



1

2

3

4

5

6 7

8

9

10

19 20

21

39

Proceeding Paper Bee product royal jelly suppress EMT and invasiveness of HCT-116 cells ⁺

Milena Jovanović ^{1,*}, Katarina Virijević ², Dejan Arsenijević ², Katarina Pecić ² and Dragana Šeklić ²

- ¹ Faculty of Science, University of Kragujevac, Serbia; milena.jovanovic@pmf.kg.ac.rs; 5012-2019@pmf.kg.ac.rs ² Institute for Information Technologies, University of Kragujevac, Serbia; msc katarina virijevic@gmail.com;
 - Institute for Information Technologies, University of Kragujevac, Serbia; msc.katarina.virijevic@gmail.com;
 - ddjacic@yahoo.com; katarinapecic13@gmail.com
- * Correspondence: milena.jovanovic@pmf.kg.ac.rs
- + Presented at the 4th International Electronic Conference on Foods: Focus on Sustainable Food Systems: Current Trends and Advances, 15–30 October 2023.

Abstract: The most frequent type of cancer, colorectal cancer (CRC), is widely recognized as the 11 most common cause of death worldwide, due to the high invasive potential of cancer cells enabling 12 metastasis. Cancer cells owe these properties to epithelial-mesenchymal transition (EMT), which 13 requires overexpressed markers Snail and vimentin. Considering that natural products have been 14 intensively investigated from anticancer point of view, we aimed to investigate effects of royal jelly, 15 natural bee product, on invasiveness of colorectal cancer cell line HCT-116 and expression of these 16 two proinvasive/EMT markers. Our study reports on inhibited expression of Snail and vimentin in 17 tested cells, due to which suppressed aforementioned potential was detected. 18

Keywords: EMT; Transwell test; immunofluorescence; collagen

1. Introduction

As the highly frequent type of cancer diagnosed in both males and females, colorectal 22 cancer (CRC), is also widely recognized as the most common cause of death worldwide. 23 Globally it is one of the cancers whose incidence is increasing, and the prognosis of cancer 24 patients' survival is often poor due to the acquisition of invasive and migratory potential 25 of cancer cells which consequently leads to metastasis [1]. Firstly, cancer cells succumb to 26 specific process - epithelial to mesenchymal transition (EMT), acquiring invasive poten-27 tial. This allows them to detach from primary cancer site, therefore, cells are enabled to 28 penetrate in the surrounding stiff extracellular matrix. For this purpose, expression of cer-29 tain markers that enable this transition is necessary to assess, such as Snail, nuclear pro-30 tein, or cytoskeletal protein Vimentin. It is known that Snail expression starts in the first 31 stage of colorectal carcinogenesis, whereat this transcriptional factor acts as repressor of 32 many epithelial hallmarks, as well as potentiator of mesenchymal markers [2]. Hence, 33 therapy designed to battle cancer from this point of cancer progression involves targeting 34 aforementioned markers. 35

We aimed to investigate effects of royal jelly, natural bee product, originated from 36 Serbia, on invasive potential of colorectal cancer cell line HCT-116 and expression of these 37 two invasive EMT proteins. 38

2. Materials and methods

HCT-116 cells, isolated from rectal region of human colon carcinoma, were purchased as immortalized cell line from American Type Culture Collection (ATCC, Manassas, USA). Cells were cultured in complete culturing medium (Dulbecco's Modified Eagle Medium – DMEM, supplemented with 10% of Fetal Bovine Serum – FBS, and penicillin/streptomycin), in humified atmosphere, at 37°C and 5% of CO₂, and seeded for assays

Citation: To be added by editorial staff during production.

