

Proceeding Paper

Nepeta nuda L. Plant Extract Preserves the Morphology of Red Blood Cells Subjected to Oxidative Stress †

Stefani Petrova ^{1,2}, Nikol Mazhdrakova ^{1,2,3}, Svetla Todinova ¹, Velichka Strijkova ^{1,4}, Miroslava Zhiponova ² and Sashka Krumova ^{1,*}

¹ Institute of Biophysics and Biomedical Engineering, Bulgarian Academy of Sciences; stefani.v-petrova@abv.bg (S.P.); nikolmazhdrakova@gmail.com (N.M.); todinova@abv.bg (S.T.); vily_strij@abv.bg (V.S.)

² Sofia University “St. K. Ohridsky”; zhiponova@biofac.uni-sofia.bg

³ Institute of Plant Physiology and Genetics, Bulgarian Academy of Sciences

⁴ Institute of Optical Materials and Technologies “Acad. Jordan Malinowski”

* Correspondence: sashka@bio21.bas.bg

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Abstract: We show that extract from catmint (*Nepeta nuda* L.) flowers is not hemotoxic and does not alter erythrocytes morphology. H₂O₂-induced oxidative stress leads to increase in lipid peroxidation, accompanied by reduction in the number of biconcave cells and increase in the amount of echinocytes. Pre-treatment of erythrocytes with extract does not reduce the lipid peroxidation level, however it results in partial restoration of the relative abundance of biconcave cells and respective reduction in echinocytes quantity. Our data reveal the concentrations at which the examined extract exhibits protective effect on erythrocytes morphology in the condition of H₂O₂-induced oxidative stress.

Keywords: plant extract; red blood cells; oxidative stress

1. Introduction

Medicinal plant extracts are widely explored for their beneficial effects on human health. A number of previous reports highlights the protective effect of plant extracts against oxidative damage, including on erythrocytes [1]. The catmint *Nepeta nuda* L. (Lamiaceae) was reported to exhibit antioxidant effects due to its phenolic compounds and iridoids [2], however to the best of our knowledge, no studies have been performed so far on the effect of catmint extract on red blood cells (RBC).

2. Materials and Methods

Plant extract (PE) from catmint flowers was prepared at 60 °C using water solvent and maceration [2]. After lyophilization the dry substance was stored at –20°C until further use. For all experiments aqueous solutions of 0.01–1 mg/L PE and 0.1 mM Trolox (TX) antioxidant (Acros Organics Fisher Scientific, water-soluble analogue of the vitamin E) were utilized.

Donor blood was used for preparation of RBC as described in Langari et al. [3]. Experiments were performed in three replicates. Samples with concentration 1 mg Hb/mL were incubated for 1 h at 37 °C with PE with concentration 1, 0.1 and 0.01 mg/mL, and 0.1 mM TX. Untreated RBC were used as control. Hemolysis was determined spectrophotometrically based on hemoglobin absorption at 543 nm corrected for non-specific absorption at 650 nm. Full (100%) hemolysis was achieved via 100 times dilution of RBC in distilled water.

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In vitro lipid peroxidation was induced by incubation of control and PE/TX treated samples with 1 mg Hb/mL and 0.8 mM H₂O₂ for 2 h. The lipid peroxidation reaction was evaluated via thiobarbituric acid reactive substances (TBARS) according to Gilbert et al. [4], and the absorption at 532 nm was corrected for non-specific absorption at 650 nm. The reaction was stopped with 150 μ L 10 mM ethylenediaminetetraacetic acid before the absorption readings.

RBC morphologic evaluation on air-dried smears was performed by means of optical microscopy (3D Optical profiler, Zeta-20, Zeta Instruments, Milpitas, CA, USA).

Data are represented as mean values and standard deviation (SD).

3. Results and Discussion

As a first step in our study, we probed the hemolytic effect of the studied PE and TX on RBC. The evaluation of the hemolysis level after 1 h of PE incubation in the selected concentration range resulted in similar cell lysis for the control and PE/TX treated cells of ca. 6–11% (Figure 1a). The morphology of the treated cells was similar to the one of control RBC—the dominant cell species was the biconcave one accounting for 74–85% followed by echinocytes (11–26%) and spherocytes (0–3%) (Figure 1b).

