

The Potential Use of Synbiotic Combinations in Bread—A Review †

Gamze Nil Yazici ^{1,*}, Ozum Ozoglu ², Tansu Taspinar ¹, Isilay Yilmaz ‡ and Mehmet Sertac Ozer ¹

¹ Department of Food Engineering, Faculty of Engineering, Cukurova University, 01250 Adana, Turkey; ttaspinar@cu.edu.tr (T.T.); msozer@cu.edu.tr (M.S.O.)

² Department of Food Engineering, Faculty of Agriculture, Bursa Uludag University, 16059 Bursa, Turkey; ozoglu@uludag.edu.tr

* Correspondence: gnboran@cu.edu.tr

† Presented at the 4th International Electronic Conference on Foods, 15–30 October 2023; Available online: <https://foods2023.sciforum.net/>.

‡ Decreased author.

Abstract: To date, the most commonly used probiotics in the potential synbiotic combinations (SC) in breads belong to the family of Lactobacillaceae and Bifidobacteriaceae. However, *Bacillus coagulans* as a heat-resistant bacteria, and *Saccharomyces boulardii* as a probiotic yeast could promising to use of SC in bread. Inulin is the most commonly direct-used prebiotic source in formulations of potential SC bread. Moreover, co-encapsulations of probiotics with prebiotics mainly including different hydrocolloids could be beneficial. Although the consumption of potential SC in bread generally including *Lactobacillus sporogenes* and inulin by diabetics had some health benefits, there is a need for more comprehensive clinical trials.

Keywords: *Bacillus coagulans*; hydrocolloid; co-encapsulation; edible film; type 2 diabetes

1. Introduction

Nowadays, the development of functional food products including probiotics, prebiotics, and synbiotics, which have an important role in the diet in terms of protecting and/or improving human health [1], come into prominence. Among those, synbiotics as a combination of probiotics and prebiotics have a synergetic health-promoting influence. In this regard, the synbiotic foods mainly include dairy (cheese, yoghurt, ice cream, and fermented milk, etc.), and non-dairy food products, such as confectioneries (chocolate, candy, etc.) [2], fermented or unfermented beverages (such as cereal-, legume- or fruit-based drinks, etc.) [3,4], and other food products such as mousse, salad dressing, and snacks [2]. However, there are limited studies in the literature regarding the potential synbiotic combinations in baked goods particularly bread as a staple food.

Inulin is the major directly-utilized prebiotic source in potential synbiotic combinations in bread, as seen in Table 1. However, there is a challenge in the production of a synbiotic bread which is generally related to the heat sensitivity of probiotic bacteria during the baking process. To protect the probiotic bacteria regarding their viability and stability during the heating process, they should be encapsulated and then added to the food structure, or should be covered with an edible film/coating on the bread crust structure [5]. In addition, PRO-PRE co-encapsulation, in other terms co-delivery or co-entrapment, of probiotics with prebiotics is a promising application regarding both stability and viability of living probiotics [6]. Moreover, recently it is revealed that some probiotic microorganisms particularly *Bacillus coagulans* could be also a promising for maintaining probiotic viability throughout thermal processes [7]. Although *B. coagulans*, which is a spore-forming probiotic [8], has resistance and/or tolerance to heat, conditions of the

Citation: Yazici, G.N.; Ozoglu, O.; Taspinar, T.; Yilmaz, I.; Ozer, M.S. The Potential Use of Synbiotic Combinations in Bread—A Review. *Biol. Life Sci. Forum* **2023**, *26*, x. <https://doi.org/10.3390/xxxxx>

Academic Editor(s): Name

Published: date



Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

gastrointestinal system, and also is in GRAS status and is declared as safe by EFSA and FDA [9], it did not have adequate coverage in synbiotic combinations in breads, as seen in Table 1. Similarly, although *Saccharomyces cerevisiae* var. *boulardii* is the only probiotic yeast and is used in several different food products [10], there is no study in the literature based on it as a probiotic source of synbiotic combinations in breads, as far as we know.

