

Exploring the inter and intra-specific variability of *Androctonus* scorpion venoms.

Ines HILAL^{*1,2}, Soukaina KHOURCHA^{1,2}, Amal SAFI², Abdelaziz HMYENE², Reto STÖCKLIN³ and Naoual OUKACHE¹

¹ Laboratory of Venoms and Toxins, Pasteur Institute of Morocco, Casablanca, Morocco

² Laboratory of Biochemistry, Environment and Food Technology, Faculty of Sciences and Techniques of Mohammedia, Morocco.

³ Atheris Laboratories, Case Postale 314, CH-1233 Bernex, Geneva, Switzerland.

* Correspondence: drineshilal@gmail.com.

Abstract: Scorpion venom possesses a lethal sting and potential medicinal properties, making it a captivating natural elixir. Our study aimed to unravel the composition of *Androctonus* scorpion venoms in Morocco. Using electrospray mass spectrometry and high-performance liquid chromatography (HPLC), we conducted a thorough analysis to gain detailed insights into venom composition. The data unveiled a wide range of molecular weights (236-622 Da), influenced by factors such as species, genus, location, age, sex, and diet. Short toxins (2000-4000 Da) predominated in the venoms, effectively blocking K⁺ channels, while larger molecular weights (>4000 Da) corresponded to long toxins that modulate Na⁺ channels. Furthermore, we made intriguing discoveries of previously unidentified peptides (<2000 Da). This study provides valuable knowledge, shedding light on the intricate composition of scorpion venoms.

Keywords: scorpion; *Androctonus*; venom; peptides; LC-MS; Peptidic maps.

1. Introduction

Scorpions are a fascinating group of arachnids. These venomous creatures pose a significant public health risk, accounting for 30-50% of poisoning cases reported in Morocco, with an alarming number of 30,000 people falling victim to scorpion stings annually, predominantly children under 15 years old [1]. Morocco boasts the highest scorpion diversity in North Africa, with 61 species, including *Androctonus* scorpions which are commonly associated with envenomation cases [2, 3]. Scorpion venom is a complex mixture of bioactive molecules, particularly neurotoxins that target ion channels. Venom composition varies greatly between species and individuals, influenced by factors such as sex, age, diet, and environmental conditions [4]. Proteomic analysis, specifically mass spectrometry, has revolutionized the study of scorpion venom, enabling the identification of toxins and peptides, aiding in the development of therapeutic agents and antivenoms [5]. Our research focuses on unravelling the mysteries of *Androctonus* scorpion venoms, utilizing cutting-edge proteomic strategies to understand their composition, variability, and potential applications. Through advanced mass spectrometry techniques, we aim to shed light on these complex venoms, paving the way for scientific breakthroughs and innovative analysis approaches.

2. Materials and Methods

2.1. Venoms

Scorpions from high-risk areas, known for severe envenomation cases, were studied. Venom milking involved electrical stimulation, scorpions received weak 12 V pulses on their post abdomen to extract venom. The collected venom was centrifuged at 10,000 rpm for 10 min, freeze-dried, and stored at -80°C until needed [6].

Citation: To be added by editorial staff during production.

Academic Editor: Firstname Last-name

Published: date



Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

2.2. Venom analysis by mass spectrometry coupled to reverse phase high performance liquid chromatography (RP-HPLC)

The analysis of Moroccan scorpion venoms was performed using a Micromass Quattro Micro triple quadrupole ESI-MS coupled with RP-HPLC [7]. Each venom sample (50 µg of protein) was loaded onto a C18 Zorbax analytical column (150 mm length, 2.1 mm internal diameter, 3 µm particle size). Venom fractionation took place over 100 min with a mobile phase of 0.1% FA (solvent A) and ACN in 0.1% FA (solvent B). Elution used a linear gradient from 0% to 100% of solution B at a constant flow rate of 0.2 mL/min. The separated peaks were directly analyzed by the Micromass Quattro Micro ESI-MS triple quadrupole mass spectrometer. The MassLynx version 4 software was used to convert the generated MS peaks into molecular masses, analyzing the obtained spectra.

