



Proceedings

# Amelioration of Glioblastoma multiforme by the combination of Simulated microgravity and oncolytic viral therapy †

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Abstract: Glioblastoma multiforme (GBM) is the most common aggressive malignant primary brain tumor afflicting approximately 3.19 per 100,000 persons in the United States with an incidence 1.6 times higher in males compared to females. GBM usually arises from the glial cells known as astrocytes and it is commonly located in the supratentorial region (cortical lobes),usually affecting the frontal lobe. A unique feature of this tumor is its rapid local growth and spread making the prognosis very poor with a 5-year survival rate less than 5 %. Treatment of GBM remains challenging. Multiple therapeutic interventions are used for GBM including surgical resection of the tumor, radiotherapy and chemotherapy. Other experimental methods for the treatment of GBM include immune therapy, gene therapy, simulated microgravity therapy, and oncolytic viral therapy. We propose a combination therapy of simulated microgravity using a clinostat-based three-dimensional culture system with an oncolytic viral therapy using an autonomous rat parvovirus H-1 (H-1PV). Our hypothesis combines the beneficial effects of simulated microgravity and oncolytic viral therapy to lyse tumor cells through induction of apoptosis, decreased cell proliferation and or induction of an immune response. This proposal provides the foundations to construct novel breakthroughs in the treatment of GBM.

**Keywords:** Brain tumor; clinostat; Epidermal Growth Factor Receptor (EGFR),incidence; mutations; parvovirus H1; primary malignant brain tumors; EGFR mutations; Parvovirus H1; Chromosomal loss, Genetically modified viruses; Suicide genes; Neurosurgery; Radiotherapy; Clinical Neurology.

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Abbreviations:

EGFR, Epidermal growth factor receptor; GBM, Glioblastoma multiforme; INF, Interferon; RIG-1, Retinoic acid-inducible gene 1; TNF, Tumor necrosis factor; TRAF3, TNF receptor-associated factor 3; TRAIL, TNF-related apoptosis-inducing ligand

#### 1. Introduction

Glioblastoma multiforme is the most common aggressive malignant primary brain tumor affecting approximately 3.19 per 100,000 persons in the United States with an incidence 1.6 times higher in males compared to females. This is the highest incidence among malignant brain tumors. Incidence is highest in the northeast and lowest in the south-central region of the United States. Whites have the highest incidence rates for GBM.GBM is more common in males compared to females(3.97 vs. 2.53).(1)

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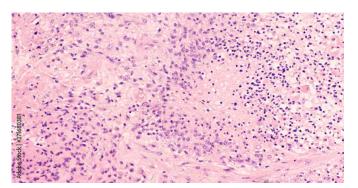
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Med. Sci. Forum **2023**, 3, × 2 of 7



**Figure 1.**: MRI showing Glioblastoma in the left frontal lobe of the brain.. Reproduced with permission from. <a href="https://www.shutterstock.com/image-photo/mri-brain-show-left-frontal-gliblastoma-1080095912">https://www.shutterstock.com/image-photo/mri-brain-show-left-frontal-gliblastoma-1080095912</a>.

GBM usually arises from the glial cells known as astrocytes and it is commonly located in the supratentorial region (cortical lobes), usually affecting the frontal lobe. It is multiformed grossly. Microscopically gross transverse sections how areas of hemorrhage and necrosis, pleomorphic nuclei and cells, pseudopalisading necrosis, and microvascular proliferation .(2) A unique feature of this tumor is its rapid local growth and spread making the prognosis very poor with a 5-year survival rate less than 5 %.(1)



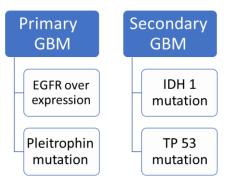
**Figure 2.** Microscopic image showing histology of a glioblastoma multiforme (GBM). Necrosis and vascular proliferation are diagnostic features of this high grade malignant tumor. Reproduced with permission from https://stock.adobe.com/search?load\_type=search&native\_visual\_search=&similar\_content\_id=&is\_recent\_search=&search\_type=usertyped&k=glioblastoma+multiforme&as-set\_id=271680381.

