

Profiling of Antibacterial Compounds from Selective Medicinal Mangrove Species

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Abstract: Mangrove is an opulent and untapped ecosystem with great phytochemical diversity, making it suitable for the discovery of novel antimicrobial compounds. The goal of the study was to explore the pharmaceutical antibacterial and antioxidant resources from *Bruguiera gymnorrhiza*, *Ceriops tagal*, *Rhizophora mucronata*, and *Aegiceras corniculatum* and gain insight into the diversity and novelty of compounds like alkaloids, flavanol, polyphenols, etc. A liquid extract was obtained by subjecting fresh mangrove leaves to Maceration & Soxhlet extraction. Plant DNA barcoding was utilized to authenticate the identity of the samples under study. A few of the obtained sequences have been communicated to GenBank (under review; accession number awaited). The phytochemical profiling revealed the presence of polyphenols (TPC = 8.71, 8.51, 8.77, 5.52 mg/gm of plant tissue resp.); flavonoids (TFC = 12.42, 8.48, 5.26, 13.903 mg/gm of plant tissue resp.); and alkaloids (2.5, 3.81, 4.98, 5.21 mg/gm of plant tissue resp.). The antioxidant potential (radical scavenging activity) was scored to be 87.8%, 89.5%, 92.07%, and 45.8% (DPPH assay was conducted). The antimicrobial analysis was performed on *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Bacillus subtilis*. The MIC revealed maximum activity in *Klebsiella pneumoniae*, while negligible activity was scored for *Staphylococcus aureus* and *Escherichia coli*. The analysis thus reveals that the plants under study may have better medicinal activity against respiratory tract organisms. In-vitro Biochemical analysis of different molecules present in the plants was done using the Swiss ADME database. The present study thus reveals the preliminary compounds from selected mangrove plants that can be promising future anti-microbial therapeutics.

Keywords:

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1. Introduction

A major peril among infectious diseases, which account for millions of deaths across the globe per year is a bacterial infection (1). The use of antibiotics or chemicals is highly recommended as a therapy. However there are instances of development of multiple drug resistance strains and incomplete bioactivity of these chemical drugs. Furthermore these chemical drugs have been reported to have profound impact on liver, kidney and several other vital organs in the human body which can have a deleterious cytopathic effect (2). Therefore, the search for novel antimicrobials for emerging and reemerging bacterial diseases is urgent. Mangroves coastline forests are well known for their ecological significance. Many bioactive substances from different mangrove species, including steroids, triterpenes, saponins, flavonoids, alkaloids, and tannins have been reported to have therapeutic potentials (3). Numerous studies have demonstrated the effectiveness of these plants against pathogens that affect people, animals, and plants. In comparison to sea-

weeds and sea grasses, it has also been claimed that these are an excellent source of anti-viral compounds (4). They were described as having incredible potential against cancer cells by Boopathy and Kathiresan (5).

The present research study aims to screen the bioactive components from *Ceriops Tagal*, *Rhizophora Mucronata*, *Brugeria gymnoriza*, and *Agicerous corniculatom* against the bacteria causing respiratory, soft tissue and gastrointestinal infections. *Ceriops tagal* is a small tree with short buttresses and knee-like breathing roots. the bark of *C. tagal* has been used for the treatment of infected wounds in Thailand, and obstetric and hemorrhagic conditions in the Philippines. The species is also used to treat sores, hemorrhages and malignant ulcers, and malaria in Asian countries. In the Philippines, the reddish-brown ground bark of *C. tagal* is added to toddy (an alcoholic drink from the inflorescence sap of coconut) as a preservative to delay the fermentation process by controlling spoilage microbes (6). *Rhizophora mucronata* is found in the Indo-Pacific region on the banks of rivers and on the edge of the sea. It has long been traditionally used for the treatment of elephantiasis, hematoma, hepatitis, ulcers and febrifuge. Its leaf is being used in the folk medicine for treating diarrhea or gastric motility disorder (7). *Bruguiera gymnorrhiza* (L.) (Rhizophoraceae) is an evergreen mangrove tree, widely distributed in tropical and subtropical coastlines. This plant has exhibited anti-tumors activity against HepG2 hepatoma cells, antibacterial activity against selected microbial species, and germicidal activity. *Aegicerous corniculatum* is a mangrove plant grows in the wetland of tropical and subtropical regions of Indus Delta valley of Pakistan, Western costline of India. It has a potential effect in many diseases like diabetes, inflammation, rheumatism, cardio vascular diseases etc. *A. corniculatum* has been used as folklore medicine since long time but there is not much scientific evidence available to justify its medicinal use

