

Some problems arising during the initiation of somatic embryogenesis in *Pinus sylvestris* L.[†]

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Abstract: The use of biotechnological tools in particular somatic embryogenesis (SE) for mass propagation of conifers is relevant, since this method allows to quickly replicate plant material with desired features. However, there are still a number of difficulties in obtaining embryogenic cell culture for *Pinus sylvestris*. One of the important and unsolved problems is the search for SE-competent genotypes. 674 megagametophytes from 22 donor plants (16 genotypes) were cultured in vitro during 2021 summer period. As a result of the experiment, callus formation was not recorded for the studied genotypes, however, 9.4±1.0% of the explants formed plants. In addition to the genotype effect, unsuitable nutrient medium or late developmental stages of zygotic embryos could be the reasons for the lack of callus induction. To solve these problems, a number of studies were carried out: (1) the effect of the nutrient medium composition and density (MS, MSG, ½LV, DCR) on the callus initiation from mature seeds was analysed, (2) the effect of various growth regulators concentrations on the initiation of callus formation was studied, (3) the analysis of the reproductive competence of donor plants was performed by the method of vegetative buds cultivation. As a result, several genotypes were found to have the ability for embryogenic callus formation, and the conditions for explants cultivation were selected.

Keywords: somatic embryogenesis, Scots pine, medium composition, mature seeds, vegetative buds, Karelia, forest seed plantation.

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1. Introduction

Somatic embryogenesis (SE) is a promising and effective biotechnological method for obtaining a large amount of coniferous plant material throughout the year. Despite the availability of data on the successful initiation of this process in Scots pine (*Pinus sylvestris* L.), many researchers agree that this species is one of the most difficult to undergo SE [1–3]. It is considered that the successful initiation of SE in *P. sylvestris* depends on multiple factors [3–6]: the efficiency of surface sterilization protocol of plant material; the explant type; donor plant (genotype) capable of SE; cultivation conditions, in particular, the composition of the nutrient medium and the content of plant growth regulators; stage of the zygotic embryo development.

For several years, the team of our laboratory conducted research aimed at initiating SE in *P. sylvestris* using megagametophytes with immature zygotic embryos collected from plus trees' clones from the Petrozavodsk Seed Orchard (SO) of the 1st order as explants, and the protocol developed by M. Abrahamsson and co-authors [7]. Thus, in 2021, 674 megagametophytes were introduced into culture *in vitro*, which were collected from 22 clones of plus trees (16 genotypes). However, no embryonic-suspensor mass was

obtained, and 9.4±1.0% of the explants formed plants. 48

In this regard, we carried out a number of experiments aimed at finding out the 49
possible reasons for the lack of SE initiation in *P. sylvestris* explants using vegetative buds 50
and mature seeds. 51

2. Materials and methods 52

2.1. Determination of the donor plants' reproductive potential 53

Vegetative buds from plus trees clones (40 years old) growing on the Petrozavodsk 54
SO of the 1st order (Karelia, Russia) [8] were collected during the period of forced dor- 55
mancy in 2021, late February – early March (16 genotypes 2 clones each); and in 2022, the 56
end of March – beginning of April (6 genotypes, 2 clones each). Buds, without detaching 57
from the shoot, were surface sterilized in a soap solution for 10 minutes, then washed 58
under running water. Under aseptic conditions, buds were placed in 5% sodium hypo- 59
chlorite solution for 10 minutes, with a three-fold treatment using sterile water, after 60
which buds were placed in 20% hydrogen peroxide for 10 minutes, with three thorough 61
washings in sterile water. Buds were cleaned from integument layers in a laminar box, 62
cut into 2-3 mm thick transverse disks, which were placed on Murashige-Skoog nutrient 63
medium modified by A. Hohtola [9], 2,4-dichlorophenoxyacetic acid (2,4-D) and 64
6-benzylaminopurine (BA) at concentrations of 2 and 1 mg/l, respectively, were used as 65
growth regulators, 10 g/l sucrose served as a carbohydrate source. 5-6 replicates were 66
provided for each tree. 4 explants were cultivated per jar (one replicate). The description 67
of the ongoing processes was performed on the 30th day of the experiment. Parameters 68
such as weight, initiation frequency, and proportion of light callus were analyzed. 69

