

The 8th International Electronic Conference on Medicinal Chemistry (ECMC 2022) 01–30 NOVEMBER 2022 | ONLINE

## Antimicrobial activity and DNA/BSA binding affinities of silver(I) and gold(III) complexes with 1,6-naphthyridine

Chaired by **DR. ALFREDO BERZAL-HERRANZ**; Co-Chaired by **PROF. DR. MARIA EMÍLIA SOUSA** 



1972



# Darko P. Ašanin<sup>1</sup>\*, Marija Nenadovic<sup>2</sup>, Tina P. Andrejević<sup>3</sup>, Sandra Vojnovic<sup>2</sup>, Miloš I. Djuran<sup>4</sup> and Biljana Đ. Glišić<sup>3</sup>

<sup>1</sup>University of Kragujevac, Institute for Information Technologies Kragujevac, Department of Science, Jovana Cvijića bb, 34000 Kragujevac, Serbia; <sup>2</sup>University of Belgrade, Institute of Molecular Genetics and Genetic Engineering, Vojvode Stepe 444a, 11042 Belgrade, Serbia; <sup>3</sup>University of Kragujevac, Faculty of Science, Department of Chemistry, R. Domanovića 12, 34000 Kragujevac, Serbia; <sup>4</sup>Serbian Academy of Sciences and Arts, Knez Mihailova 35, 11000 Belgrade, Serbia

\* Corresponding author: darko.asanin@uni.kg.ac.rs

### Antimicrobial activity and DNA/BSA binding affinities

of silver(I) and gold(III) complexes with 1,6-naphthyridine



### Abstract

Silver(I) and gold(III) complexes with aromatic nitrogen-containing heterocycles have shown an effective and widespectrum antimicrobial activity. The possible mechanism of their antimicrobial activity can be attributed to the interactions of these complexes with biomolecules, including DNA and proteins. In the present study, new silver(I) complex with 1,6-naphthyridine (1,6-naph), {[Ag(1,6-naph)(H<sub>2</sub>O)](BF<sub>4</sub>)}<sub>n</sub> (**1**) was synthesized and characterized by NMR, IR and UV-Vis spectroscopy, and its crystal structure was determined by single-crystal X-ray diffraction analysis. The complex **1** and the previously reported analogue gold(III) complex [1], [AuCl<sub>3</sub>(1,6-naph)] (**2**), were evaluated for antimicrobial activity against the panel of representative microorganisms, while their cytotoxicity was tested against normal human lung fibroblast cell line (MRC-5). The binding affinity of these complexes with calf thymus DNA (ct-DNA) and bovine serum albumin (BSA) was studied to clarify the mode of their antimicrobial activity [2].

Keywords: Silver(I) complexes; Gold(III) complexes; 1,6-Naphthyridine; Antimicrobial activity; DNA/BSA interaction.

B. Đ. Glišić, B. Warżajtis, M. Hoffmann, U. Rychlewska, M. I. Djuran, RSC Adv. 10 (2020) 44481-44493.
 D. P. Ašanin, M. Nenadovic, T. P. Andrejević, S. Vojnovic, M. I. Djuran, B. Đ. Glišić, manuscript in preparation

### есмс 2022

#### Introduction

- Silver(I) and gold(III) complexes with aromatic nitrogen-containing heterocycles have shown an effective and widespectrum antimicrobial activity
- ✓ One of the possible mechanism of their antimicrobial activity can be attributed to interactions of these complexes with biological targets, including DNA and proteins



1,6-naphthyridine (1,6-naph)

### ECMC 2022

### **Results and discussion**



[1] B. Đ. Glišić, B. Warżajtis, M. Hoffmann, U. Rychlewska,M. I. Djuran, RSC Adv. 10 (2020) 44481-44493.



### **Structural characterization**

### Crystal structure of silver(I) complex 1





### $\checkmark$ <sup>1</sup>H NMR spectroscopic characterization of 1



### ✓ UV-Vis stability of 1



Time stability of complex **1** followed by UV-Vis spectrophotometry at room temperature in DMSO

#### ✓ Electrochemical characterization of 1



Cyclic voltammogram of complex **1** recorded at the GC electrode in DMSO and 0.1 M tetrabutylammonium hexafluorophosphate (TBAHP) as a supporting electrolyte at a scan rate of 50 mV/s. The conditions are given as follows:  $E_{begin} = -2.0 \text{ V}$ ,  $E_{end} = 2.0 \text{ V}$  and  $E_{step} = 0.002 \text{ V}$ 

### ECMC 2022

### **Antimicrobial activity**

Antimicrobial activity of silver(I) 1 and gold(III) 2 complexes and the corresponding ligand expressed as MIC (μg/mL) in comparison to their cytotoxicity against healthy human fibroblasts MRC-5 (IC<sub>50</sub>, μg/mL)

Test organism	. 1	2	1,6-naph	
Compounds	T	2		
Staphylosossus aureus NCTC 6571	31.25	62.50	>500	
Listeria monocytogenes NCTC 1194	31.25	62.50	>500	
Pseudomonas aeruginosa NCTC 10338	31.25	62.50	>500	
<i>Escherichia coli</i> NCTS 9001	7.81	62.50	>500	
Candida albicans ATCC 10231	3.90	>500	>500	
<i>Candida parapsilosis</i> ATCC 22019	0.49	>500	>500	
Klebsiella pneumoniae ATCC BAA	15.62	>500	>500	
MRC-5	12 ± 0.8	120 ± 8	500 ± 10	