The present research study highlights the screening and quantification of bioactive compound of the four plants understudy applying in-silico tools basal ADME has also been gauged. the anti-bacterial potentials have been screened against *Klebsiella Pneumonia*, *E.coli*, *S.aureus* & *Bacillus subtilis*.

2. Material and Methods

Four different species of mangroves were collected from a mangrove nursery established at shores of Gorai, Mumbai, India. Collected Plant samples were maintained in the greenhouse of SBDYPUNM, India, under its favorable conditions until further studies

Extraction of phytochemicals-

Soxhlet Method

Plant leaves were sorted (less bruised) and washed under running tap water. Leaves were then ground to a fine powder using a mortar and pestle with the help of liquid nitrogen. Then the extract was packed in a thimble and was extracted using different solvents such as petroleum ether, Chloroform, methanol, etc. for 3hrs. Resulting extracts in different solvents were evaporated and concentrated to dryness using the rotary evaporator at 50°C. Powder was dissolved in the solvents used for extraction (8).

Qualitative and Quantitive estimation of bioactive compounds

A preliminary qualitative screening of bioactive compounds from plants under study was followed by their quantitative estimation. The procedures described by Syahidah and N Subekti (2019) (9) were adopted for the study with some modifications.

Alkaloid determination

5g of each dried ground sample was placed in a 250 ml beaker. 200ml of 10% acetic acid in ethanol was added, covered, and allowed to stand for 4 hours. This was filtered

and the extract was concentrated in a water bath to 1/4 of the original volume. Concentrated ammonium hydroxide was added dropwise to the content until precipitation was complete. The resulting solution was allowed to settle and the precipitate was collected and washed with dilute ammonium hydroxide and then filtered. The residue was the alkaloid which was dried and weighed.

Tannin determination

500mg of each sample was placed in a 50-ml plastic bottle. 50ml of distilled water was added and shaken for 1h in a mechanical shaker. This was filtered into a 50ml volumetric flask and made up to the mark. Then 5 ml of the filtrate was pipetted into a test tube and mixed with 2 ml of 0.1M ferric chloride in 0.1N hydrochloric acid and 0.008M potassium ferrocyanide. The absorbance was measured at 280nm within 10 mins. Tannin contents were expressed as a percentage of the dried fraction.

Flavonoid determination

1g of each ground sample was extracted repeatedly with 100 ml of 80% aqueous methanol at room temperature. The whole solution was filtered through Whatman filter paper No. 42 (125mm). The filtrate was later transferred into a crucible and evaporated to dryness (constant weight) over a water bath. The flavonoid content was calculated as a percentage of the dried fraction.

Reducing sugars determination

1g of each ground sample was diluted with water (10 ml) and titrated with a standard Benedict reagent. The sample was hydrolyzed with standard acid (0.5N HCl). The hydrolyzed fraction gave the total reducing sugars. The results obtained were calibrated using the standard curve of glucose.

Estimation of total phenolic content

The Folin-Ciocalteu method will be used to estimate the total phenolic content. 1ml of extract solution of concentration from 100-500µg/ml will be added to 2.5ml of 10%(w/v) Folin-Ciocalteu reagent. 2ml of 75% Na₂CO₃ will be mixed into the above solution after 5 minutes and will be incubated at 50°C for 10 minutes. Then the sample will be cooled, and absorbance will be measured at 765nm by a UV spectrophotometer against the blank solution. The data will be expressed as mg/g of gallic acid equivalents in milligrams per gram of dry extract.