The cytological analyses of the calluses obtained were conducted. The callus was 70
placed on a glass slide, kept for 1-2 minutes in the dye (0.2% safranin water solution with 71
the addition of a methylene blue drop) [6]. Squashed preparations were viewed under 72
the light microscope (Carl Zeiss Primo Star) at 4× 10 × magnifications. 73

2.2. Study on the effect of plant growth regulators' different concentrations 74

The impact of phytohormones various concentrations on the megagametophytes 75
reaction was performed on DCR medium [10]. 12 medium types were prepared, which 76
differed in the content of plant growth regulators (PGR) and sucrose (Table 1). We used 77
population mixtures of mature seeds collected from *P. sylvestris* trees located on the Pet- 78
rozavodsk SO and in a park on the territory of Petrozavodsk (the age of the trees is 20 79
years) as explants. Explants were megagametophytes containing mature zygotic embryo- 80
s. Sterilization of plant material was carried out in accordance with the protocol de- 81
scribed above. Megagametophytes were extracted from mature seeds, peeled and placed 82
horizontally on a medium, 4 explants per jar (5 replicates). The formation of plants and/or 83
calluses was registered on the 21st day of experiment. 84

Table 1. The content of growth regulators and sucrose in different types of the DCR medium. 85

Component	Nutrient medium number											
	1	2	3	4	5	6	7	8	9	10	11	12
2,4-D, μM	9.0	13.6	2.2	9.0	4.4	13.6	–	9.0	9.0	–	13.6	–
NAA, μM	–											2.7
BA, μM	4.4	2.2	2.2	9.0	4.4	13.6	4.4	–	2.2	–	9.0	9.0
Sucrose, g l ⁻¹	30											10

Note. NAA – 1-naphthylacetic acid.

2.3. Study of the influence of density and composition of the nutrient medium

To study the effect of the composition and density of the nutrient medium on the reaction of *P. sylvestris* megagametophytes from mature seeds, we used explants from various habitats: the natural phytocoenosis of the Medvezhyegorsk region of the Republic of Karelia (the age of the trees is 80–100 years) and the Petrozavodsk city park. During the study nutrient media MSG [5], MS [11], ½ LV [6], DCR [7] with the same content of phytohormones 9.0 µM 2,4-D and 4.4 µM BA, which differ in the composition of micro- and macroelements, sucrose content and have two density options (standard content and reduced content of gelling agent marked with “-”) were used (Table 2). Sterilization, introduction and description of explants into culture *in vitro* was performed according to the protocol described above.

Table 2. The content of gelling agent and sucrose in different types of nutrient media

Component g l ⁻¹	Nutrient medium							
	MSG	MSG-	MS	MS-	½ LV	½ LV-	DCR	DCR-
Sucrose	10		30		30		10	
Agar	7	3,5	6	3	7	3,5	–	3,5
Gelrite	–		–		–		3,5	–

2.4. Statistic analysis

Data were statistically processed with Microsoft Excel 2007 and PAST (4.0). Spearman rank correlation was used to measure the statistical dependence. All assays were performed at the Core Facility of the Karelian Research Centre RAS.

3. Results and discussion

3.1. The evaluation of the donor-plants reproductive potential

It was found that in 2021 the callus from the *P. sylvestris* vegetative buds was formed on 5–11th day of cultivation. Data analysis showed that with an increase in the average mass of buds callus (from 0 to 1 g), the proportion of light callus (from 4 to 61%) and the frequency of its initiation (from 38 to 90%) increases (Spearman correlation $r = 0.52$, $p = 0.002$ and $r = 0.38$, $p = 0.03$, respectively). Based on the data obtained 6 genotypes were selected, which were capable on callus formation from buds with the highest mass (516, 856, 876, 1025, 1026). In 2022 explants were collected from these trees with further introduction into culture medium. On the 30th day of the study, the frequency of callus initiation in explants collected from different plus trees clones varied on average from 20 to 60%. Cytogenetic analysis showed that there are two types of cells forming the callus (Fig. 1): meristematic (rounded) and parenchymal (elongated). Moreover emerging single somatic embryos were registered in the genotype 1025-5 culture, which, probably, may indicate a predisposition of this genotype to SE. There is information in the literature about the formation of somatic embryoids in cell culture, where vegetative buds of *P. sylvestris* were used as explants [12].