Academic Editor: Firstname Lastname

Published: date



Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/). at 80-90% of confluency. Royal jelly sample (produced by *Apis mellifera* L. species) was 1 collected in Central region of Serbia and diluted fresh in Phosphate Buffer Solution (PBS) 2 and complete cell culturing medium. This way two working concentrations 10 and 100 3 μ g/mL were obtained and applied for all the following assays, while the effects of RJ was 4 analyzed after 24 h. 5

Invasive potential of control (untreated) and HCT-116 cells treated with RJ was investigated using assay with Transwell inserts coated with thin collagen layer, which was appied as an extracellular matrix. This assay was performed as described earlier [3]. Absorbances were read using Multiskan SkyHigh Microplate Spectrophotometer (Thermo Scientific, USA) (595 nm wavelength) and results from three independent experiments performed in triplicates are presented in form of invasive index.

In order to assess the protein expression of Snail and Vimentin, immunofluorescent 12 assay was performed in three independent experiments and in triplicates, as described 13 earlier (Jovanovic et al., 2022). Inverted fluorescent microscope Eclipse Ti (Nikon Instruments Inc., Tokyo, Japan) was used to obtain micrographs (at 600× magnification) which 15 were furtherly analyzed to quantify relative fluorescence intensity of targeted proteins. 16 For this purpose, ImageJ software package was applied following the procedure described 17 earlier [4]. 18

Statistical comparison of obtained results was done by IBM SPSS statistical software19package (NY, USA) using One-way Anova test. Results from all performed assays are20presented as mean \pm standard error, where *p < 0.05 designate a statistically significant21difference between treatments and control, and #p < 0.05 designate a statistically significant22cant difference between effects of treatment concentrations.23

3. Results

According to our results showed in Figure 1, the suppression of invasiveness of 25 tested colorectal cancer cell line, HCT-116, was observed 24 h after treatment with two applied RJ concentrations (10 and 100 μ g/mL). This suppression is obvious and statistically significant when compared to control values. 28



Figure 1. Invasive potential of control and HCT-116 cells treated with RJ in two selected concentrations (10 and 100 μ g/mL) presented as mean values ± SE; *p < 0.05 is considered as statistically significant difference between treatments and control values.</th>303132

In order to assess the possible mechanism of RJ's antiinvasive potential, the protein 33 expression of regulatory factor Snail and effector marker vimentin was investigated using 34 immunofluorescent method, and the results can be observed on representative micro-35 graphs, as well as in graphs (Figure 2). Firstly, the expression intensity of Snail in control 36 (untreated) HCT-116 cells was at very high level, and the localization of this protein was 37 in both cell nuclei and cytoplasm, indicating its abundance in cells (Figure 2 a). Mean-38 while, the treatment was able to suppress it significantly and restrict its localization only 39 to cell nuclei. In this suppression, the lower RJ concentration exerted more significant ef-40 fect, than higher concentration applied, which correlates with its antiinvasive potential 41 (Figure 1). 42

24

29

On the other hand, control (untreated) HCT-116 cells contained relatively high level 1 of vimentin, found in both cell nuclei and cytoplasm. When treatment with RJ was ap-2 plied, the significantly lower level of this protein expression was observed, which was mainly localized in cell nuclei (Figure 2 b). The lower RJ concentration had slightly stronger suppressive effect on this protein expression, when compared to higher RJ concentration. These results regarding vimentin protein expression also correlate with anti-6 invasive properties exerted by RJ, as showed earlier (Figure 1). 7



Figure 2. Representative micrographs showing Snail (a) and vimentin (b) fluorescence intensity in 8 control and treated HCT-116 cells. Micrographs were obtained 24 after treatment with RJ in two 9 selected concentrations (10 and 100 µg/mL). Results of relative protein fluorescence intensity are 10 presented as changes compared to control values (mean \pm SE), where *p < 0.05 is considered as sta-11 tistically significant difference between treatments and control values, and #p < 0.05 is considered 12 as statistically significant difference between treatment concentrations. 13