2. The Potential Synbiotic Combinations in Breads

2.1. The Influence of Potential Synbiotic Combinations in Breads on Probiotic Viability

Up to now, the most commonly used probiotic bacteria in potential synbiotic combinations in breads are *Lactobacillus acidophilus*, *Lacticaseibacillus casei*, *Lactiplantibacillus plantarum*, and *Lacticaseibacillus rhamnosus*, which belong to Lactobacillaceae family and also *Bifidobacterium animalis*, *Bifidobacterium adolescentis* and *Bifidobacterium longum* which are species of Bifidobacteriaceae family (Table 1). Moreover, the mostly utilized wall materials are high-amylose maize starch (Hi-maize), chitosan, and some hydrocolloids such as pectin, xanthan gum, gum arabic, gellan gum, tragacanth gum, carboxymethyl cellulose, hydroxypropyl methylcellulose for preparing co-encapsulated probiotic bacterial strains in potential synbiotic combinations of breads, as shown in Table 1.

Un-encapsulated *L. rhamnosus* lost its viability after bread baking [11,12]. The viability of *L. rhamnosus* was increased by nearly 3% when incorporating baobap pulp (BP) into the culture medium. The inclusion of baobap pulp has a prebiotic potential in bread formulation resulting in a further increase in probiotic viability by about 3%. Those were interrelated with its higher pectin content which enhanced the metabolic activities of *L. rhamnosus* and also its interaction with BP due to an increase in hydrogen bonds, electrostatic forces, and steric hindrance. Throughout the resident time of simulated gastric (90 min) and intestinal phase (180 min), the viability of encapsulated probiotics were significantly increased in BP-enriched bread. Further increase was also determined with pre-cultured probiotics with BP powder. At the end of the simulated gastric phase, the viability of encapsulated probiotic was increased by nearly 6–11.5%. This was ascribed to the proliferation of probiotics because of the increase in carbon sourced from starch and phenolic contents of BP [11]. In another study, the number of viable encapsulated *L. rhamnosus* with different wall materials (sodium alginate, high-amylose maize starch, cassava starch, and chitosan) was in the range of nearly 4.94–9.2 log CFU/g after baking at different baking conditions (180 °C-30 min, 220 °C-20 min, 250 °C-15 min). The triple-layered capsules composed of sodium alginate combined with high amylose-resistant starch and chitosan had the highest probiotic viability after baking and in the simulated gastrointestinal system [12]. In simulated gastric conditions, the highest relative viability of *L. acidophilus* was obtained in the first layer composed of alginate using at 1%, and then, chitosan was significantly more effective than alginate as a second layer coating material at the same concentrations in fresh and 1-day stored bread. However, the relative viability of double-layered encapsulated probiotics was decreased with short-time storage (1 day) at room temperature irrespective of coating material [13]. The viability of probiotics (*L. acidophilus*, *L. plantarum*) irrespective of strain was increased by microencapsulation independent of wall material throughout gluten-free bread baking and also storage ongoing more than 2 logarithmic cycles. However, this influence was more pronounced in encapsulated probiotics by tragacanth gum which is attributed to its higher prebiotic effect than sago starch [14]. In a similar vein, the viability of *L. casei* in the edible films based on konjac glucomannan was gradually reduced with a decrease of 2 log CFU/portion after 7-day storage at room temperature [15]. In another study, the higher probiotic viability in the edible coating was obtained in the combination of whey and sodium alginate than in high amylose maize starch and gelatin [16]. Although inulin concentration did not make a significant difference in *Bacillus coagulans* count in fresh and stored bread, it was more than 6 log CFU/ which was still met the minimum level complying with the WHO recommendation to provide health benefits [5].