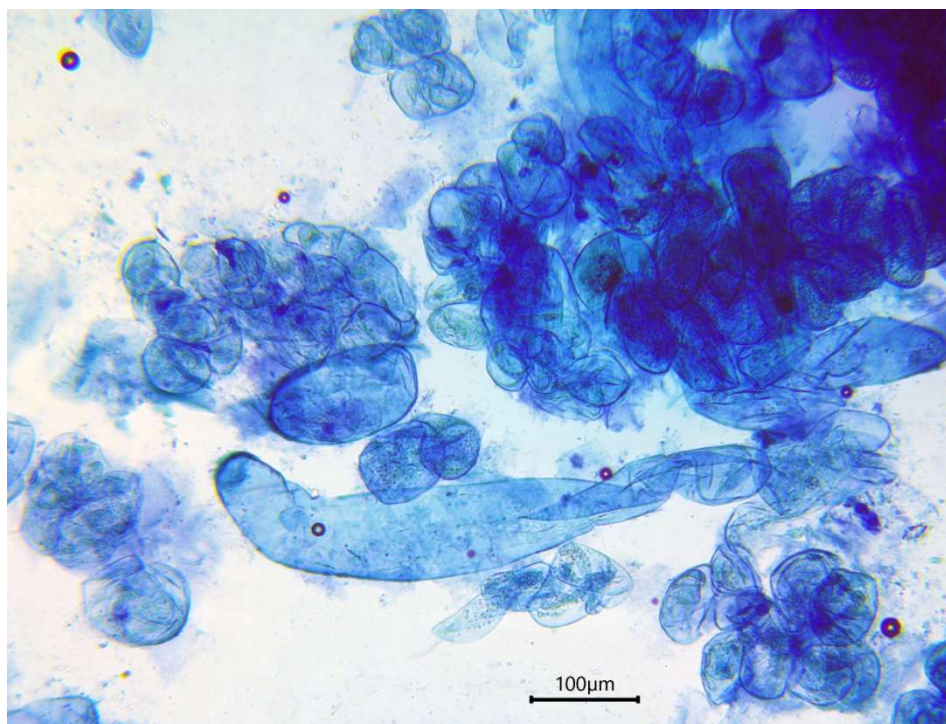


Figure 1. Meristematic and parenchymal types of somatic cells in *P. sylvestris* callus.

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3.2. Study of the influence of different growth regulators concentrations

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As a result of studying the effect of a substrate with different content of phytohormones, it was found that megagametophytes from plus trees clones located on the Petrozavodsk SO more often formed calluses on nutrient media № 3 и 5 (Table 2) while mature seeds from Petrozavodsk park formed a cell culture on substrate № 4 (Fig. 2). It is important to note that explants collected from SO formed callus twice more often than seeds from the park. The auxin/cytokinin ratio (2:1) in the composition of nutrient medium is the most commonly used for SE initiation in conifers [6, 7, 13, et al.]. However, it was revealed in our study that the extracted from mature seeds megagametophytes predominantly formed calluses on substrates with 1:1 auxin/cytokinin ratio. It should be noted that seed population mixture was used in this experiment which contributed to a more effective assessment of nutrient media.

