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The 2-nd International Electronic Conference on Biomolecules:  
Biomacromolecules and the Modern World Challenges

# **Structure, properties and biological activity of chitosan salts with *L*- and *D*-aspartic acid**

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Tatiana Lugovitskaya and Irina Zudina**



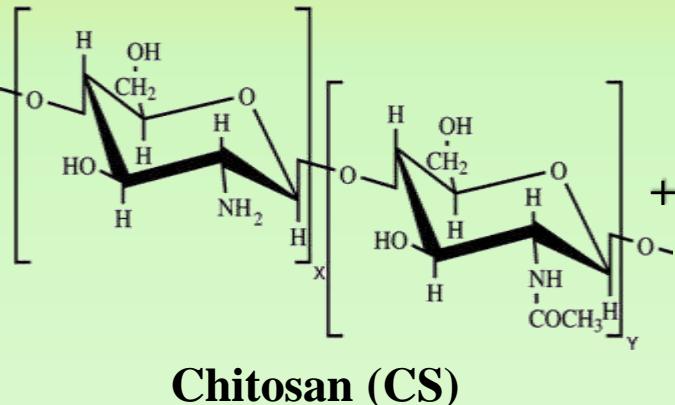
**Saratov State University, Russian Federation**

**1–15 November 2022**

# Abstract

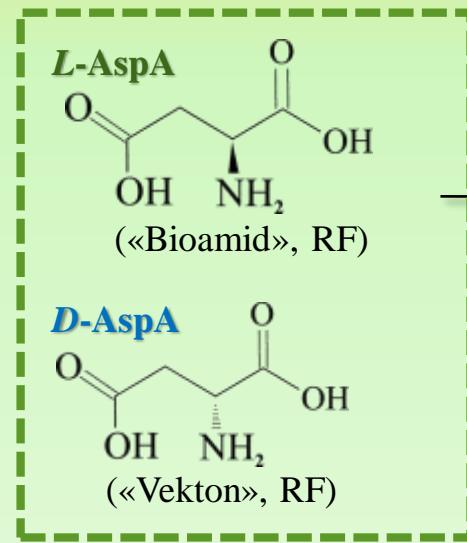
A comprehensive study of the structure, properties and biological functionality of salt chitosan complexes with *L*- and *D*-aspartic acid (AspA) was carried out. It has been established that these polymer salts differ in spatial organization, chirooptic characteristics, surface charge, and macrocoil size. In experiments *in vitro* on a wide range of biological objects (unicellular algae, planktonic crustaceans, aerobic bacterial microorganisms, cell cultures, and test plants) it was found that the chitosan salt with D-AspA exhibited the best biological activity. The results obtained confirm our hypothesis that the biological homochiral hierarchy principles are most consistent with chitosan (D-aminoglucan) derivatives with the D-antipode of the acid.

## Preparation of chitosan aspartate

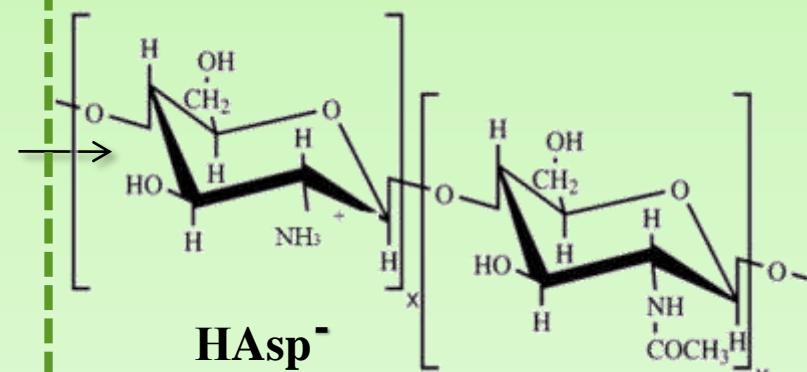


**Chitosan (CS)**

Molecular weight of 200 kDa  
Degree of deceleration DD = 80 mol.%  
("Bioprogress Ltd.", RF)