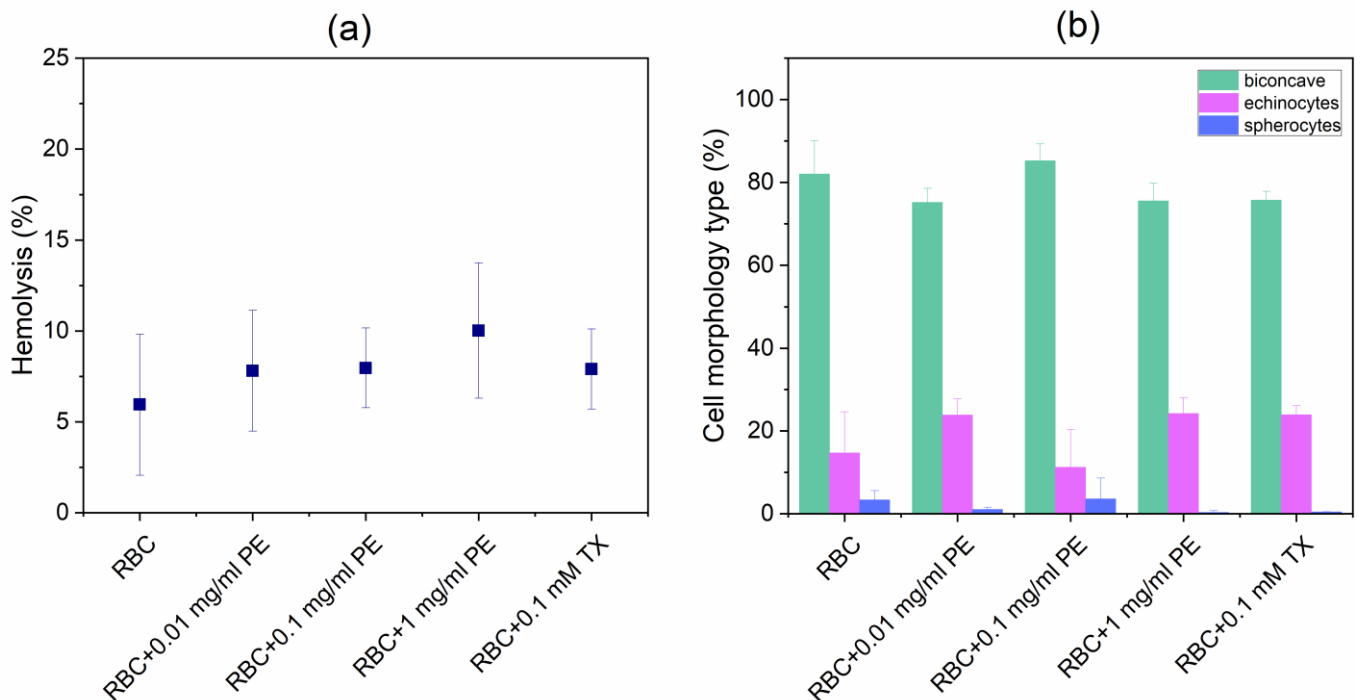


Figure 1. Hemolysis (a) and relative abundance of different morphological types (b) of RBC subjected to 1 h treatments with different concentrations of catmint plant extract (PE) and 0.1 mM Trolox (TX). Mean \pm SD.

Next, we evaluated the PE effect in the condition of H₂O₂-induced oxidative stress. The applied incubation with H₂O₂ resulted in a five-fold increase in TBARS level in RBC (designated RBCoxi). The incubation with 0.01 and 0.1 mg/mL PE, and 0.1 mM TX had no additional effect, while 1 mg/mL PE induced further increase in TBARS values (Figure 2a). Lipid peroxidation led also to four-fold decrease in the relative abundance of biconcave RBC and five-fold increase in echinocytes population, while the number of spherocytes was not substantially affected, as compared to RBCoxi sample. All three applied concentrations of PE, as well as TX, led to an increase in the number of biconcave cells by 1.5–2.1 times and reduction of echinocytes by 0.6–0.8 times under the conditions of oxidative stress (Figure 2b).