Estimation of flavonoids

The mixture of 10ml solution will be made by mixing 1ml of plant extract, 3ml of 70% ethanol, 0.2ml of 10% aluminum chloride, 0.2ml of potassium acetate (1M), and 5.6ml of distilled water. The solution will be incubated for 30mins at room temperature, and then a UV spectrophotometer at 415nm will be used to measure the absorbance of the solution against the blank solution. 5mg of Quercetin will be mixed with 1 ml of methanol to prepare the stock solution, then different concentrations (5-200µg/ml) of standard quercetin solution will be prepared. The concentration of total flavonoid content in the test samples will be calculated from the calibration plot ($Y = 0.0162x + 0.0044$, $R^2 = 0.999$) and expressed as mg quercetin equivalent (QE)/g of dried plant material. All the determinations were carried out in triplicate.

Anti-oxidant Activity (DPPH) Free Radical Scavenging assay

1 ml of 0.135 mM DPPH in methanol solution will be added to tubes containing 1 ml of plant extracts, vitamin C, and gallic acid at varying doses (0.2–1.0 mg/ml). The mixture will be shaken and then kept at room temperature for 30 minutes in the dark. After that,

the mixture's absorbance will be measured spectrophotometrically at 517 nm. As benchmarks, vitamin C and gallic acid will be utilized.

The DPPH radical scavenging activity will be calculated by:

$$= \frac{\text{Abs control} - \text{Abs sample}}{\text{Abs control}} \times 100\%$$

Where Abs control is the absorbance of DPPH and methanol; Abs sample is the absorbance of DPPH radical + sample extract or standards (Vitamin C and gallic acid).

Anti-bacterial activity-

A bacterial culture was prepared by inoculating a fresh colony from an overnight culture into a sterile broth medium and incubating it for 4-6 hours at the appropriate temperature with agitation until the bacterial density reaches 0.5 McFarland standard ($1-2 \times 10^8$ CFU/mL). *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Bacillus subtilis* were the strains used in this assay. Serial dilutions were prepared for the test sample in a suitable broth medium (e.g. Mueller-Hinton broth) in a 96-well plate, starting from the highest concentration. Add the bacterial inoculum to each well, except for the negative control wells (only broth medium). Positive control wells with known antibacterial agents to ensure the validity of the assay. Cover the plate and incubate it at the appropriate temperature (e.g. 37°C) for 18-24 hours without agitation. After incubation, The MIC (minimum inhibitory concentration) is defined as the lowest concentration of the test sample that completely inhibits the visible growth of bacteria. MIC was determined by visually inspecting the wells or by using a microplate reader to measure the optical density (OD) at 600 nm (8).

In-silico study

Here we have derived the small chemical structures, from the literature sources and Pubchem chemical compound database (10). The compounds were initially screened for physiochemical property, drug likeness, ADME properties, PAINS, Brenk alerts, Synthetic accessibility using SWISS-ADME online server (11).

3. Results

Preliminary phytochemical screening revealed the presence of alkaloid, flavonoids, phenolic, and tannin in both of the acetone and methanol extracts. On the contrary, terpenoid, steroid and saponin were absent (Figure 2). Total phenolic content of four mangrove samples in their crude and 1:10 diluted forms was calculated and found to be 00.07274, 0.87119, 0.83833, 0.851305, 0.8625, 0.8625 0.87785, 0.536425, and 0.5525 GAE/wet weight (Gallic acid equivalent) for crude (A), 1:10 (A), 1:10(A), Crude (B), 1:10 (B), Crude (C), 1:10 (C), Crude(D), 1:10(D). The total flavonoids content of four mangrove samples in their crude and 1:10 diluted forms was calculated and found to be -0.57115, 1.24231, 0.53846, 0.848075, 1.33846, -0.52692, 2.10565, and 13.9038 GAE/wet weight (gallic acid equivalent) for crude (A), 1:10 (A), crude (B), 1:10 (B), crude (C), 1:10 (C), and crude (110), respectively. The study revealed that flavonoids, alkaloids and phenol were found to be present the extract. Several studies reported that, flavonoids show anti-allergic, anti-inflammatory, anti-microbial and anti-cancer activity.