**Aspartic acid (AspA)**



**HAsp<sup>-</sup>**

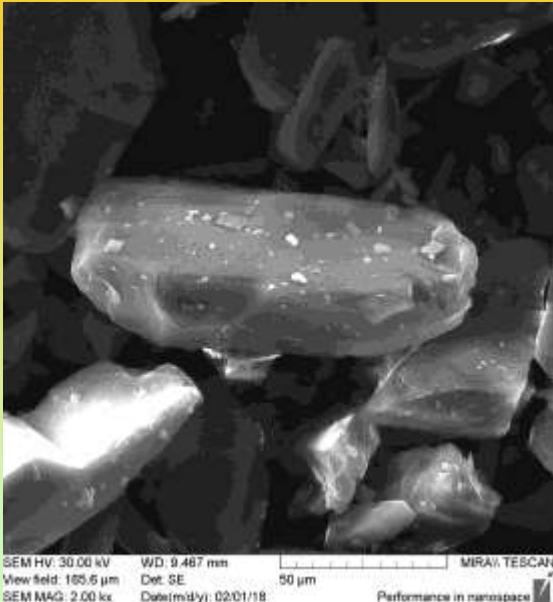
**CS·L-AspA**

or

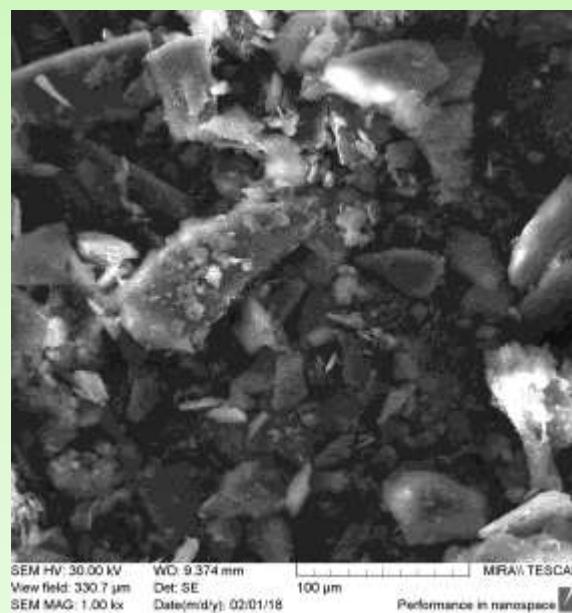
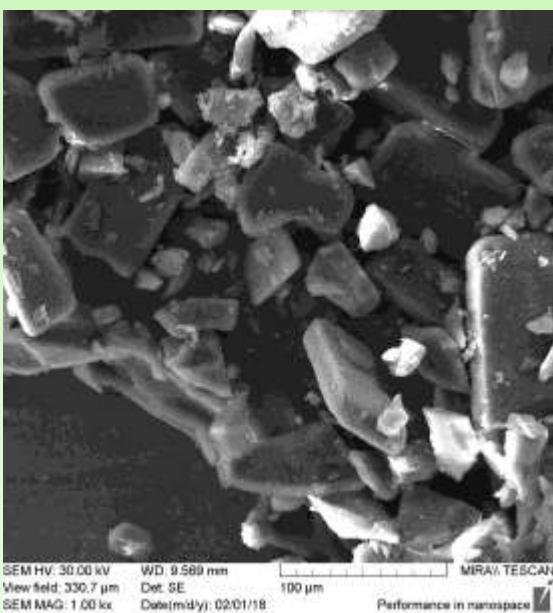
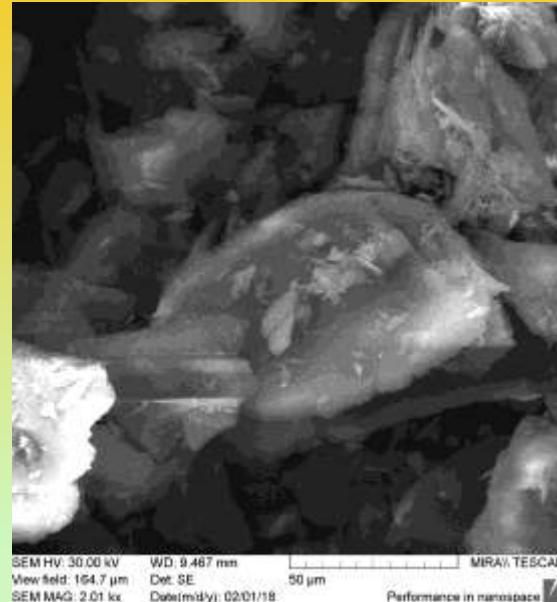
**CS·D-AspA**

