://doi.org/10.1099/ijms.0.013649-0>
- Ettoumi B, Guesmi A, Brusetti L, Borin S, Najjari A, Boudabous AF, Cherif A (2013) Microdiversity of deep-sea Bacillales isolated from Tyrrhenian Sea sediments as revealed by ARISA, 16S rRNA gene sequencing and BOX-PCR fingerprinting. *Microbes and Environ* 28:361–369. <https://doi.org/10.1264/jmsme2.ME13013>
- Daffonchio D, Cherif A, Borin S (2000) Homoduplex and heteroduplex polymorphisms of the amplified ribosomal 16S–23S internal transcribed spacers describe genetic relationships in the *Bacillus* cereus group. *Appl Environ Microbiol* 66:5460–5468. <https://doi.org/10.1128/AEM.66.12.5460-5468.2000>
- Balouiri M, Sadiki M, Ibsouda SK (2016) Methods for in vitro evaluating antimicrobial activity: a review. *J Pharm Anal* 6:71–79. <https://doi.org/10.1016/j.jpha.2015.11.005>
- Kim HY, Heo DY, Park HM, Singh D, Lee CH (2016) Metabolomic and transcriptomic comparison of solid-state and submerged fermentation of *Penicillium expansum* KACC 40815. *PLoS ONE* 11(2):e0149012. <https://doi.org/10.1371/journal.pone.0149012>

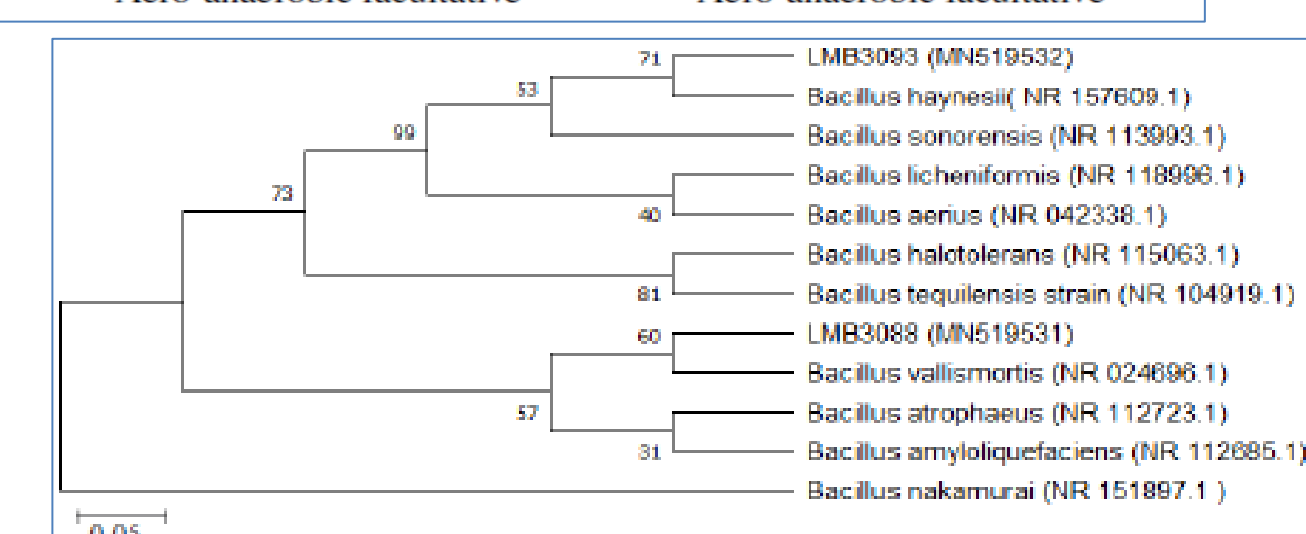
## Results

**Table 1.** Morphological, physiological, and biochemical characteristics of LMB3051, LMB3073, and LMB3093 strains

Characteristics	LMB3051	LMB3073	LMB3093
Gram staining	+	+	+
Motility	Motile	Motile	Motile
Cell morphology	Long and straight rods	Short rods	Long big rods
Arrangement	Singly or in chains	Singly and in irregular clumps	Singly
Spore	Ellipsoidal subterminal	Ellipsoidal subterminal	Ellipsoidal subterminal
Colonies (24 h on nutrient agar at 15% of NaCl)	3 to 4 mm in diameter, cream, opaque, circular, smooth, shiny, slimy, convex, and odorless	3 to 4 mm in diameter, whitish, opaque, circular, smooth, shiny, slimy, convex, no smell	2 to 4 mm in diameter, cream, opaque, circular, smooth, slimy, shiny, flat, and odorless
NaCl tolerance	0 to 20% (w/v)	0 to 20% (w/v)	0 to 20% (w/v)
NaCl optimum (%)	0%	0%	0%
Range of temperature for growth	15 to 60 °C	15 to 60 °C	15 to 60 °C
Temperature optimum °C	30–40 °C	30–37 °C	30–37 °C
Range of pH for growth	5 to 11	5 to 11	5 to 11
pH optimum	6–7	6–7	6–7
Catalase	+	+	+
Oxidase	+	+	+
Respiratory test	Aero-anaerobic facultative	Aero-anaerobic facultative	Aero-anaerobic facultative