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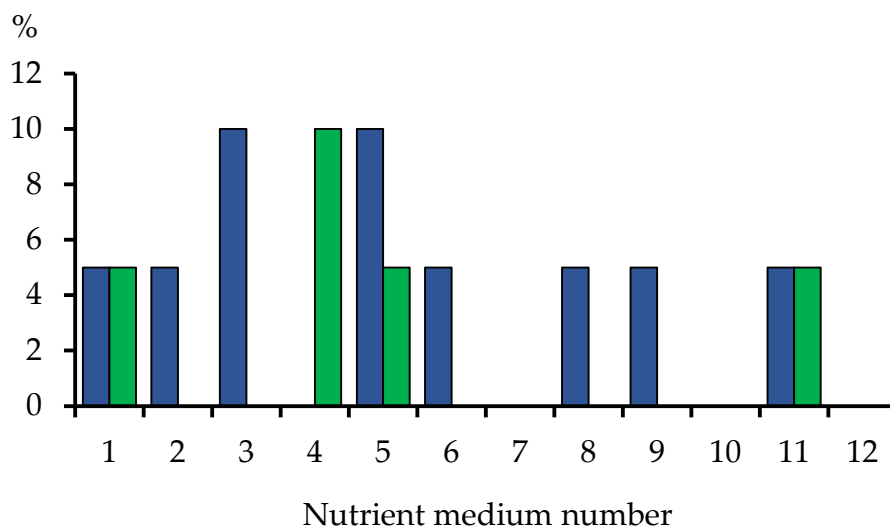


Figure 2. Frequency of callus formation on different nutrient media. Note: blue bars indicate Petrozavodsk SO, green bars – Petrozavodsk park.

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3.3. Study of the influence of content and density of the nutrient medium

It is known that the availability of water in the nutrient medium affects the development of the embryonic mass [13, 14]. Several authors have shown that stress (including water deficiency) can trigger or improve embryogenesis in recalcitrant species [15, 16]. It was found in our study that, in terms of the frequency of callus formation, the 1/2LV medium with the standard agar concentration turned out to be the most successful for *P. sylvestris* megagametophytes with mature embryos (Table 3). On the DCR- substrate, the proportion of explants (collected from trees in the natural phytocoenosis) which formed callus averages 8.33±3.3%, which is also a high value in this experiment. Analysis of the data obtained showed that the population mixture of seeds collected in the Medvezhyegorsk region of Karelia formed callus 4 times and plants 14 times more often than from megagametophytes of the Petrozavodsk park.

Table 3. Mean frequency of callus/plant formation from *Pinus sylvestris* mature seeds megagametophytes from different habitats on the nutrient media differed in composition and density.

Event / Medium, %	DCR	DCR-	MS	MS-	MSG	MSG-	1/2LV	1/2LV-
Medvezhyegorsk region								
Callus	1.67±1.7	8.33±3.3	3.33±3.4	5.0±2.8	6.67±3.97	3.33±2.4	9.67±3.3	3.33±2.4
Plant	8.33±4.1	6.67±3.1	0	0	11.67±5.6	0	6.33±2.9	0
Petrozavodsk park								
Callus	1.25±1.3	0	–	–	1.25±1.3	3.75±2.1	5.0±2.4	0
Plant	1.25±1.3	0	–	–	0	1.25±1.3	0	0

Note. Values in the table are the arithmetic mean of the frequency of callus/plant initiation±standard error.

4. Conclusions

Thus, the data obtained indicate that the use of vegetative buds and mature seeds as explants can help identify *P. sylvestris* genotypes predisposed to SE, as well as select cultivation conditions throughout the year. As part of the experiments, it was found that there are genotypes on the Petrozavodsk SO that are probably capable of forming an embryonic-suspensor mass from immature embryos. The study showed that callus formed 5 times less frequently on the DCR nutrient medium than on the 1/2LV substrate. Perhaps, when *P. sylvestris* immature embryos collected in the middle taiga phytocoenoses of Karelia are introduced into culture *in vitro*, it is necessary to use this nutrient medium to initiate SE.