The antioxidant potential (radical scavenging activity) was scored to be 87.8%, 89.5%, 92.07%, and 45.8% (DPPH assay was conducted)(figure 2) Antibacterial assays for plant extracts of mangrove plants and the controls were also carried out. The extracts exhibited different percentage of inhibitions for different microbial strains under study (figure 3,4,5). The antimicrobial analysis was performed on *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Bacillus subtilis*. The MIC revealed maximum activity in *Klebsiella pneumonia*, while negligible activity was scored for *Staphylococcus aureus* and *Escherichia coli*.

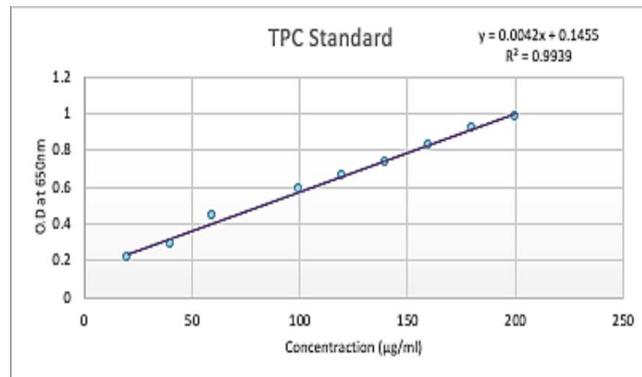


Figure 1. Graphical Representation of total phenolic content standards and samples.

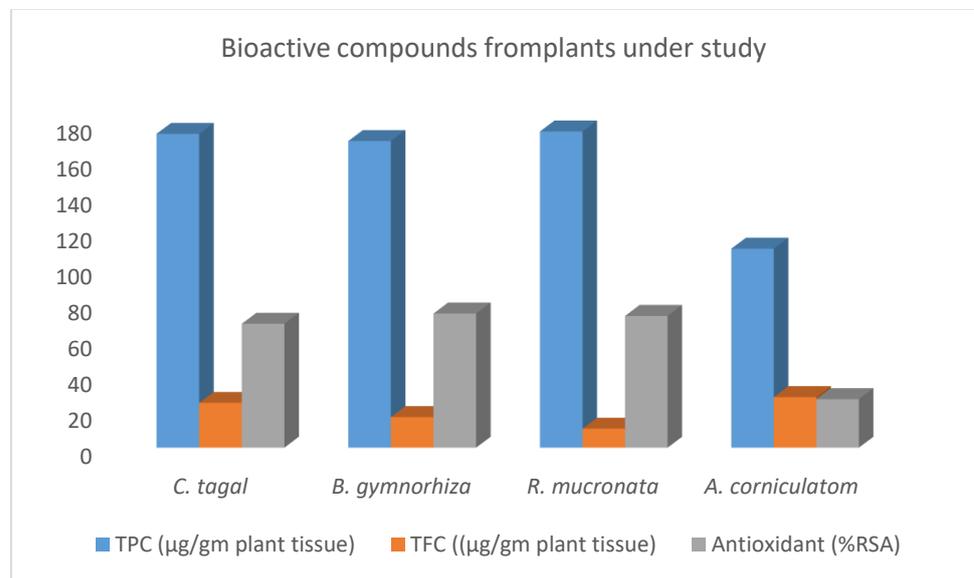


Figure 2. Graphical representation of quantified bioactive components for plants under study.