### **DNA binding study**

### ✓ DNA interaction of complexes 1 and 2 was studied by florescence spectroscopy

Complex	K <sub>sv</sub> (M <sup>-1</sup> )	Hypochromism (%)	<i>K<sub>q</sub></i> (M <sup>-1</sup> s <sup>-1</sup> )	<i>К<sub>А</sub></i> (М <sup>-1</sup> )	n
1	(9.89±0.02)·10 <sup>2</sup>	11.90	9.89 <sup>.</sup> 10 <sup>10</sup>	6.11 <sup>.</sup> 10 <sup>3</sup>	1.21
2	(7.75±0.01) <sup>.</sup> 10 <sup>5</sup>	53.92	7.75 <sup>.</sup> 10 <sup>13</sup>	4.57 <sup>.</sup> 10 <sup>10</sup>	2.55

[complex 2] =  $0 - 70 \mu$ M, Phosphate buffer saline (pH = 7.4)



ECMC 2022

### **BSA binding study**

### ✓ BSA interaction of complexes 1 and 2 was studied by florescence spectroscopy

Complex	<i>K<sub>sv</sub></i> (M <sup>-1</sup> )	Hypochromism (%)	<i>K<sub>q</sub></i> (M <sup>-1</sup> s <sup>-1</sup> )	<i>K<sub>A</sub></i> (M⁻¹)	n
1	(5.05±0.01)·10 <sup>3</sup>	41.96	5.05 <sup>.</sup> 10 <sup>11</sup>	2.12 <sup>.</sup> 10 <sup>4</sup>	1.18
2	(4.90±0.05) <sup>.</sup> 10 <sup>4</sup>	61.50	4.90 <sup>.</sup> 10 <sup>12</sup>	1.26 <sup>.</sup> 10 <sup>6</sup>	1.45

 $[\text{complex 1}] = 0 - 160 \,\mu\text{M}, \text{Phosphate buffer saline (pH = 7.4)}$ 



### ECMC 2022

### Synchronous fluorescence spectroscopy

- ✓ Synchronous fluorescence spectroscopy was used to explore the structural changes in BSA in the presence of the investigated complexes
- ✓ When  $\Delta\lambda$  is 15 nm, the synchronous fluorescence is characteristic of the tyrosine (Tyr) residue, while a larger  $\Delta\lambda$  value of 60 nm is due to tryptophan (Trp)

	Complex	<i>K<sub>sv</sub></i> (M⁻¹)	Hypochromism (%)	<i>K<sub>q</sub></i> (M⁻¹s⁻¹)	<i>K<sub>A</sub></i> (M⁻¹)	n
Δλ = 15 nm	1	(4.32±0.01) <sup>.</sup> 10 <sup>3</sup>	39.99	4.32 <sup>.</sup> 10 <sup>11</sup>	7.85 <sup>.</sup> 10 <sup>3</sup>	1.07
	2	(2.12±0.06) <sup>.</sup> 10 <sup>5</sup>	69.28	2.12 <sup>.</sup> 10 <sup>13</sup>	6.88 <sup>.</sup> 10 <sup>7</sup>	1.86
Δλ = 60 nm	1	(3.70±0.01) <sup>.</sup> 10 <sup>3</sup>	35.56	3.70 <sup>.</sup> 10 <sup>11</sup>	2.34 <sup>.</sup> 10 <sup>4</sup>	1.22
	2	(1.02±0.04) <sup>.</sup> 10 <sup>5</sup>	68.86	1.02 <sup>.</sup> 10 <sup>13</sup>	1.27 <sup>.</sup> 10 <sup>7</sup>	1.68



✓ Synchronous fluorescence spectra of BSA in the absence and presence of increasing concentrations of complex 1



### Conclusions

- New silver(I) complex with with 1,6-naphthyridine (1,6-naph), {[Ag(1,6-naph)(H<sub>2</sub>O)](BF<sub>4</sub>)},
  (1) was synthesized and structurally characterized
- ✓ 1,6-naph ligand is monodentately coordinated to the Ag(I) ion through the nitrogen atom, leading to the formation of [Ag(1,6-naph)(H<sub>2</sub>O)]<sup>+</sup> complex cation
- ✓ The synthesized silver(I) complex 1 has shown a good antimicrobial activity, especially against *Candida* strains, being, in all cases, more active than gold(III) analogue 2
- ✓ Both Ag(I) and Au(III) complexes, 1 and 2, respectively, have the ability to interact with DNA and BSA, with Au(III) complex being more reactive towards these two biomolecules

### ECMC 2022





### **Acknowledgments**

This research has been financially supported by the Ministry of Education, Science and Technological Development of the Republic of Serbia (Agreement No. 451-03-68/2022-14/20042, 451-03-68/2022-14/200122 and 451-03-68/2022-14/200378) and the Serbian Academy of Sciences and Arts (project F128).