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References

1. Häggman, H., Jokela, A., Krajnakova, J., Kauppi, A., Niemi, K., Aronen, T., Somatic embryogenesis of Scots pine: cold treatment and characteristics of explants affecting induction. *J. Exp. Bot.* **1999**, *50*(341), 1769-1778. doi:10.1093/jxb/50.341.1769
2. Lelu, M.A., Bastien, C., Drugeault, A., Gouez, M.L., Klimaszewska, K. Somatic embryogenesis and plantlet development in *Pinus sylvestris* and *Pinus pinaster* on medium with and without growth regulators. *Physiol. Plant.* **1999**, *105*(4), 719-728. doi:10.1034/j.1399-3054.1999.105417.x
3. Bonga, J. M., Klimaszewska, K. K., Von Aderkas, P. Recalcitrance in clonal propagation, in particular of conifers. *PCTOC* **2010**, *100*(3), 241-254. doi:10.1007/s11240-009-9647-2
4. Keinonen-Mettälä, K., Jalonen, P., Eurola, P., Arnold, S., Weissenberg, K. Somatic embryogenesis of *Pinus sylvestris*. *Scand. J. For. Res.* **1996**, *11*(1), 242-250. doi:10.1080/02827589609382933
5. Niskanen, A.M., Lu, J., Seitz, S., Keinonen, K., Von Weissenberg, K., Pappinen, A. Effect of parent genotype on somatic embryogenesis in Scots pine (*Pinus sylvestris*). *Tree Physiol.* **2004**, *24*(11), 1259-1265. doi:10.1093/treephys/24.11.1259
6. Treťyakova, I.N., Voroshilova, E.V., Shuvaev, D.N. Callusogenesis and somatic embryogenesis induction in hybrid embryos from the seeds of *Pinus sibirica*. *Russ. J. Plant Physiol.* **2014**, *61*(2), 274-280. doi: 10.1134/S1021443714020162
7. Abrahamsson, M., Clapham, D., Arnold, S.V. Somatic embryogenesis in Scots pine (*Pinus sylvestris* L.). Step wise protocols for somatic embryogenesis of important woody plants, Shri Mohan Jain, Gupta, P., Springer: Gewerbstrasse 11, 6330 Cham, Switzerland **2018**, Volume 1, pp. 123-134. doi:10.1007/978-3-319-89483-6_9
8. Raevsky, B.V., Kuklina, K.K., Schurova, M.L. Genetic and breeding assessment of Scotts pine plus trees in Karelia. *Proceedings of the Karelian Research Centre of the Russian Academy of Sciences* **2020**, *3*,45-59. doi: 10.17076/eb1163
9. Hohtola, A. Seasonal changes in explant viability and contamination of tissue cultures from mature Scots pine. *PCTOC* **1988**, *15*(3), 211-222. doi:10.1007/BF00033645
10. Gupta, P. K., Durzan, D. J. Shoot multiplication from mature trees of Douglas-fir (*Pseudotsuga menziesii*) and sugar pine (*Pinus lambertiana*). *Plant. Cell Rep.* **1985**, *4*(4), 177-179. doi:10.1007/BF00269282
11. Murashige, T., Skoog, F. A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiol. Plant.* **1962**, *15*(3), 473-497. doi:10.1111/j.1399-3054.1962.tb08052.x
12. Trontin, J-F., Aronen, T., Hargreaves, C., Montalbán, I.A., Moncaleán, P., Reeves, C., Quoniou, S., Lelu-Walter, M-A., Klimaszewska, K. International effort to induce somatic embryogenesis in adult pine trees. *Vegetative Propagation of Forest Trees NIFoS* **2016**, pp. 211-260.
13. Klimaszewska, K., Park, Y.S., Overton, C., Maceacheron, I., Bonga, J.M. Optimized somatic embryogenesis in *Pinus strobus* L. *In Vitro Cell. Dev. Biol. – Plant.* **2001**, *37*(3), 392-399. doi:10.1007/s11627-001-0069-z
14. Montalbán, I. A., Moncaleán, P. *Pinus radiata* (D. Don) somatic embryogenesis. In *Step Wise Protocols for Somatic Embryogenesis of Important Woody Plants*, Shri Mohan Jain, Gupta, P., Springer: Gewerbstrasse 11, 6330 Cham, Switzerland, **2018**, Volume 1, pp. 1-11.
15. Aderkas, P., Bonga, J. M. Influencing micropropagation and somatic embryogenesis in mature trees by manipulation of phase change, stress and culture environment. *Tree Physiol.* **2000**, *20*(14), 921-928. doi: 10.1093/treephys/20.14.921
16. Neilson, K. A., Gammulla, C. G., Mirzaei, M., Imin, N., Haynes, P. A. Proteomic analysis of temperature stress in plants. *Proteomics* **2010**, *10*(4), 828-845. doi: 10.1002/pmic.200900538