Multiple risk factors have been linked to GBM such as exposure to radiation, a weak immune system, and increased age. The median age of diagnosis of 64 years. It is uncommon in children accounting only approximately 3% of all brain and CNS tumors reported among individuals aged 0 to 19 years. Other risk factors include high socioeconomic status. Decreased susceptibility to allergy and the use of anti-inflammatory medications have been linked to increased risk for GBM. (1)

Several genetic and molecular mechanisms have been identified to play a role in the development of GBM. Primary *de novo* GBM without evidence of a less malignant precursor is associated with Epidermal Growth Factor Receptor (EGFR)over-expression, pleitrophin mutation, and loss of chromosome 10. Secondary GBM arising from a low grade astrocytoma or anaplastic astrocytoma is associated with IDH1 mutations, TP53

Med. Sci. Forum **2023**, 3, × 3 of 7

mutations causing alpha synclean protein over expression; the same protein involved in Parkinson's disease, and chromosome 19q loss (3),(4),(5),(6)



**Figure 3.** :Schematic diagram to represent the genetic and molecular mechanisms of primary and secondary glioblastoma multiforme (GBM). GBM: Glioblastoma multiforme; EGFR: Epidermal Growth Factor Receptor (EGFR).

Treatment of GBM remains challenging .Multipletherapeutic interventions are used for GBM including surgical resection of the tumor, post operative radiotherapy, and chemotherapy. The extent of resection is determined after assessing the preoperative prognosis of the case and the location of the tumor. Since GBM is a highly aggressive tumor, total surgical resection would lead to permanent neurological deficits especially if the tumor is located in critical areas (cortical areas) which is commonly the case (7). Survival benefits have been reported with postoperative radiation therapy to doses of 5,000–6,000 (8). Common chemotherapeutic agents used for GBM include temozolomide which can be given concomitant with radiotherapy and after it. Bevacizumab can be used along with temozolomide (TMZ) to improve progression free survival (9). Despite all these therapeutic interventions, GBM has a very poor prognosis with a 5-year survival rate less than 5 %, hence novel interventions are required for treatment of this highly malignant tumor.

# 2. Methods- the Hypothesis

We hypothesize that the growth of malignant glioma cells will be aborted by the application of a combination of simulated microgravity and oncolytic viral therapy.

Malignant glioma cells (Takeda, 2009) in Dulbecco's modified medium supplemented by penicillin (100 units/ml), streptomycin (100 micrograms/ml), 10% fetal bovine serum, white blood cells, granulocyte-monocyte colony stimulating factor (GMSCF), and parvovirus H1 at 37  $^{\circ}$  C with a humidified 5% CO2 atmosphere will be placed in a simulated microgravity using a clinostat-based three-dimensional (3D) culture system. The necrosis of malignant glioma cells week by verified by inverted contrast phase and quantum microscopy, flow cytometry mitochondrial membrane potentials. After two weeks of this intervention, all malignant glioma cells will be dead.

## 3. Results and discussion

Evidence from in vitro, animal and clinical studies were collected and evaluated. The data described in the next slides suggest that our hypothesis could provide a new therapy for glioblastoma multiforme. We aim to conduct an experiment to confirm this hypothesis and, if confirmed, to conduct clinical trials:

## A) Simulated microgravity effect on tumor cells:

## 1. -Simulated microgravity effects on Glioblastoma multiforme:-

Multiple studies have been conducted by Kentucky State University to examine the effect of microgravity during spaceflights on different tumor cells including Glioblastoma multiforme and the results showed that microgravity causes decreased cell proliferation,

Med. Sci. Forum **2023**, 3, × 4 of 7

decreased secretory activity, and induction of apoptosis. Simulated microgravity can be produced on earth by floating or using a device called 3D Clinostat to generate gravitational waves in 3 dimensions resulting in an environment with an average of  $10^{-3}$  G (10),(11)

A study was conducted in Japan to examine the effects of simulated microgravity using a 3D clinostat on cell proliferation of Glioblastoma multiforme, mitochondrial activity and sensitivity of tumor cells to cisplatin; a chemotherapy drug. Various cell lines were used in this study, including the D54MG (human glioma; wild p53), U251MG (human glioma; mutant p53), and T98G (human glioma; mutant p53) cell lines. The samples were divided into 2 groups; C group; cells under 100 G force and CL group; cells incubated in 1 G force generated by a 3D clinostat. An inverted phase contrast microscope was used to examine the morphological changes in tumor cells. The fluorescent dye rhodamine123 was used to measure the mitochondrial membrane potential. A Define first FACS caliber flow cytometer was used to measure cell cycle distribution. The results of this experiment showed that after 3 days, simulated microgravity induces Glioblastoma cells growth inhibition by worsening mitochondrial activity and increasing the sensitivity of tumor cells to cisplatin (12)