3. Results

3.1. Fractionation of venoms by reverse phase high performance liquid chromatography (RP-HPLC)

The venom of *A. mauritanicus* from Tadla exhibited 24 peaks, while *A. mauritanicus* from Oualidia and *A. bicolor* from Draa Valley displayed 37 peaks. The majority of these peaks eluted within a retention time (RT) range of 17 minutes to 74.45 minutes. However, a few minority peaks were eluted at an RT of less than 5 minutes in the venom of *A. maroccanus* from Marrakech, *A. barbouri* from Agadir, *A. amoreuxi* from Tata, and in the venom of all three *A. mauritanicus* specimens. Intraspecific variability is evident in the venoms of scorpions within the same species. This variability is exemplified by the venom profiles of *A. mauritanicus* specimens from Oualidia and Essaouira, which are complex and nearly identical. In contrast, the venom of the Tadla specimen differs and is less complex. This intraspecific variability is likely influenced by their geographical locations.

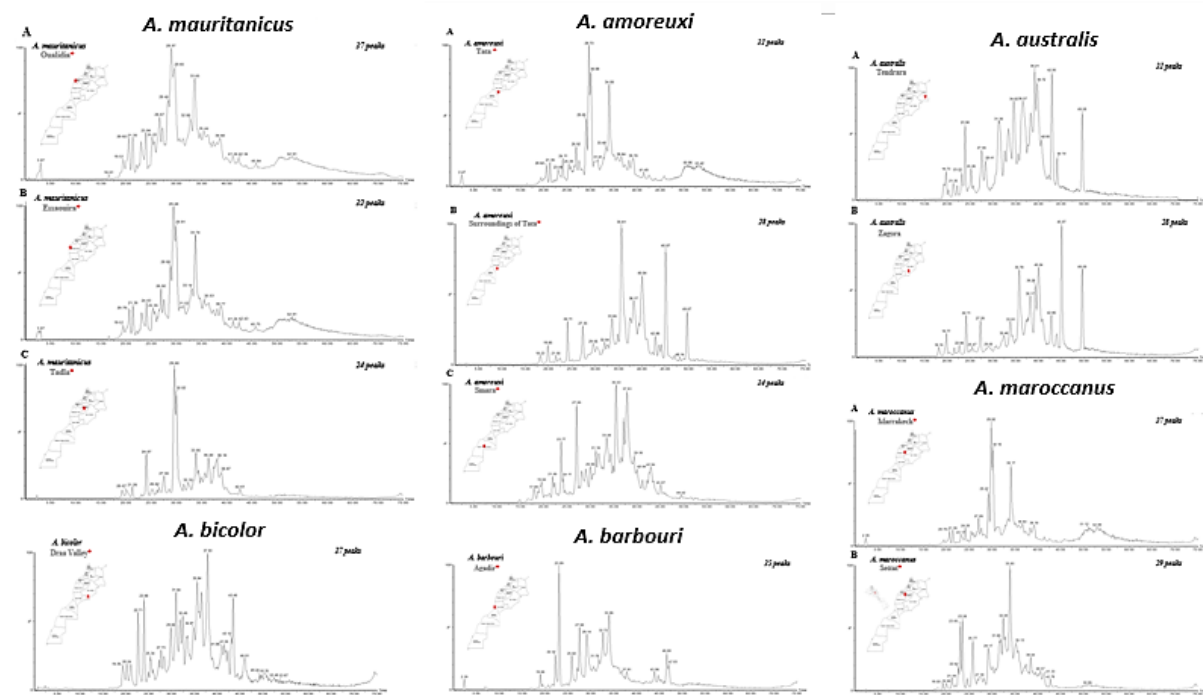


Figure 1. RP-HPLC profiles of venom from *Androctonus* specimens collected from different regions in Morocco.