# SEM photos of the powders AspA

*L*-AspA

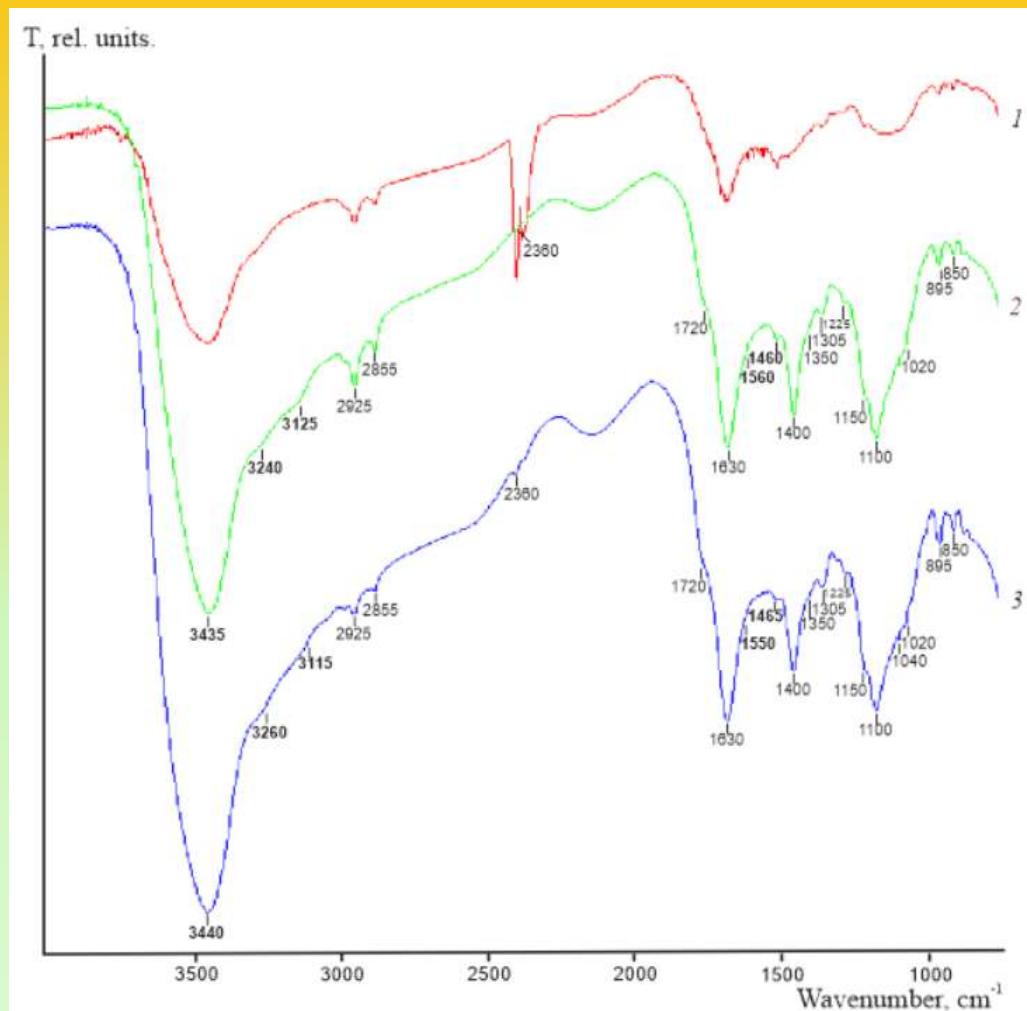


*D*-AspA



# IF spectroscopy

Oscillation types	Wave numbers $\nu$ , $\text{cm}^{-1}$	
	CS·L-AspA	CS·D-AspA
$\nu_{\text{O-H}}$	<b>3435</b>	<b>3440</b>
$\nu_{\text{N-H}}$	<b>3240</b>	<b>3260</b>
$\nu_{\text{N-H}} (\text{NH}_3^+)$	<b>3125</b>	<b>3115</b>
$\nu_{\text{as}}(\text{CH})$	2925	
$\nu_s(\text{CH})$	2855	
$\nu_{\text{as}}(\text{C-N})$	2360	
$\nu_{\text{C=O}}$ (COOH)	1720	
$\delta_{\text{C=O}}$ (Amide I)	1630	
$\delta_{\text{N-H}}$ (Amide II)	<b>1560</b>	<b>1550</b>
$\nu_{\text{C-O}}$ (COO <sup>-</sup> )	<b>1460</b>	<b>1465</b>
$\delta_{\text{as}}(\text{CH})$	1400	
$\nu_{\text{C-N}}$ (Amide III)	1350, 1305	
$\delta(\text{OH})$	1225	
	1150	
	1100	
$\nu(\text{C-C}), \nu(\text{C-O}),$ $\nu(\text{C-N}), \delta(\text{C-H})$	1020	1040, 1020
Glucopyranose ring	895	

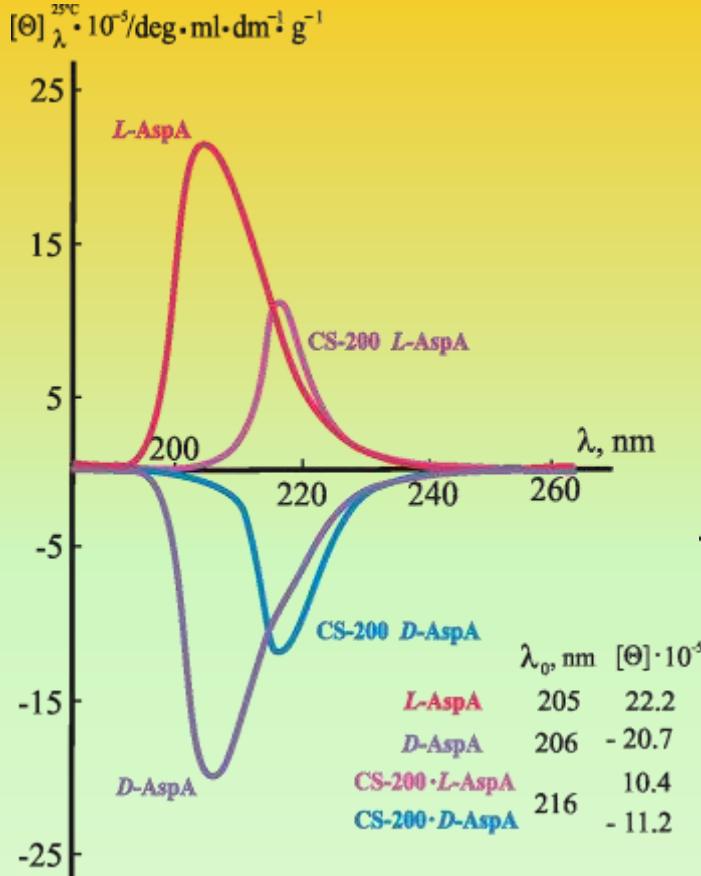


IR spectra of CS powders (1), CS·L-AspA (2) and CS·D-AspA (3)

Vertex 70v vacuum FTIR spectrometer  
(Billerica, MA, USA)