# 2 Simulated Microgravity effects on Thyroid Cancer cells:

Different ESA ground-based facilities, including the RPM and the 2D and 3D clinostat S-g, have been used to expose thyroid cancer cells. These exposures have resulted in a variety of changes in the exposed TCC, including early changes to the cytoskeleton, ECM, focal adhesion molecules, proliferation, the rate of apoptosis, migration, and growth ([13, 14, 15, 16, 17, 18) The development of MCS was the main discovery. After varying exposure times, the examined TCC and cells from other cancers (such as breast carcinoma and prostate carcinoma) developed in r- and s-g in the form of 3D MCS (19, 20, 21, 22, 23, 24, 25, 26).

# B) Oncolytic Viral Therapy.

## HSV and Melanoma .-1

A new promising treatment for cancer is oncolytic viral therapy, in which a virus or a genetically modified virus is injected into tumor cells directly or via a systemic route (eg, intravenously) to induce direct lysis of tumor cells and/or systemic induction of an immune response against the tumor cells. Due to increased research on virus biology and tumor immunology, there is increased interest in oncolytic viruses. A clinical trial of advanced melanoma was treated with herpes simplex virus type 1 (HSV-1) encoding granulocyte-macrophage colony-stimulating factor (GM-CSF). This virus, called Ltalimogene-laherparepvec, is expected to be approved by the FDA in the near future. (28

## Parvo Virus and Glioblastoma multiforme .-2

Parvo H1 virus (12) has been used in experiments on glioblastoma multiforme due to its kinetics being able to cross the blood brain barrier.(29

Multiple factors are involved in clearing the viral particles in Glioblastoma multifore cells such as TRAF3, INF-related factor 3, INF7 and RIG-1. These factors activate JAK-STAT pathway which activate PKR pathway leading to termination of protein synthesis and cell death .(16) The other mechanism by which oncolytic viruses cause tumor cell death is induction of a systemic immune response , multiple cytokines such as type 1 INF , TNF alpha, INF gamma and IL-12 which play a role in stimulation of antigen presenting cells(APCs) such as dendiritic cells then activation of T-helper cells CD 4 and C-cytotoxic cells CD 8 occurs leading to tumor cell lysis.(30

Glioblastoma cells may exhibit counteracting evading mechanisms via surface receptors which inactivate the effector immune cells and secrete inhibitory cytokines such as

Med. Sci. Forum 2023, 3, x 5 of 7

> IL-10,TGF-B and IDO to recruit immune suppressive cells. However, the viruses can be genetically modified to escape this suppressive microenvironment.(31)

#### 3. -. Reovirus and Pancreatic cancer:

A reovirus being tested in a Phase II clinical study for pancreatic cancer is called Oncolytics Biotech Inc., Calgary, Alberta, Canada[32]. The research is still inconclusive. However, it has been demonstrated that administering reovirus intraperitoneally in the management of peritoneal metastases in hamsters with pancreatic ductal adenocarcinoma carcinomatosis was efficient and secure[33.

Overexpression of CD46, a viral entrance receptor also present in many cancer cells, is required for measles virus development. An altered measles virus previously demonstrated oncolytic activity in pancreatic tumor xenografts in mice, leading to tumor shrinkage and improved survival [35]. In a different study, the virus was changed to specifically target prostate stem cell antigen, a protein found in pancreatic cancer.

Overexpression of CD46, a viral entrance receptor also present in many cancer cells, is required for measles virus development. 34) In a prior work, mice with pancreatic tumor xenografts treated with a modified measles virus shown oncolytic activity, which resulted in tumor shrinkage and improved survival [35]. In a different investigation, the virus was changed to target the purine nucleoside phosphorylase drug and the protein prostate stem cell antigen, which is expressed in pancreatic cancer. The researchers came to the conclusion that immunocompromised mice treated with viral treatment showed anticancer activity[36]

# *C)* Limitations of our hypothesis:

Unfavorable results would be decreased efficacy due to the inhibitory effects of microgravity on the human immune system which is one of the mechanisms oncolytic viruses utilize to abort the growth of tumor cells. We suggest adding Immune stimulants such as GM-CSF to escape this inhibitory effect. (

**Conclusion:** 28

The treatment of glioblastoma remains a challenge. We are pleased to publish our hypothesis that could offer a novel therapy for this highly malignant CNS tumor. We aim to conduct an experiment to confirm this hypothesis and, if confirmed, to conduct clinical trials.