Additionally, intraspecific variability is observed in the venom of *A. amoreuxi*, where the three venom profiles demonstrate differences in appearance and complexity.

This difference persists even among scorpions collected from two adjacent regions, Tata and its surroundings.

Similarly, intraspecific variability is noted in the venom of the two specimens of *A. australis* and the two specimens of *A. maroccanus*.

3.2. Analysis by mass spectrometry

The RP-HPLC-separated peaks were subjected to analysis using triple quadrupole ESI-MS mass spectrometry. The resulting data were then processed using MassLynx 4 software to identify the molecular masses associated with each peak. The table 1 presents the number of molecular masses identified in the venoms of *Androctonus* scorpions.

Table 1. Molecular masses generated after the analysis of the different peaks by the MassLynx4 software.

Genus	Species	Number	Region	Molecular masses
Androctonus	<i>Androctonus mauritanicus</i>	3	Oualidia	469
			Essaouira	410
			Tadla	328
	<i>Androctonus amoreuxi</i>	3	Tata	374
			Tata surroundings	452
	<i>Androctonus australis</i>	2	Smara	309
			Zagora	336
	<i>Androctonus bicolor</i>	1	Draa Valley	578
	<i>Androctonus maroccanus</i>	2	Marrakech	312
			Settat	338
<i>Androctonus barbouri</i>	1	Agadir	236	

The total number of molecular masses observed ranges from 236 to 578. *A. bicolor* venom exhibits the highest number of different masses (578), followed by *A. mauritanicus* from Oualidia with 469 masses. The least complex venoms are found in *A. australis* from Zagora (336 different masses) and *A. barbouri* from Agadir with 236 molecular masses. The mass spectrometry results validate the RP-HPLC fractionation data and demonstrate both inter and intraspecific variability in the venom of Moroccan scorpions. Regarding the distribution of molecular weights in the venoms, masses between 2001 and 5000 Da (corresponding to neurotoxins targeting K⁺, Cl⁻, and Ca²⁺ channels) are the most abundant across all species analyzed. The venom of *A. maroccanus* from Settat exhibits the highest percentage at 58.28%. On the other hand, masses between 5001 and 10,000 Da (corresponding to neurotoxins targeting Na⁺ channels) are more prevalent in the venom of the three specimens of *A. mauritanicus*, with the highest percentage in the Essaouira specimen (36.42%). Similarly, *A. amoreuxi* shows a significant percentage in the venom of all three specimens, with the highest in the Tata surroundings specimen (34.71%). In *A. australis*, the Tendrara specimen has a percentage of 25.86%, while *A. bicolor* from Draa Valley exhibits 21.33%. However, these molecular masses are less abundant in the venom of *A. barbouri* from Agadir (3.39%).

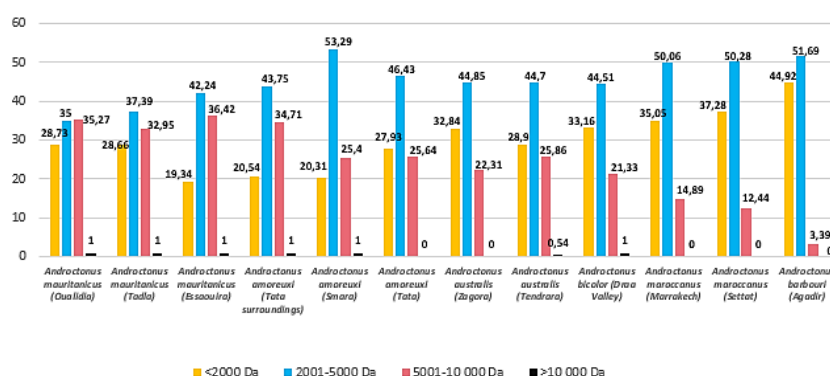


Figure 2. Molecular weight distribution of venoms from *Androctonus* species.