# Chirooptic properties of solutions

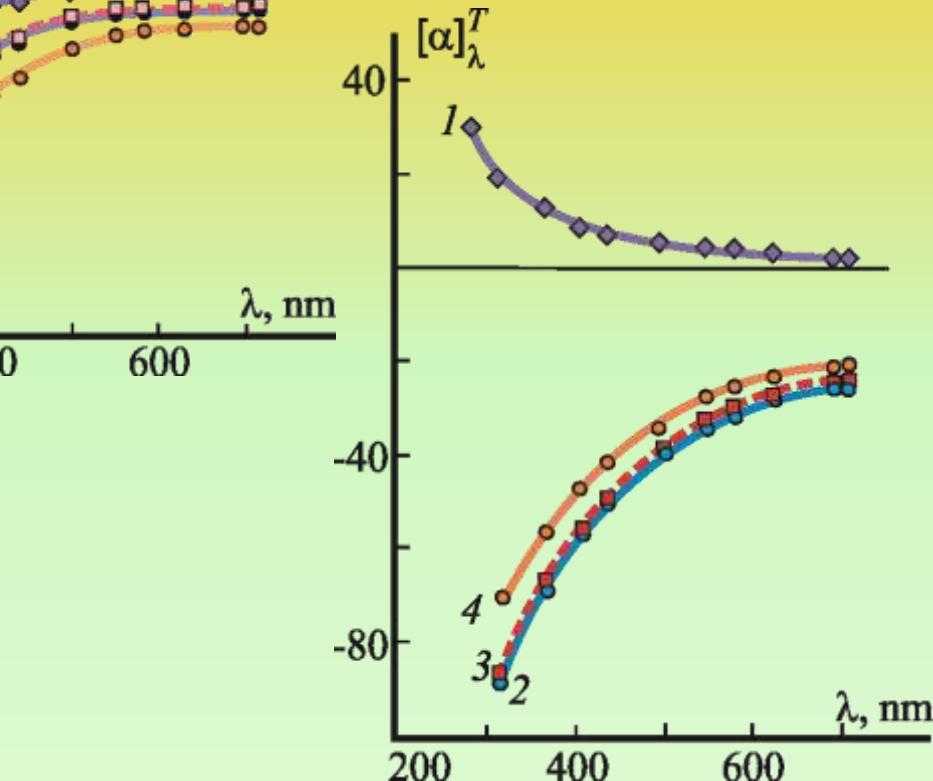
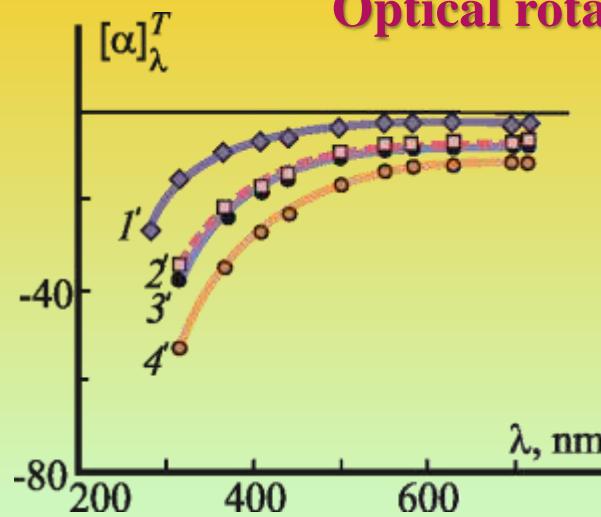
## Circular dichroism (CD)



CD spectra of aqueous solutions of *L*-AspA (1), *D*-AspA (2), CS in *L*-AspA (3), and CS in *D*-AspA (4); the molar ratio  $[\text{AspA}]/[\sim\text{NH}_2] = 0.85$  mol/mol.

Chirascan™ Spectrometer (Applied Photophysics Ltd, USA)

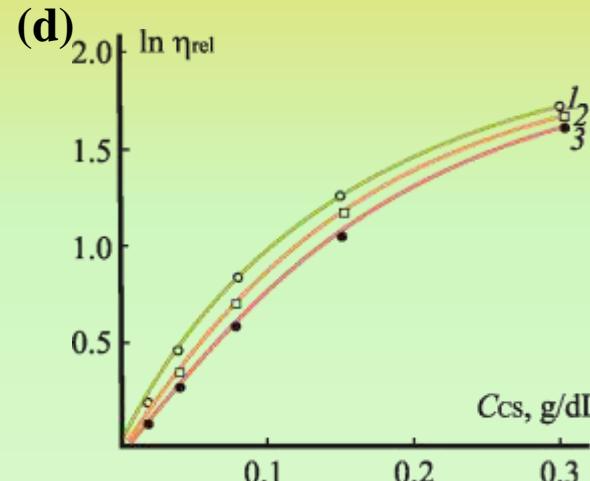
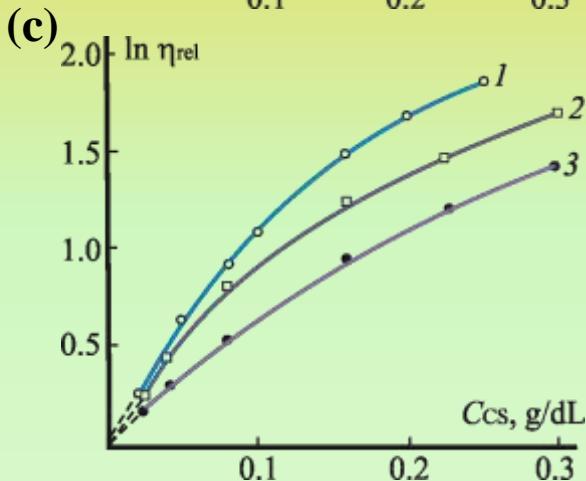
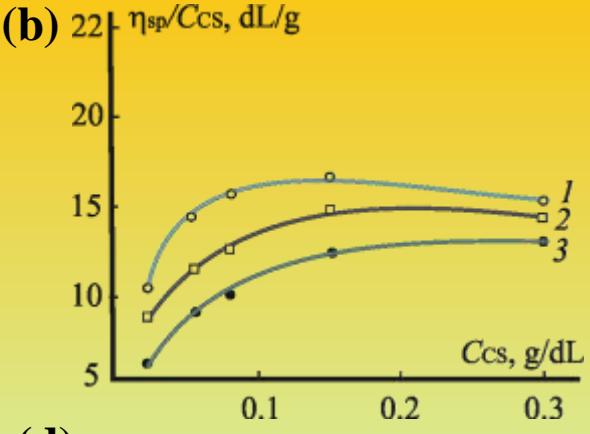
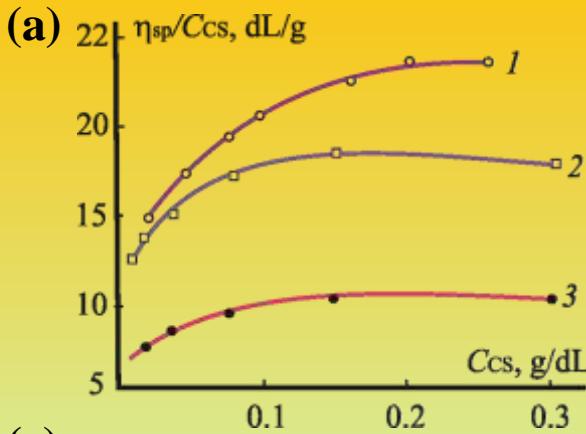
## Optical rotation dispersion (ORD)