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References 44

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Med. Sci. Forum **2023**, 3, x 6 of 7

Ostrom QT, Gittleman H, Farah P, Ondracek A, Chen Y, Wolinsky Y, et al. CBTRUS statistical report: Primary brain and central nervous system tumors diagnosed in the United States in 2006-2010. Neuro Oncol [Internet]. 2013 [cited 2022 Mar 23];15 Suppl 2(suppl 2):ii1-56. Available from: https://academic.oup.com/neuro-oncology/article/15/suppl\_2/ii1/1042185?login=false

- D'Alessio A, Proietti G, Sica G, Scicchitano BM. Pathological and molecular features of glioblastoma and its peritumoral tissue. Cancers (Basel) [Internet]. 2019 [cited 2022 Mar 23];11(4):469. Available from: https://www.mdpi.com/2072-6694/11/4/469
- Wrensch M, Fisher JL, Schwartzbaum JA, Bondy M, Berger M, Aldape KD. The molecular epidemiology of gliomas in adults. Neurosurg Focus [Internet]. 2005 [cited 2022 Mar 23];19(5):E5. Available from: https://theins.org/focus/view/journals/neurosurg-focus/19/5/foc.2005.19.5.6.xml
- 4 Ryskalin L, Biagioni F, Morucci G, Busceti CL, Frati A, Puglisi-Allegra S, et al. Spreading of Alpha Synuclein from Glioblastoma Cells towards Astrocytes Correlates with Stem-like Properties. Cancers [Internet]. 2022 Mar 10 [cited 2022 Jul 22];14(6):1417. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8946011/
- Elshourbagy T, Brašić JR, Syed AB. Guidelines for the Diagnosis and Treatment of Parkinson's Disease. Biology and Life Sciences Forum [Internet]. 2021 [cited 2022 Jul 22];9(1):9. Available from: https://www.mdpi.com/2673-9976/9/1/9/htm
- Elshourbagy T, Syed AB, Amer MAM, Brasic JR. Precision medicine to identify optimal diagnostic and therapeutic interventions for Parkinson's Disease. Medical Science and Discovery. 2021 Sep 11;8(9):514–9. Available from: https://doi.org/10.36472/msd.v8i9.595
- Yong RL, Lonser RR. Surgery for glioblastoma multiforme: striking a balance. World Neurosurg [Internet]. 2011 [cited 2022 Mar 23];76(6):528–30. Available from: http://dx.doi.org/10.1016/j.wneu.2011.06.053
- 8 Barani IJ, Larson DA. Radiation Therapy of Glioblastoma. In: Cancer Treatment and Research. Cham: Springer International Publishing; 2015. p. 49–73
- 9 Fernandes C, Department of Medical Oncology, Centro Hospitalar de São João, Porto, Portugal, Costa A, Osório L, Lago RC, Linhares P, et al. Current standards of care in glioblastoma therapy. In: Glioblastoma. Codon Publications; 2017. p. 197–241
- 10 Harvard.edu. [cited 2022 Apr 9]. Available from: https://ui.adsabs.harvard.edu/abs/2012cosp...39..274C/abstract
- Topal U, Zamur C. Microgravity, stem cells, and cancer: A New Hope for cancer treatment. Stem Cells Int [Internet]. 2021 [cited 2022 Mar 23];2021:1–9. Available from: http://dx.doi.org/10.1155/2021/5566872