4. Discussion

The aim of this study was to characterize the venoms of scorpions belonging to the *Androctonus* genus collected from various regions in Morocco. The venoms were analyzed using MS mass spectrometry coupled with RP-HPLC, revealing differences in chromatographic profiles, including peak number, intensity, and retention time, both within and between species. For example, in *A. mauritanicus*, the venom profiles of specimens from Oualidia and Essaouira were highly similar and complex, while the venom from the Tadla specimen was less complex and distinct. The RP-HPLC profiles obtained provide a partial representation of the venom composition for each species, with characteristic peaks that can aid in taxonomic identification and differentiation.

Analysis of the peaks using MassLynx4 software showed that species from the *Androctonus* genus exhibited a high number of molecular masses, ranging from 236 (*A. barboursi*) to 578 (*A. bicolor*). A similar number of molecular masses were detected in the venoms of *Tityus metuendus* and *Rhopalurus junceus* species (200 masses); *Serradigitus gertschi* (204 masses); *Tityus discrepans* (205 masses); *Paravaejovis schwenkmeyeri* (212 masses); *Leiurus quinquestriatus quinquestriatus* (380 masses); *Tityus serrulatus* (382 masses); *Pandinus cavimanus* (390 masses); *Centruroides limpidus* (395 masses); *Tityus bahiensis* (464 masses); *Leiurus quinquestriatus hebraeus* (554 masses) and *Tityus stigmurus* (632 masses) [8,9,10,11,12,13,14,15], while a low number of mass was identified in the venoms of some scorpions, namely *Leiurus abdullahbayrami* (45 masses); *Buthacus macrocentrus* (60 masses); *Scorpio maurus palmatus* (73 masses); *Androctonus mauretanicus mauretanicus* (74 masses), *Androctonus crassicauda* (80 masses); *Tityus stigmurus* (100 masses) and *Opisthacanthus elatus* (106 masses) [16,17,18,19,20,21].

The molecular weight distribution of *Androctonus* venoms revealed a predominance of molecular masses between 2001 and 5000 Da, corresponding to neurotoxins that target K^+ , Cl^- , and Ca^{2+} channels. A high percentage of these masses was found in the venom of *A. maroccanus*, followed by masses between 5001 and 10,000 Da, representing neurotoxins that act on Na^+ channels, with a significant presence in the venom of *A. mauritanicus* from Essaouira. These findings align with previous studies demonstrating the prevalence of molecular masses between 2001 and 5000 Da in the venom of *Scorpio maurus palmatus* and *Buthus occitanus* [22, 23].

Neurotoxins targeting Na^+ ion channels, which are responsible for envenomation symptoms, were prominently represented in the venom of *Androctonus* scorpions. In particular, the three specimens of *A. mauritanicus* exhibited percentages of 32.95% (Tadla), 35.27% (Oualidia), and 36.42% (Essaouira), while the three specimens of *A. amoreuxi* showed percentages of 25.4% (Tata specimen), 25.4% (surroundings of Tata), and 34.71% (Smara). The two specimens of *A. australis* displayed percentages ranging from 22.31% (Zagora) to 25.86% (Tendrarra), and *A. bicolor* from Agadir had a percentage of 21.33%.

These results support the literature's depiction of the *Androctonus* genus as the most dangerous worldwide, particularly in North Africa, the Middle East, and Asia. The findings also provide insights into the areas at high risk of envenomation, mainly concentrated in the center of the kingdom. This correlation aligns with the distribution of severe envenomation cases according to the CAPM (Centers for Antipoison and Pharmacovigilance of Morocco) [1].

Thus, the mass peptide maps generated from the analysis of different venoms can be similar to an overview of the genome of the species studied, allowing the genetic matching of specimens and then, the use of venom profiles for taxonomic purposes must be demonstrated.