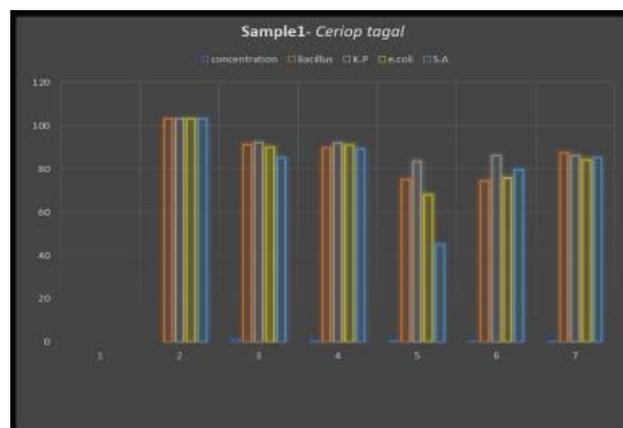


Figure 3. Percentage inhibition of bactreial cells in presence of *C. tagal* extracts.

Mole cule 4	C9H8 O3	High	Yes	No	No	No	No	No	No	0	0.85	0	1	1.61
Mole cule 5	C9H1 6O4	High	Yes	No	No	No	No	No	No	0	0.85	0	0	1.57
Mole cule 10	C7H6 O2	High	Yes	No	No	No	No	No	No	0	0.55	0	1	1
Mole cule 17	C12H 14O4	High	Yes	No	No	No	No	No	No	0	0.85	0	0	1.99
Mole cule 3	C15H 24O	High	Yes	No	No	Yes	Yes	No	No	0	0.55	0	2	4.35
Mole cule 20	C16H 33NO	High	Yes	No	Yes	No	No	No	No	0	0.55	0	0	2.12
Mole cule 1	C18H 32O2	High	Yes	No	Yes	No	Yes	No	No	1	0.85	0	1	3.1
Mole cule 21	C18H 35NO	High	Yes	No	Yes	No	Yes	No	No	1	0.55	0	1	2.97
Mole cule 22	C16H 32O2	High	Yes	No	Yes	No	Yes	No	No	1	0.85	0	0	2.31
Mole cule 18	C18H 30O3	High	Yes	No	Yes	No	Yes	Yes	No	0	0.85	0	2	3.44
Mole cule 16	C18H 28O3	High	Yes	No	Yes	Yes	Yes	Yes	No	0	0.85	0	1	4.2
Mole cule 8	C9H1 0O5	High	No	No	No	No	No	No	No	0	0.56	0	0	1.7
Mole cule 9	C8H1 5NO	High	No	No	No	No	No	No	No	0	0.55	0	0	3.05
Mole cule 12	C24H 38O4	High	No	No	No	No	No	No	Yes	1	0.55	0	1	3.41

Mole cule 11	C9H6 O4	High	No	No	Yes	No	No	No	No	0	0.55	1	2	2.61
Mole cule 24	C18H 36O2	High	No	No	Yes	No	No	No	No	1	0.85	0	0	2.54
Mole cule 2	C15H 10O5	High	No	No	Yes	No	No	Yes	Yes	0	0.55	0	0	2.96
Mole cule 6	C15H 10O6	High	No	No	Yes	No	No	Yes	Yes	0	0.55	0	0	3.14
Mole cule 15	C15H 10O6	High	No	No	Yes	No	No	Yes	Yes	0	0.55	1	1	3.02
Mole cule 23	C18H 34O2	High	No	No	Yes	No	Yes	No	No	1	0.85	0	1	3.07
Mole cule 7	C30H 48O3	Low	No	No	No	No	No	No	No	1	0.85	0	1	6.21
Mole cule 13	C21H 20O11	Low	No	No	No	No	No	No	No	2	0.17	1	1	5.04
Mole cule 14	C21H 20O11	Low	No	No	No	No	No	No	No	2	0.17	1	1	5.17
Mole cule 19	C30H 50O2	Low	No	No	No	No	No	No	No	1	0.55	0	1	5.68
Mole cule 25	C22H 45NO	Low	No	No	Yes	No	No	No	No	1	0.55	0	0	2.82