- 12 Takeda M, Magaki T, Okazaki T, Kawahara Y, Manabe T, Yuge L, et al. Effects of simulated microgravity on proliferation and chemosensitivity in malignant glioma cells. Neurosci Lett [Internet]. 2009;463(1):54–9. Available from: http://dx.doi.org/10.1016/j.neulet.2009.07.045 ,
- 13 Svejgaard, B.; Wehland, M.; Ma, X.; Kopp, S.; Sahana, J.; Warnke, E.; Aleshcheva, G.; Hemmersbach, R.; Hauslage, J.; Grosse, J.; et al. Common Effects on Cancer Cells Exerted by a Random Positioning Machine and a 2D Clinostat. PLoS ONE 2015, 10, e0135157.
- Warnke, E.; Pietsch, J.; Wehland, M.; Bauer, J.; Infanger, M.; Görög, M.; Hemmersbach, R.; Braun, M.; Ma, X.; Sahana, J.; et al. Spheroid formation of human thyroid cancer cells under simulated microgravity: A possible role of CTGF and CAV1. Cell Commun. Signal. 2014, 12, 32. [
- 15 Kopp, S.; Warnke, E.; Wehland, M.; Aleshcheva, G.; Magnusson, N.E.; Hemmersbach, R.; Corydon, T.J.; Bauer, J.; Infanger, M.; Grimm, D. Mechanisms of three-dimensional growth of thyroid cells during long-term simulated microgravity. Sci. Rep. 2015, 5, 16691
- 16 Grimm, D.; Bauer, J.; Kossmehl, P.; Shakibaei, M.; Schöberger, J.; Pickenhahn, H.; Schulze-Tanzil, G.; Vetter, R.; Eilles, C.; Paul, M.
- et al. Simulated microgravity alters differentiation and increases apoptosis in human follicular thyroid carcinoma cells. FASEB J. 2002, 16, 604–606. [
- 17 Kossmehl, P.; Shakibaei, M.; Cogoli, A.; Infanger, M.; Curcio, F.; Schönberger, J.; Eilles, C.; Bauer, J.; Pickenhahn, H.; Schulze-Tanzil.
- G.; et al. Weightlessness induced apoptosis in normal thyroid cells and papillary thyroid carcinoma cells via extrinsic and intrinsic pathways. Endocrinology 2003, 144, 4172–4179
- Infanger, M.; Kossmehl, P.; Shakibaei, M.; Schulze-Tanzil, G.; Cogoli, A.; Faramarzi, S.; Bauer, J.; Curcio, F.; Paul, M.; Grimm, D. Longterm conditions of mimicked weightlessness influences the cytoskeleton in thyroid cells. J. Gravit. Physiol. 2004, 11, P169–P172.
- 19 Grimm, D.; Infanger, M.; Westphal, K.; Ulbrich, C.; Pietsch, J.; Kossmehl, P.; Vadrucci, S.; Baatout, S.; Flick, B.; Paul, M.; et al. A delayed type of three-dimensional growth of human endothelial cells under simulated weightlessness. Tissue Eng. Part A 2009, 15, 2267–2275
- 20 Freed, L.E.; Langer, R.; Martin, I.; Pellis, N.R.; Vunjak-Novakovic, G. Tissue engineering of cartilage in space. Proc. Natl. Acad. Sci
- USA 1997, 94, 13885-13890
- 21 Morey-Holton, E.R.; Globus, R.K. Hindlimb unloading of growing rats: A model for predicting skeletal changes during space flight. Bone 1998, 22, 83s–88s.
- Borst, A.G.; van Loon, J.J.W.A. Technology and Developments for the Random Positioning Machine, RPM. Microgravity Sci. Technol. 2008, 21, 287
- Melnik, D.; Krüger, M.; Schulz, H.; Kopp, S.; Wehland, M.; Bauer, J.; Baselet, B.; Vermeesen, R.; Baatout, S.; Corydon, T.J.; et al. The CellBox-2 Mission to the International Space Station: Thyroid Cancer Cells in Space. Int. J. Mol. Sci. 2021, 22, 8777