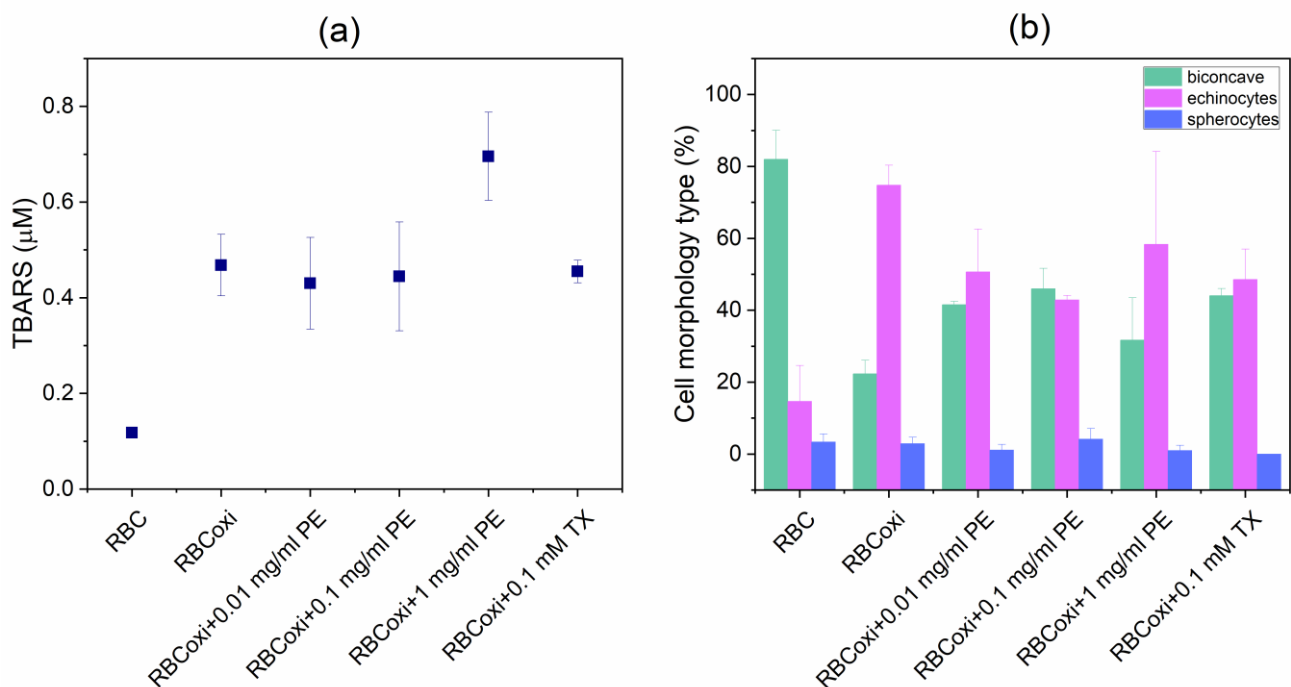


Figure 2. TBARS amount per mg Hb (a) and relative abundance of different morphological types (b) of RBC subjected to 1 h treatments with different concentrations of catmint plant extract (PE) and 0.1 mM trolox (TX) followed by 2 h in vitro oxidation by H_2O_2 . Control RBCs are not exposed to either PE, TX, or H_2O_2 treatment, while RBCoxi are cells subjected to in vitro H_2O_2 oxidation. Mean \pm SD.

The presented data clearly show that under our experimental conditions, PE itself has no detrimental effect on RBC features. However, in the conditions of in vitro H_2O_2 oxidation PE does not express any protective effect towards the formation of TBARS and even induces further increase in their values at the highest tested concentration. Surprisingly TX, a compound with well-established antioxidant properties, also did not reduce TBARS, which might be due to the short incubation period and/or the utilized concentration in the applied experimental protocol. Indeed, Antosik et al. [5] demonstrated a dose- and time-dependent effect of TX in long term stored RBC on the membrane integrity, lipid peroxidation and cellular morphology, with clear protective function only after 20 days of storage in TX supplemented medium.

A clear protective effect of PE and TX was observed in the morphological features of oxidized RBC, with a similar extent for the 0.01–1 mg/mL PE range and 0.1 mM TX. This important finding strongly suggests that PE stabilizes RBC shape, which is essential for their functioning and resistance to different pathologies and ageing [6,7]. The mechanism of this process will be the subject of our future studies.

4. Conclusions

Our data showed that incubation of RBC with 0.01–1 mg/mL aqueous catmint extract exhibits a protective effect on cell morphology in the conditions of H_2O_2 -induced oxidative stress.

Author Contributions:

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Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki, and approved by the Ethics Committee of Institute of Biophysics and Biomedical Engineering, Bulgarian Academy of Sciences (protocol code 1271ND).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

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Conflicts of Interest: The authors declare no conflict of interest.

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