ORD curves for aqueous solutions of *L*-AspA (1), *D*-AspA (1'), CS in *L*-AspA (2–4), and CS in *D*-AspA (2'–4'); the molar ratio  $[\text{AspA}]/[\sim\text{NH}_2] = 0.85$  (2, 2'), 1.0 (3, 3') and 2.0 mol/mol (4, 4').

Automatic SPU-E spectropolarimeter (RF)

# Hydrodynamic properties of solutions



Concentration dependences of the viscosity number (a, b) and the logarithm of the relative viscosity (c, d) of CS solutions in L-AspA (a, c) and D-AspA (b, d) with  $C_{AspA} = 1.5 \cdot 10^{-3}$  (1),  $3.0 \cdot 10^{-3}$  (2) and  $6.0 \cdot 10^{-3}$  mol/dL (3).

Ubbelohde capillary viscometer  
(Cannon Instrument Company", USA)  
 $d = 0.56$  mm

$C_{AspA} \cdot 10^3$ , mol/dL	[ $\eta$ ], dL/g		$R_{ef}$ , HM	
	CS·L- AspA	CS·D- AspA	CS·L- AspA	CS·D- AspA
1.5	12.5	8.8	73	65
3.0	11.1	8.4	71	64
	3.5*	2.1*	48*	41*
6.0	7.5	6.8	62	60

\*  $I = const$  (0.17 M NaCl)

Limit number of viscosity

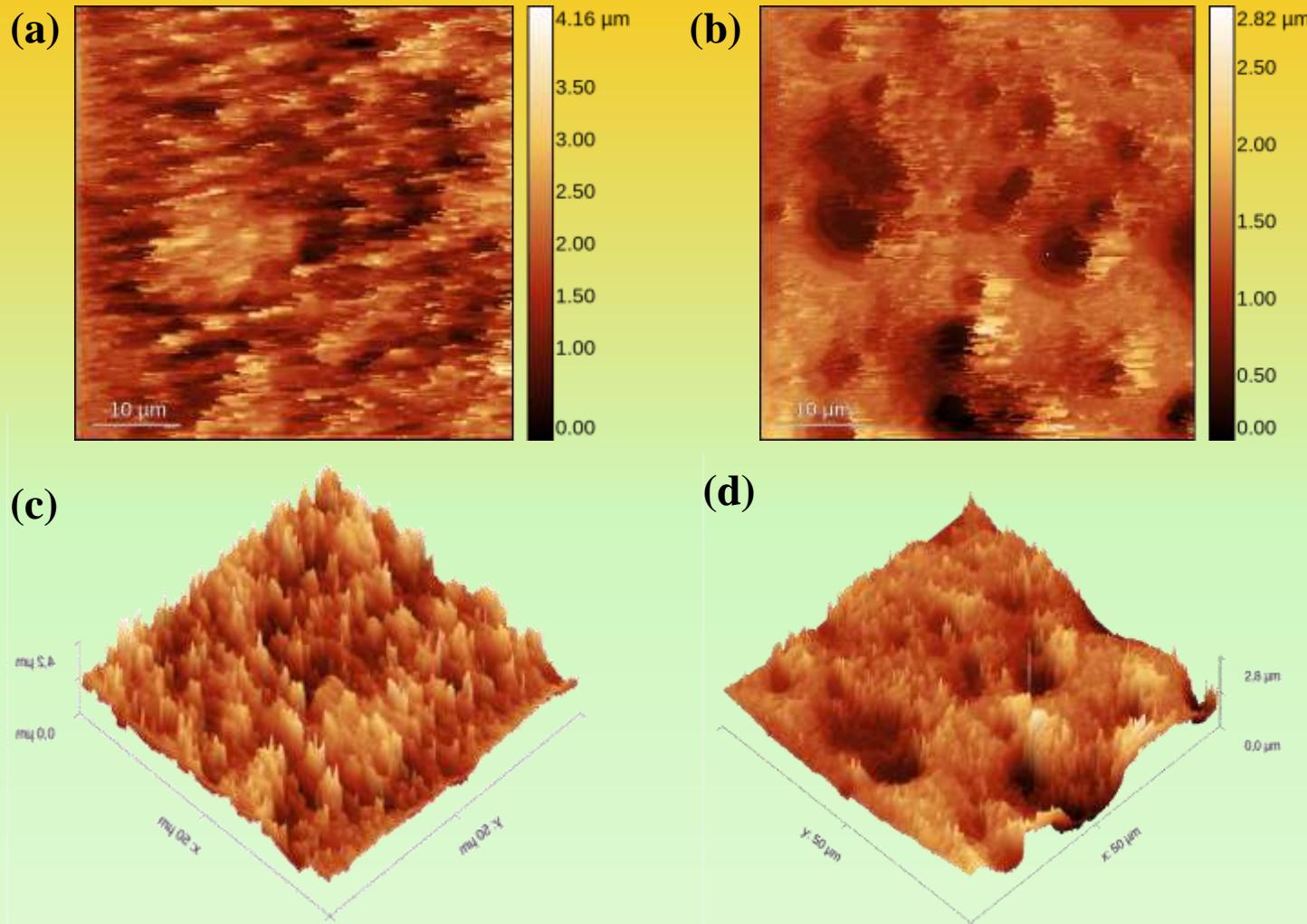
$$[\eta] = \partial \ln \eta_{rel} / \partial C_{CS}$$