Med. Sci. Forum **2023**, 3, × 7 of 7

Pietsch, J.; Ma, X.; Wehland, M.; Aleshcheva, G.; Schwarzwälder, A.; Segerer, J.; Birlem, M.; Horn, A.; Bauer, J.; Infanger, M.; et al. Spheroid formation of human thyroid cancer cells in an automated culturing system during the Shenzhou-8 Space mission. Biomaterials 2013, 34, 7694–7705.

- Ma, X.; Pietsch, J.; Wehland, M.; Schulz, H.; Saar, K.; Hübner, N.; Bauer, J.; Braun, M.; Schwarzwälder, A.; Segerer, J.; et al. Differential gene expression profile and altered cytokine secretion of thyroid cancer cells in space. FASEB J. 2014, 28, 813–835.
- Infanger, M.; Kossmehl, P.; Shakibaei, M.; Bauer, J.; Kossmehl-Zorn, S.; Cogoli, A.; Curcio, F.; Oksche, A.; Wehland, M.; Kreutz, R.;
- et al. Simulated weightlessness changes the cytoskeleton and extracellular matrix proteins in papillary thyroid carcinoma cells. Cell Tissue Res. 2006, 324, 267–277
- 27 Carmeliet G, Nys G, Bouillon R. Microgravity reduces the differentiation of human osteoblastic MG-63 cells. J Bone Miner Res [Internet]. 1997;12(5):786–94. Available from: http://dx.doi.org/10.1359/jbmr.1997.12.5.786
- Zawit M, Swami U, Awada H, Arnouk J, Milhem M, Zakharia Y. Current status of intralesional agents in treatment of malignant melanoma. Ann Transl Med [Internet]. 2021 [cited 2022 Mar 23];9(12):1038. Available from: http://dx.doi.org/10.21037/atm-21-491
- Martuza RL, Malick A, Markert JM, Ruffner KL, Coen DM. Experimental therapy of human glioma by means of a genetically engineered virus mutant. Science [Internet]. 1991;252(5007):854–6. Available from: http://dx.doi.org/10.1126/science.1851332
- 30 Kaufman HL, Kohlhapp FJ, Zloza A. Oncolytic viruses: a new class of immunotherapy drugs. Nat Rev Drug Discov [Internet]. 2015 [cited 2022 Mar 23];14(9):642–62. Available from: https://www.nature.com/articles/nrd4663
- Clemens MJ. Targets and mechanisms for the regulation of translation in malignant transformation. Oncogene [Internet]. 2004 [cited 2022 Mar 23];23(18):3180–8. Available from: https://www.nature.com/articles/1207544
- 32 Galanis E, Markovic SN, Suman VJ, Nuovo GJ, Vile RG, Kottke TJ, Nevala WK, Thompson MA, Lewis JE, Rumilla KM, Roulstone V, Harrington K, Linette GP, Maples WJ, Coffey M, Zwiebel J, Kendra K. Phase II trial of intravenous administration of Reolysin(®) (Reovirus Serotype-3-dearing Strain) in patients with metastatic melanoma. Mol Ther 2012; 20: 1998-2003 [PMID: 22871663 DOI: 1
- 33 Hirano S, Etoh T, Okunaga R, Shibata K, Ohta M, Nishizono A, Kitano S. Reovirus inhibits the peritoneal dissemination of pancreatic cancer cells in an immunocompetent animal model. Oncol Rep 2009; 21: 1381-1384 [PMID:
- 34 Galanis E. Therapeutic potential of oncolytic measles virus: promises and challenges. Clin Pharmacol Ther 2010; 88: 620-625 [PMID: 20881957 DOI: 10.1038/clpt.2010.211
- Penheiter AR, Wegman TR, Classic KL, Dingli D, Bender CE, Russell SJ, Carlson SK. Sodium iodide symporter (NIS)-mediated radiovirotherapy for pancreatic cancer. AJR Am J Roentgenol 2010] 349-341:195; PMID: 20651188 DOI: 10.2214/AJR.09.3672
- 36 Bossow S, Grossardt C, Temme A, Leber MF, Sawall S, Rieber EP, Cattaneo R, von Kalle C, Ungerechts G. Armed and targeted measles virus for chemovirotherapy of pancreatic cancer. Cancer Gene Ther 2011; 18: 598-608 [PMID: 21701532 DOI: 10.1038/cgt.2011.30]
- 37 Kärre K, Ljunggren HG, Piontek G, Kiessling R. Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategy. Nature [Internet]. 1986 [cited 2022 Mar 23];319(6055):675–8. Available from: https://www.nature.com/articles/319675a0
- Ruffell B, Affara NI, Coussens LM. Differential macrophage programming in the tumor microenvironment. Trends Immunol [Internet]. 2012 [cited 2022 Mar 23];33(3):119–26. Available from: https://www.cell.com/trends/immunology/fulltext/S1471-4906(11)00213-
  - 4?\_returnURL=https%3A%2F%2Flinkinghub.elsevier.com%2Fretrieve%2Fpii%2FS1471490611002134%3Fshowall%3Dtru
- Martinelli LK, Russomano T, Dos Santos MA, Falcao FP, Bauer ME, Machado A, et al. Effect of microgravity on immune cell viability and proliferation: simulation using 3-D clinostat. IEEE Eng Med Biol Mag [Internet]. 2009;28(4):85–90. Available from: https://ieeexplore.ieee.org/document/5165230
- 40 Agha NH, Mehta SK, Rooney BV, Laughlin MS, Markofski MM, Pierson DL, et al. Exercise as a countermeasure for latent viral reactivation during long duration space flight. FASEB J [Internet]. 2020;34(2):2869–81. Available from: http://dx.doi.org/10.1096/fj.201902327R
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