**Fig 1.** Phylogenetic tree constructed by performing bootstrap analysis of 1000 data sets using the MEGA 6 program, based on the analysis of 16S rRNA partial sequences showing the relationships between isolates LMB3051 and LMB3073 and related species. *Bacillus capparidis* was used as an outgroup. The bar indicates 0.005 substitution per nucleotide position. Accession numbers are given in parentheses



**Fig 2.** Phylogenetic analyses of the isolate LMB3093 based on 16S rRNA partial sequences. Phylogenetic dendrogram was evaluated by performing bootstrap analysis of 1000 data sets using MEGA 6. 16S rRNA sequence. *Bacillus nakamurai* was used as an outgroup. The bar indicates 5 substitution per 100 nucleotide. Accession numbers of the reference strains were indicated in parentheses

**Table 2.** Inhibitory effects of the chloroform and ethyl acetate extracts of the isolates using disc diffusion method

Test microorganisms	Zone's diameter (mm)					
	LMB3051		LMB3073		LMB3093	
	Chloroform	Ethyl acetate	Chloroform	Ethyl acetate	Chloroform	Ethyl acetate
<i>Bacillus cereus</i> ATCC 11,778	15.33 ± 0.57 <sup>a</sup>	14.33 ± 0.57 <sup>a</sup>	17.33 ± 0.57 <sup>a</sup>	22.33 ± 0.57 <sup>a</sup>	13.33 ± 0.57 <sup>a</sup>	12.66 ± 0.57 <sup>a</sup>
<i>Staphylococcus aureus</i> ATCC 25,923	19.66 ± 0.57 <sup>a</sup>	17.66 ± 0.57 <sup>a</sup>	16 ± 1 <sup>a</sup>	19.33 ± 0.57 <sup>a</sup>	15.33 ± 1.52 <sup>a</sup>	14 ± 1 <sup>a</sup>
<i>Escherichia coli</i> ATCC 25,922	13.33 ± 0.57 <sup>a</sup>	12.33 ± 0.57 <sup>a</sup>	14 ± 1 <sup>a</sup>	14 ± 1 <sup>a</sup>	11.33 ± 1.15 <sup>a</sup>	10 ± 0 <sup>a</sup>
<i>Acinetobacter baumannii</i> ATCC 19,606	10 ± 0 <sup>a</sup>	9.66 ± 1.52 <sup>a</sup>	12.66 ± 1.15 <sup>a</sup>	15.66 ± 0.57 <sup>a</sup>	09.66 ± 1.52 <sup>a</sup>	09 ± 1 <sup>a</sup>
<i>Candida albicans</i> IP 444	13 ± 0 <sup>a</sup>	12 ± 1 <sup>a</sup>	11.66 ± 0.57 <sup>a</sup>	16 ± 0 <sup>a</sup>	10.33 ± 1.52 <sup>a</sup>	10 ± 1 <sup>a</sup>
<i>Candida albicans</i> ATCC 10,231	12.66 ± 0.57 <sup>a</sup>	11.66 ± 0.57 <sup>a</sup>	13.66 ± 1.15 <sup>a</sup>	11 ± 1 <sup>a</sup>	10 ± 1 <sup>a</sup>	10 ± 0 <sup>a</sup>
<i>Aspergillus flavus</i> MNHN994294	33.33 ± 0.57 <sup>a</sup>	31.66 ± 0.57 <sup>a</sup>	34.66 ± 1.52 <sup>a</sup>	37.66 ± 0.57 <sup>a</sup>	30.33 ± 0.57 <sup>a</sup>	29 ± 1 <sup>a</sup>

Values were expressed as mean ± standard deviation (n=3). Values in same columns with superscript (a) were strongly significant different (p < 0.001)

**Table 2.** Chemical composition of the chloroformic and ethyl acetate extracts of the isolate LMB3051

Solvent	Compounds identified	CAS n	%
Chloroform	Hexadecanoic acid bis (2ethylhexyl) ester	103-23-1	100
	3-n-hexylthiolane, S, S, dioxid	71053-07-1	6.059
Ethyl acetate	1,3-Di-tert-butyl benzene	1014-60-4	1.360
	2,5-Di-tert-butyl-benzene	2460-77-7	0.512
	1,4-benzoquinone	719-22-2	27.646
	1,4-benzoquinone	719-22-2	27.646
	2,4-Di-tert-butyl-phenol	96-76-4	8.310
	phenol	None	13.734
	1-Nonyl cycloheptane	None	13.734
	3,5-Di-tert-butyl-4-hydroxybenzaldehyde	1620-98-0	5.963
	hexadecanoic acid bis (2ethylhexyl) ester	103-23-1	35.957
	Hexadecanoic acid bis (2ethylhexyl) ester	103-23-1	35.957