[Wolf B.A. // Macromolecular Rapid Communications, 2007. Vol.28. No.2. P.164-170]

Effective radius of a macromolecular coil

$$R_{ef} = \sqrt[3]{\frac{3[\eta] \bar{M}_n}{10\pi N_a}}$$

# AFM of the surface of hydrogel plates



AFM images of the surface of hydrogel plates made of CS•L-AspA (a, c) and CS•D-AspA(b, d) obtained at the molar ratio of [AspA]/[~ -NH<sub>2</sub>] = 0.85 mol/mol.

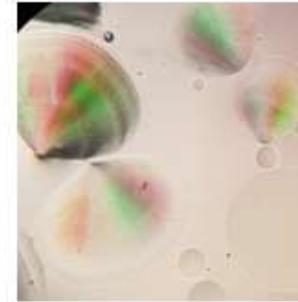
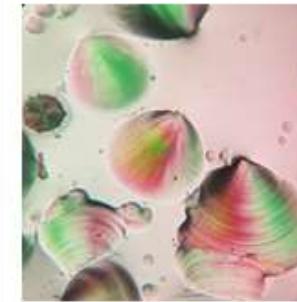
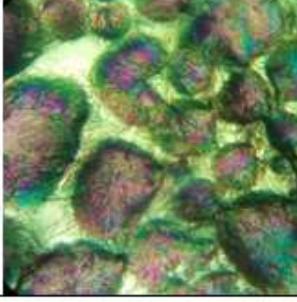
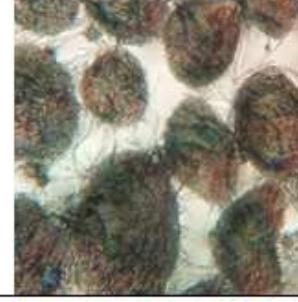
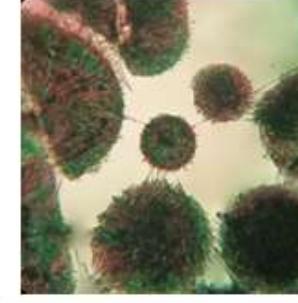
# Multi-angle dynamic light scattering

Parameter	$C_{CS} \cdot 10^3$ , monomol/dL	Temperature, °C					
		25			37		
		Molar ratio [AspA]/[~NH <sub>2</sub> ], mol/monomol					
		0.85	1.0	2.0	0.85	1.0	2.0
<b>CS·L-AspA</b>							
$\zeta$ -potential, mV	0.9	34.2	39.3	40.5	37.1	38.2	38.9
	2.4	39.3	39.3	35.6	33.3	37.9	35.9
	4.7	37.0	39.6	—	29.2	35.4	—
Wall $\zeta$ -potential, mV	0.9	51.0	51.8	41.5	52.2	53.6	49.5
	2.4	44.9	38.9	42.5	38.6	41.5	48.6
	4.7	37.2	29.6	—	28.6	37.3	—
Conductivity, mS/cm	0.9	0.40	0.39	0.51	0.47	0.46	0.61
	2.4	0.71	0.67	0.84	0.85	0.80	1.01
	4.7	1.30	1.24	—	1.56	1.50	—
<b>CS·D-AspA</b>							
$\zeta$ -potential, mV	0.9	36.5	32.3	33.7	38.0	31.0	36.2
	2.4	37.9	39.4	42.7	37.6	36.4	37.6
	4.7	42.3	33.3	—	34.8	35.5	—
Wall $\zeta$ -potential, mV	0.9	36.2	52.6	59.5	45.5	46.7	47.9
	2.4	37.3	40.2	36.0	39.0	39.0	42.9
	4.7	27.9	26.3	—	21.9	29.9	—
Conductivity, mS/cm	0.9	0.44	0.39	0.51	0.52	0.46	0.61
	2.4	0.83	0.76	0.85	0.99	0.76	1.02
	4.7	1.68	1.19	—	2.02	1.44	—

Zetasizer Ultra/Pro (Malvern Panalytical, England)

# Polarizing microscopy

Kinetics of crystallization of *L*-menthol in aqueous solutions of CS·*L*-AspA and CS·*D*-AspA

$C_{Cs} \cdot 10^3, \text{monomol/dL}^*$	CS· <i>L</i> -AspA		CS· <i>D</i> -AspA	
	Extractive crystallization time, min			
	5	30	5	30
1.8				
3.5				
4.7				

\*  $[\text{AspA}] / [\sim\text{-NH}_2] = 0.85 \text{ mol/monomol}$

Polarization microscope LaboPol-2 (RF), magnification  $\times 10$

# Complex formation with metal ions

System	Multiplicity of breeding, times	pH	Complex formation ***, %
<i>L</i> -AspA*	10	5.5	47.9±0.7
	100	6.4	21.4±1.1
	1000	6.7	12.4±0.9
<i>D</i> -AspA*	10	5.6	39.0±1.2
	100	6.5	10.1±0.8
	1000	6.7	6.40±0.7
CS· <i>L</i> -AspA**	10	5.9	89.6±1.4
	100	6.8	62.6±1.3
	1000	6.9	37.4±2.1
CS· <i>D</i> -AspA**	10	5.7	97.6±1.2
	100	6.6	92.2±0.9
	1000	6.8	79.8±1.7