3.1. Extractive Value of Bark and Root Extract of *Myrica esculenta* Plant

4. Discussion

Mangroves are one of the most prolific and unmapped ecosystem that roughly covers one fourth of the world coastline with high assortment of thriving organisms. Mangroves support the conservation of biological diversity for a number of endangered species by providing habitats, nurseries, nutrients, and spawning grounds (12). Mangroves play

also a key role in human sustainability and livelihoods, being heavily used for food, timber, fuel and medicine. They offer protection from calamitous events, such as tsunamis, tropical cyclones and tidal bores and can dampen shoreline erosion (13).

Flavonoids are now considered as an indispensable component in a variety of nutraceutical, pharmaceutical, medicinal and cosmetic applications. This is attributed to their anti-oxidative, anti-inflammatory, anti-mutagenic and anti-carcinogenic properties coupled with their capacity to modulate key cellular enzyme function (9). Alkaloids have been reported as powerful poison and many alkaloids derived from medicinal plants show biological activities like, anti-inflammatory, antimalarial, antimicrobial, cytotoxicity, antispasmodic and pharmacological effects. Similarly, steroids derived from plants are known to have cardiogenic effect and also possess antibacterial and insecticidal properties. They are very often used in medicines due to their well-known biological activities. Tannins, according to research, are known to have antibacterial antitumor and antiviral activities. These alkaloids show bioactivity against Gram-positive bacteria and cytotoxicity against leukemia and HeLa cell lines. Alkaloids, flavonoids and xanthenes that are potent inhibitors of various oxidative processes in both in vitro and in vivo system. These phytochemical compounds identified in the extracts may be responsible for the biological activities of the mangrove leaf extract (14).

Tannin and phenol are necessary for the repair and maintenance. Polyphenolic compounds have an aromatic benzene ring with substituted hydroxyl groups, including their functional derivatives. These are able to absorb free radicals and can chelate metal ions that could catalyze formation of ROS which promotes lipid peroxidation. Among polyphenols, flavonoids are of great importance because they help human body to fight against diseases. The ability of flavonoids to act as potent antioxidants depends on their molecular structures, the position of the hydroxyl group and other features in its chemical structure. They are abundantly found in plants as their glycoside. The most abundant flavonol which has a good antioxidant property is quercetin, as it has all the right structural features for free radical scavenging activity (15).

The findings of our study revealed that the methanol extracts of the mangroves exhibited substantial antibacterial effects against the bacterial strains tested. Notably, the maximum inhibition was observed against *Klebsiella pneumoniae*, indicating potential effects against bacterial infections of the respiratory and urinary systems. Since these plants are halophytes, which thrive under high salt stress conditions, our results suggest that these mangroves may produce certain bioactive compounds that hold promise for drug development to treat both acute and chronic diseases.

From the in-silico analysis we can identify that molecule 4, 5, 10 and 17 have exhibited acceptable properties of drug likeness and within the acceptable toxicity range. Furthermore, further investigations are warranted to evaluate the antimycobacterial, antiviral, and antiparasitic activities of these plant extracts. Additionally, other parts of these mangrove species should be studied to assess their potential as sources of novel antimicrobial agents.

5. Conclusions

The research found that the four plant species contain high amounts of organic compounds that have potential medicinal properties, including alkaloids, flavonoids, polyphenols, tannins, and total proteins. DNA barcoding was used to identify the four plant species and may have submitted their results to NCBI for validation or comparison with other DNA sequences in the public database. Antibacterial activity of the four plant species was tested against three bacterial strains and found that one plant species showed substantial microbial inhibition against *Klebsiella*, while another plant species showed moderate microbial inhibition against *E. coli* and *Bacillus subtilis*. The research suggests that the four plant species may have potential medicinal benefits for respiratory tract infections, based on their observed antibacterial activity against *Klebsiella*, a respiratory

pathogen. However, further research is needed to confirm this hypothesis and investigate the safety and mechanism of action of these plant compounds.

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