4. Discussion

So far, the common treatments for colon cancer in clinics are chemoradiotherapy, 2 surgery and immunotherapy. However, many disadvantages have been noticed, which is 3 why scientists are turning to development of alternative types of therapeutic approaches 4 to this disease. Natural products have been used in folk medicine for centuries, and many 5 of them proved to be effective in treating cancer [5]. Royal jelly, as natural product, already 6 showed to possess anticancer properties by suppressing cancer growth and aggressive-7 ness [6]. Therefore, we aimed to assess antiinvasive potential of royal jelly sampled in 8 Serbia, on colorectal cancer cell line HCT-116 which is already described as highly inva-9 sive and aggressive [7]. Therefore, the suppression of this cell behavior is highly desirable 10 approach for designing anticancer therapeutics. 11

Present study report on suppression of invasiveness of tested colorectal cancer cells 12 by treatment with RJ, obviously induced by lowered protein level of Snail and Vimentin. 13 Previous investigations confirm that natural products possess the ability to repress levels 14 of these two invasive markers *in vitro*, as well *in vivo* [8]. 15

It is generally accepted that a fundamental characteristic responsible for formation of 16 metastasis is acquisition of invasive potential and is associated with Snail and Vimentin 17 expression [2]. Overexpression of Snail, as zing-fingered transcriptional factor, is respon-18 sible for resistance to chemotherapeutics, lymph node metastasis, activation of EMT pro-19 gram, stemness of cancer and poor prognosis of CRC patients [9,10]. Activation of Vi-20 mentin expression by Snail was already reported, resulting in increased invasive and mi-21 gratory potential of cancer cells, enabling their dissemination (metastasis). Also, the oc-22 currence of this nuclear factor has not been detected in normal (healthy) epithelial cells, 23 however, it was found present in invasive front of cancer tissue [11]. The tight connection 24 between two investigated EMT markers with significant role in acquisition of invasive-25 ness in cancer cells resulted in suppression of this very undesirable trait of HCT-116 cell 26 line. 27

5. Conclusions

RJ exerted significant antiinvasive activity against very aggressive HCT-116 colorec-29tal cell line by attenuating Snail and vimentin invasive markers, which is significant result30of present study, suggesting this natural product as valuable source of anticancer effects.31We anticipate that these findings will be focus of increasing attention in both scientific and32clinical field of research.33

Supplementary Materials:The following supporting information can be downloaded at:34www.mdpi.com/xxx/s1, Figure S1: title;Table S1: title; Video S1: title.35

Author Contributions: Conceptualization, D.Š. and M.J.; methodology, K.V., D.A. and K.P.; software, D.A.; validation, D.Š. and M.J.; formal analysis, K.V., D.A. and K.P.; investigation, K.V., D.A.36and K.P.; resources, D.Š.; data curation, D.Š.; writing — original draft preparation, K.V., D.A. and37K.P.; writing — review and editing, D.Š.; visualization, M.J.; supervision, D.Š.; project administration, D.Š.; funding acquisition, D.Š. All authors have read and agreed to the published version of the manuscript.40

Funding: This research was funded by MINISTRY OF EDUCATION, SCIENCE AND TECHNO-42LOGICAL DEVELOPMENT OF THE REPUBLIC OF SERBIA, grant number 451-03-68/2023-4314/200124 and 451-03-68/2023-14/200122.44

Institutional Review Board Statement: Not applicable.	45
Informed Consent Statement: Not applicable.	46
Data Availability Statement: Not applicable.	47
Acknowledgments: The authors would like to thank Dr Jelena Rakobradović for providing the royal jelly sample.	48 49
Conflicts of Interest: The authors declare no conflict of interest.	50