5. Conclusion

This study significantly enhances our understanding of *Androctonus* scorpion venom. By employing proteomic techniques, we successfully characterized the venom proteome of scorpions from the *Androctonus* genus across different regions of Morocco. The findings highlight the remarkable variability of scorpion venoms, influenced by factors such as scorpion biology and ecological conditions. Additionally, the study identifies the most potent venom sources and high-risk regions for envenomation. This knowledge is invaluable for developing effective antivenoms and potential drugs.

Author Contributions: Conceptualization, I.H. and N.O.; methodology, I.H. and S.K.; software, I.H. and S.K.; validation, R.S. and N.O.; formal analysis, I.H.; investigation, I.H. and N.O.; resources, R.S.; data curation, I.H. and N.O.; writing—original draft preparation, I.H.; writing—review and editing, A.S. and A.H.; visualization, R.S. and N.O.; supervision, A.S. and N.O.; project administration, N.O.; All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Acknowledgments: We thank Atheris Laboratories for the technical support.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. El Oufir, R. Piqures et Envenimations Scorpioniques (PES); Rapports general et specifiques. Revue toxicologie Maroc; Centre Anti Poison du Maroc (CAPM): Rabat, Morocco, 2019; Volume 43, p. 11.
2. Touloun O. Liste Actualisee et commentee de la Faune scorpionique du Maroc (Arachnida: Scorpiones) Rev Iberica Aracnol. 30 juin 2019;34:126-32.
3. Hilal I, Khourcha S, Safi A, Hmyene A, Asnawi S, Othman I, et al. Comparative Proteomic Analysis of the Venoms from the Most Dangerous Scorpions in Morocco: *Androctonus mauritanicus* and *Buthus occitanus*. Life. mai 2023;13(5):1133.
4. Cid-Urbe JI, Veytia-Bucheli JI, Romero-Gutierrez T, Ortiz E, Possani LD. Scorpion venomomics: a 2019 overview. Expert Rev Proteomics. 2 janv 2020;17(1):67-83.
5. Escoubas P, Quinton L, Nicholson GM. Venomomics: unravelling the complexity of animal venoms with mass spectrometry. J Mass Spectrom. 2008;43(3):279-95.
6. Oukkache N, Chgoury F, Lalaoui M, Cano AA, Ghalim N. Comparison between two methods of scorpion venom milking in Morocco. J Venom Anim Toxins Trop Dis. 2013;19(1):5.
7. Vestal ML. High-Performance Liquid Chromatography-Mass Spectrometry. Science. 19 oct 1984;226(4672):275-81.
8. Jenkins TP, Ahmadi S, Bittenbinder MA, Stewart TK, Akgun DE, Hale M, et al. Terrestrial venomous animals, the envenomings they cause, and treatment perspectives in the Middle East and North Africa. Chippaux JP, éditeur. PLoS Negl Trop Dis. 2 déc 2021;15(12):e0009880.
9. Pimenta AMC, Stöcklin R, Favreau P, Bougis PE, Martin-Eauclaire MF. Moving pieces in a proteomic puzzle: mass fingerprinting of toxic fractions from the venom of *Tityus serrulatus* (Scorpiones, Buthidae): Mass fingerprinting of *Tityus serrulatus* toxic fractions. Rapid Commun Mass Spectrom. 15 sept 2001;15(17):1562-72.
10. Diego-García E, Batista CVF, García-Gómez BI, Lucas S, Candido DM, Gómez-Lagunas F, et al. The Brazilian scorpion *Tityus costatus* Karsch: genes, peptides and function. Toxicon. 1 mars 2005;45(3):273-83.