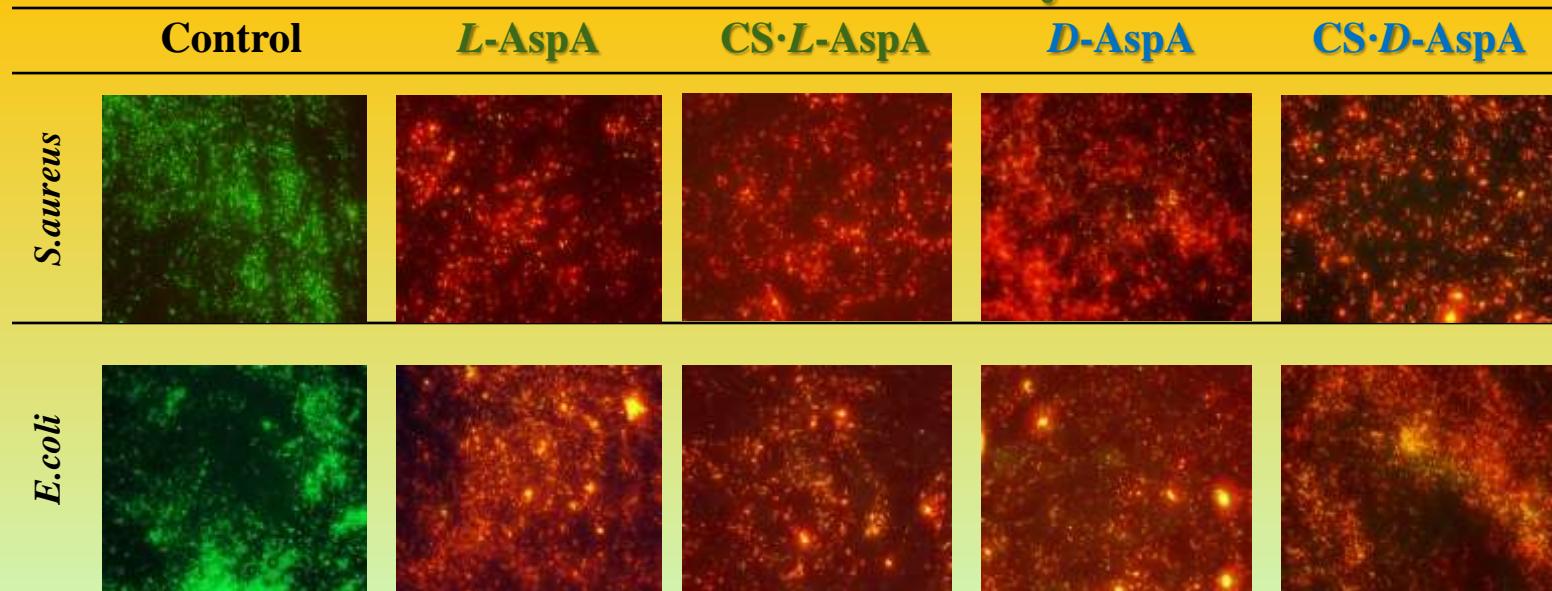
\*  $C_{\text{AspA}} = 0.8 \cdot 10^{-3} \text{ mol/dL}$

\*\*  $C_{\text{CS}} = 0.8 \cdot 10^{-3} \text{ monomol/dL}$ ,  $[\text{AspA}] / [\sim\text{NH}_2] = 1.0 \text{ mol/monomol}$

\*\*\* Complex formation was assessed by the change in the chlorophyll fluorescence intensity of *S. quadricauda* microalgae cultivated in Prat's medium,  $t = 4$  days.

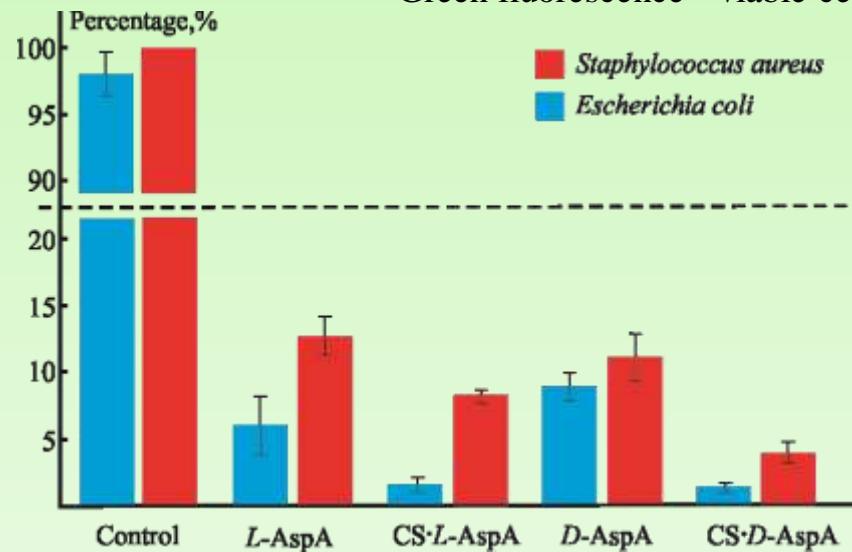
Reagent	Concentration, g/dm <sup>3</sup>
Nutrient medium Prat	
Potassium nitrate, KNO <sub>3</sub>	0.1
Potassium phosphate disubstituted, K <sub>2</sub> HPO <sub>4</sub> ·3H <sub>2</sub> O	0.01
Magnesium sulfate, Mg <sub>2</sub> SO <sub>4</sub> ·7H <sub>2</sub> O	0.01
Iron chloride, FeCl <sub>3</sub> ·6H <sub>2</sub> O	0.001
Bioobject	
<i>S. quadricauda</i>	30 thousand cells/cm <sup>3</sup>