**Table 2.** Chemical composition of the chloroformic and ethyl acetate extracts of the isolate LMB3073

Solvent	Compounds identified	CAS n	%
Chloroform	1,3-Di-tert-butyl benzene	1014-60-4	1.548
	1-Fluorododecane	334-68-9	5.125
	1-Nonyl cycloheptane	None	0.767
	2,6-Di-tert-butyl-1,4-benzoquinone	719-22-2	0.597
	2,6-di-tert-butyl-4-(dimethylaminomethyl) phenol	88-27-7	24.741
	2,4-di-tert-butyl phenol	96-76-4	4.649
	3,5-di-tert-butyl-4-hydroxyacetophenone	14035-33-1	1.075
	2,4,6-Tri-tert-butylphenol	732-26-3	0.874
	3,5-Di-tert-butyl-4-hydroxybenzaldehyde	1620-98-0	13.340
	4,4'-Ethylenebis(2,6-di-tert-butylphenol)	1516-94-5	2.756
	Pyrrrolo [1,2-a] pyrazine-1,4-dione, hexahydro-3-	5654-86-4	6.928
	2-Propenoic acid, 3-(4-methoxyphenyl)	830-09-1	2.969
	Tetracontane, 3,5,24,trimethyl	55162-61-7	3.335
	Carbonic acid ticosyl vinyl ester	None	5.74
Ethyl acetate	9-Octadecenoamide, (z)-	301-02-0	3.728
	1-Nonyl cycloheptane	None	11.205
	1,3-Di-tert-butyl benzene	1014-60-4	1.449
	2 (3H)-Naphthalene, 4, 4a, 5, 7,8-hexahydro-4a-5-dimethyl-3-(1-methyl ethylidene)-4ar-cis	1014-60-4	1.449
	2,4-di-tert-butyl phenol	96-76-4	4.667
	Cyclo pentane acetic acid, 3-oxo-2-pentyl-methyl ester	24851-98-7	4.066
	Tridecanoic acid, 12-methyl-methyl ester	5129-58-8	0.788
	3,5-Di-tert-butyl-4-hydroxybenzaldehyde	1620-98-0	3.382
	7-Acetyl-6-ethyl-1,4,4-tetrahydronaphthalene	88-29-9	3.103
	Hexadecanoic acid methyl ester	112-39-0	5.807
Heptadecanoic acid, 16 methyl-methyl ester	5129-61-3	4.806	
Hexadecanoic acid bis (2 ethylhexyl) ester	103-23-1	36.781	

**Table 2.** Chemical composition of the chloroformic and ethyl acetate extracts of the isolate LMB3093

Solvent	Compounds identified	CAS n	(%)	
Chloroform	1,3-Di-tert-butyl benzene	1014-60-4	1.955	
	1-Fluorododecane	334-68-9	8.089	
	1-Nonyl cycloheptane	None	1.459	
	2,6-Di-tert-butyl-1,4-benzoquinone	719-22-2	0.735	
	2,6-di-tert-butyl-4-(dimethylaminomethyl) phenol	88-27-7	19.297	
	2,4-di-tert-butyl phenol	96-76-4	5.059	
	3,5-di-tert-butyl-4-hydroxyacetophenone	14035-33-1	1.433	
	2,4,6-Tri-tert-butylphenol	732-26-3	1.188	
	3,5-Di-tert-butyl-4-hydroxybenzaldehyde	1620-98-0	11.839	
	Propanohydrizide, N2-(3,5-di-tert-butyl-4-hydroxybenzyl)-N2-phenyl	304870-86-8	2.687	
	Pyrrrolo [1,2-a] pyrazine-1,4-dione, hexahydro-3-(2-methylpropyl)-	5654-86-4	10.012	
	Tritetracontane	7098-21-7	1.818	
	4,4'-Ethylenebis (2,6-di-tert-butylphenol)	1516-94-5	1.270	
	2-Propenoic acid, 3-(4-methoxyphenyl)	830-09-1	2.185	
	Z-8-methyl-9-tridecanoic acid	None	1.018	
	9-Octadecenoamide, (z)-	301-02-0	0.509	
	Hexadecanoic acid bis (2ethylhexyl) ester	103-23-1	1.579	
	13-Docosanamide, (Z)	112-84-5	5.160	
	1-Undecene	2243-98-3	0.840	
	3-n-hexylthiolane, S, S, dioxid	71053-07-1	5.467	
	Ethyl acetate	1-Tetradecyne	765-10-6	1.189
		2,5-di-tert-butyl-1,4-benzoquinone	2460-77-7	0.464
2,6-Di-tert-butyl-1,4-benzoquinone		719-22-2	4.103	
2,4-di-tert-butyl phenol		96-76-4	6.031	
1-Nonyl cycloheptane		None	10.564	
2-Propenoic acid, 3-(4-methoxyphenyl)		830-09-1	4.75	
Oleyl alcohol, trifluoro acetate		None	1.004	
Benzoic acid, tridecyl ester		None	2.966	
Hexadecanoic acid bis (2ethylhexyl) ester		103-23-1	3.609	
Benzoic acid, pentadecyl ester		None	1.805	
Z,5 Methyl-6-Heneicosen-11-one	None	1.254		