11. Barona J, Batista CVF, Zamudio FZ, Gomez-Lagunas F, Wanke E, Otero R, et al. Proteomic analysis of the venom and characterization of toxins specific for Na⁺- and K⁺-channels from the Colombian scorpion *Tityus pachyurus*. *Biochim Biophys Acta BBA - Proteins Proteomics*. 1 janv 2006;1764(1):76-84.
12. Batista CVF, D'Suze G, Gómez-Lagunas F, Zamudio FZ, Encarnación S, Sevcik C, et al. Proteomic analysis of *Tityus discrepans* scorpion venom and amino acid sequence of novel toxins. *PROTEOMICS*. 2006;6(12):3718-27.
13. Nascimento DG, Rates B, Santos DM, Verano-Braga T, Barbosa-Silva A, Dutra AAA, et al. Moving pieces in a taxonomic puzzle: Venom 2D-LC/MS and data clustering analyses to infer phylogenetic relationships in some scorpions from the Buthidae family (Scorpiones). *Toxicon*. 1 mai 2006;47(6):628-39.
14. Diego-García E, Peigneur S, Clynen E, Marien T, Czech L, Schoofs L, et al. Molecular diversity of the telson and venom components from *Pandinus cavimanus* (Scorpionidae Latreille 1802): Transcriptome, venomics and function. *PROTEOMICS*. 2012;12(2):313-28.
15. Romero-Gutiérrez M, Santibáñez-López C, Jiménez-Vargas J, Batista C, Ortiz E, Possani L. Transcriptomic and Proteomic Analyses Reveal the Diversity of Venom Components from the Vaejovid Scorpion *Serradigitus gertschi*. *Toxins*. 5 sept 2018;10(9):359.
16. Caliskan F, García BI, Coronas FIV, Batista CVF, Zamudio FZ, Possani LD. Characterization of venom components from the scorpion *Androctonus crassicauda* of Turkey: Peptides and genes. *Toxicon*. 1 juill 2006;48(1):12-22.
17. Batista CVF, Román-González SA, Salas-Castillo SP, Zamudio FZ, Gómez-Lagunas F, Possani LD. Proteomic analysis of the venom from the scorpion *Tityus stigmurus*: Biochemical and physiological comparison with other *Tityus* species. *Comp Biochem Physiol Part C Toxicol Pharmacol*. 1 juill 2007;146(1):147-57.
18. Daoudi K, Malosse C, Lafnoune A, Darkaoui B, Chakir S, Sabatier JM, et al. Mass spectrometry-based top-down and bottom-up approaches for proteomic analysis of the Moroccan *Buthus occitanus* scorpion venom. *FEBS Open Bio*. juill 2021;11(7):1867-92.
19. Abdel-Rahman MA, Quintero-Hernandez V, Possani LD. Venom proteomic and venomous glands transcriptomic analysis of the Egyptian scorpion *Scorpio maurus palmatus* (Arachnida: Scorpionidae). *Toxicon*. 1 nov 2013;74:193-207.
20. Erdeş E, Doğan TS, Coşar İ, Danışman T, Kunt KB, Şeker T, et al. Characterization of *Leiurus abdullahbayrami* (Scorpiones: Buthidae) venom: peptide profile, cytotoxicity and antimicrobial activity. *J Venom Anim Toxins Trop Dis*. 2 déc 2014;20:1-8.
21. Estrada-Gómez S, Vargas Muñoz LJ, Saldarriaga-Córdoba M, Quintana Castillo JC. Venom from *Opisthacanthus elatus* scorpion of Colombia, could be more hemolytic and less neurotoxic than thought. *Acta Trop*. 1 janv 2016;153:70-8.
22. Abdel-Rahman MA, Quintero-Hernandez V, Possani LD. Venom proteomic and venomous glands transcriptomic analysis of the Egyptian scorpion *Scorpio maurus palmatus* (Arachnida: Scorpionidae). *Toxicon*. 1 nov 2013;74:193-207.
23. Martin-Eauclaire MF, Bosmans F, Céard B, Diochot S, Bougis PE. A first exploration of the venom of the *Buthus occitanus* scorpion found in southern France. *Toxicon*. 1 mars 2014;79:55-63.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.