# Antibacterial activity



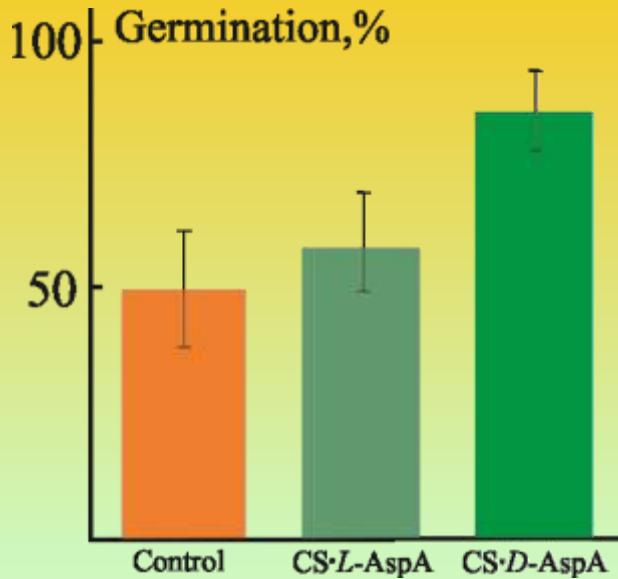
Fluorescent microscopy of cultures of *S. Aureus* 209 P and *E. Coli* 113-13 cultures in meat-peptone broth without (control) and with the addition of aqueous solutions of *L*-AspA, *D*-AspA, CS·*L*-AspA and CS·*D*-AspA; the molar ratio [AspA]/[~NH<sub>2</sub>] = 0.85 mol/monomol; 37°C, *t* = 24 hours.

Green fluorescence - viable cells; red fluorescence - dead cells. Magnification: x 400.



Number of living cells in cultures of *S. aureus* 209 P and *E. coli* 113-13 cultures after 24 hours of growth in meat-peptone broth without (control) and with the addition of aqueous solutions of *L*-AspA, *D*-AspA, CS·*L*-AspA and CS·*D*-AspA; the molar ratio [AspA]/[~NH<sub>2</sub>] = 0.85 mol/monomol; 37°C.

# Biostimulatory activity (laboratory conditions)



Germination of test seeds of *Lunum usitatissimum* in laboratory conditions on aqueous extract from the initial soil-soil (control) and from the soil-soil after biodegradation ( $t = 40$  days) of hydrogel plates made from CS·L-AspA and CS·D-AspA obtained with the molar ratio [AspA]/[ $\sim\text{NH}_2$ ] = 1.0 mol/monomol.

Photo of germinated test-seed of *Linum usitatissimum*



CS·L-AspA

$\text{H}_2\text{O}$

CS·D-AspA

# Biostimulatory activity (field conditions)

Sample test plants	Irrigation liquid	Time, days	Germination (field), %	Growth stimulating activity	
				Shoot height, mm	Mass of shoots, kg
Radish <i>Raphanus sativus</i>	$\text{H}_2\text{O}$	6	64±2	2.5-3.0	—
		10	76±4	3.8-4.2	—
		25	—	95-100	1.03±0.04
	CS·L-Asp + $\text{H}_2\text{O}^*$	6	90±3	3.5-4.0	—
		10	93±2	6.0-8.0	—
		25	—	105-115	1.36±0.03
	CS·D-Asp+ $\text{H}_2\text{O}^*$	6	92±2	3.4-4.5	—
		10	96±2	6.5-8.3	—
		25	—	110-120	1.42±0.06
Mustard <i>Sinapis alba</i>	$\text{H}_2\text{O}$	6	20±5	4.5-5.0	—
		10	75±5	7.5-8.5	—
		25	—	130-140	0.61±0.03
	CS·L-Asp+ $\text{H}_2\text{O}^*$	6	40±4	5.0-7.0	—
		10	85±3	10.0-10.5	—
		25	—	150-170	0.82±0.02
	CS·D-Asp+ $\text{H}_2\text{O}^*$	6	60±3	6.0-8.0	—
		10	90±1	11.5-12.5	—
		25	—	170-190	0.85±0.01



\*  $C_{\text{CS}} = 0.2 \cdot 10^{-3}$  monomol/dL,  
 $[\text{AspA}] / [\sim\text{NH}_2] = 1.0$  mol/monomol

Photo of a bed of radish (*Raphanus sativus*) and mustard (*Sinapis alba*) 25 days after planting in the field conditions (open ground), watered and sprinkled with a solution of CS·L-AspA and CS·D-AspA with  $C_{\text{CS}} = 0.2 \cdot 10^{-3}$  monomol/dL and the molar ratio composition  $[\text{AspA}] / [\sim\text{NH}_2] = 1.0$  mol/monomol.

*Raphanus  
sativus*

*Sinapis  
alba*

# Biostimulatory activity (field conditions)

Radish *Raphanus sativus*



H<sub>2</sub>O

CS·L-AspA

CS·D-AspA

Mustard *Sinapis alba*



H<sub>2</sub>O

CS·L-AspA

CS·D-AspA

Photos of shoots of radish (*Raphanus sativus*) and mustard (*Sinapis alba*), cut off 25 days after planting in open ground, watered and sprinkled with a solution of CS·L-AspA and CS·D-AspA with  $C_{CS} = 0.2 \cdot 10^{-3}$  monomol/dL and the molar ratio [AspA]/[~-NH<sub>2</sub>] = 1.0 mol/monomol.

*Thank you for your attention!*

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