1

28

References

- Sawicki, T.; Ruszkowska, M.; Danielewicz, A.; Niedźwiedzka, E.; Arłukowicz, T.; Przybyłowicz, K.E. A review of colorectal cancer in terms of epidemiology, risk factors, development, symptoms and diagnosis. *Cancers (Basel)* 2021, 13(9), 2025. doi: 3
 10.3390/cancers13092025.
- Brzozowa, M.; Michalski, M.; Wyrobiec, G.; Piecuch, A.; Dittfeld, A.; Harabin-Słowińska.; M, Boroń, D.; Wojnicz, R. The role of Snail1 transcription factor in colorectal cancer progression and metastasis. *Contemp Oncol (Pozn)* 2015, 19(4), 265-70. doi: 10.5114/wo.2014.42173.
- 3. Brekhman, V.; Neufeld, G. A novel asymmetric 3D in-vitro assay for the study of tumor cell invasion. *BMC cancer* **2009**, *9*, 415. https://doi.org/10.1186/1471-2407-9-415
- Šeklić, D.S.; Stanković, M.S.; Milutinović, M.G.; Topuzović, M.D.; Štajn, A.Š.; Marković, S.D. Cytotoxic, antimigratory and pro/ antioxidative activities of extracts from medicinal mushrooms on colon cancer cell lines. *Arch Biol Sci* 2016, 68(1), 93-105. https://doi.org/10.2298/ABS150427131S
- Šeklić, D.S.; Obradović, A.D.; Stanković, M.S.; Živanović, M.N.; Mitrović, T.L.; Stamenković, S.M.; Marković, S.M. Proapoptotic and antimigratory effects of *Pseudevernia furfuracea* and *Platismatia glauca* on colon cancer cell lines. *Food Technol Biotechnol* 2018, 56(3), 421-30. https://doi.org/10.17113/ftb.56.03.18.5727
- Miyata, Y.; Sakai, H. Anti-cancer and protective effects of royal jelly for therapy-induced toxicities in malignancies. *Int J Mol Sci* 2018, 19(10), 3270. https://doi.org/10.3390/ijms19103270
- 7. Šeklić, D.S.; Đukić, T.; Milenković, D.; Jovanović, M.M.; Živanović, M.N.; Marković, Z.; Filipović, N. Numerical modelling of WNT/β-catenin signal pathway in characterization of EMT of colorectal carcinoma cell lines after treatment with Pt(IV) complexes. *Comput Meth Prog Bio* 2022, 226,107158. https://doi.org/10.1016/j.cmpb.2022.107158
- Ji, Q.; Liu, X.; Han, Z.; Zhou, L.; Sui, H.; Yan, L.; Jiang, H.; Ren, J.; Cai, J.; Li, Q. Resveratrol suppresses epithelial-to-mesenchymal transition in colorectal cancer through TGF-β1/Smads signaling pathway mediated Snail/E-cadherin expression. *BMC Cancer* 2015, *15*(97). https://doi.org/10.1186/s12885-015-1119-y
- Shin, N.R.; Jeong, E.H.; Choi, C.I.; Moon, H.J.; Kwon, C.H.; Chu, I.S.; Kim, G.H.; Jeon, T.Y.; Kim, D.H.; Lee, J.H.; Park, D.Y. Over expression of Snail is associated with lymph node metastasis and poor prognosis in patients with gastric cancer. *BMC Can- cer* 2012, 521. https://doi.org/10.1186/1471-2407-12-521
- Ma, S. Y.; Park, J.H.; Jung, H.; Ha, S.M.; Kim, Y.; Park, D. H.; Lee, D.H.; Lee, S.; Chu, I.H.; Jung, S.Y.; Kim, I.H.; Choi, I.W.; Choi, C.S.; Park, S. Snail maintains metastatic potential, cancer stem-like properties, and chemoresistance in mesenchymal mouse breast cancer TUBO-P2J cells. *Oncol Rep* 2017, 38(3), 1867-1876. doi:10.3892/or.2017.5834
- Krakhmal, N.V.; Zavyalova, M.V.; Denisov, E.V.; Vtorushin, S.V.; Perelmuter, V.M. Cancer invasion: patterns and mechanisms. 30 *Acta Naturae* 2015, 7(2), 17-28.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to 33 people or property resulting from any ideas, methods, instructions or products referred to in the content. 34

1

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

27

28

29