2.2. The Influence of Potential Symbiotic Combinations in Breads on Technological Properties

The different encapsulation wall materials (sodium alginate individually, double or triple layered with chitosan, cassava starch, and Hi-maize resistant starch) for *L. rhamnosus* did not make a significant change in weight and specific volume of breads, and also its nutritional value (protein, fat, carbohydrate, ash, and fiber content) [12]. The higher specific volume and oven spring values in gluten-free bread including probiotics encapsulated with tragacanth gum were recorded. This could be explained by the hydroxyl group of tragacanth gum which has the capability of water absorption. Moreover, the hardness values did not significantly affect by probiotic sources (*L. acidophilus*, *L. plantarum*), but did by encapsulation wall material. In this regard, the softest in other terms the lowest hardness values were belong the gluten-free breads with probiotics encapsulated by tragacanth gum individually which is followed by using it combined with sago starch [14]. A significant moisture reduction was observed with increasing inulin levels in fresh bread consistent with water absorption values obtained from the farinograph. Increasing in content of inulin as a prebiotic source in bread formulation, resulted in a significant reduction in specific volume values. This could be attributed to the prohibition of proper expansion which is associated with gluten dilution, disruption of gluten-starch matrix, and lower water vapor because of the lower moisture content of the bread dough. A significant increase in hardness values in bread with increasing concentration of inulin was observed and several potential reasons were stated such as the formation of cross-linkages between inulin and starch and/or protein which led to a decrease in gas retention capacity, recrystallization of inulin while cooling, co-crystallization of inulin and also amylopectin, strengthen the solids around the gas cells and lower moisture content with inulin addition. While the *L* and *b* values of the crust part of the bread were significantly reduced with inulin addition, an opposite effect was observed in *a* values that resulted in darker and reddish color. This is related to a decrease in moisture content and partial hydrolyzation of inulin to glucose and fructose during the baking process which develops crust color via Maillard reaction and caramelization [5].

Table 1. The potential use of synbiotic combinations in bread.

| Product | Probiotic Source(s) | Prebiotic or Potential Prebiotic Source(s) | References |
|-----------------------------|---|---|------------|
| Bread | <i>Lactobacillus acidophilus</i> | Inulin ^{a, b} , carboxymethylcellulose ^b , pectin ^{a, b} , fresh agave sap ^{a, b} | [17] |
| Bread | <i>Lactobacillus acidophilus</i> | Xanthan gum ^b , gellan gum ^b , chitosan ^b | [13] |
| Cream bread | <i>Lactobacillus acidophilus</i> | Xanthan gum ^b , maltodextrin ^b | [18] |
| Gluten-free "Barbari" bread | <i>Lactobacillus acidophilus</i> , <i>Lactiplantibacillus plantarum</i> | Tragacanth gum ^b , sago starch ^b | [14] |
| Bread | <i>Lactiplantibacillus plantarum</i> | Inulin ^b , gum arabic ^{a, b} , maltodextrin ^{a, b} | [19] |
| Pan bread | <i>Lacticaseibacillus rhamnosus</i> | Baobab pulp ^a , high- amylose maize starch ^b , chitosan ^b | [11] |
| Pan bread | <i>Lacticaseibacillus rhamnosus</i> | High-amylose maize starch ^b , cassava starch ^b , chitosan ^b | [12] |
| Pan bread, hamburger bread | <i>Lactobacillus acidophilus</i> , <i>Lacticaseibacillus casei</i> | Inulin ^a , high-amylose maize starch ^b , chitosan ^b | [20] |
| Bread bun | <i>Lacticaseibacillus casei</i> | Inulin ^b , konjac glucomannan ^b | [15] |
| Bread | <i>Streptococcus salivarius</i> subsp. <i>thermophilus</i> , <i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> , <i>Lactobacillus acidophilus</i> , <i>Acetobacter aceti</i> , <i>Bifidobacterium bifidum</i> , <i>Bifidobacterium adolescentis</i> , <i>Bifidobacterium longum</i> , | Whey, glycerol ^b ; high amylose maize starch ^b | [16] |

Bifidobacterium animalis, *Lactobacillus acidophilus*,
Lactococcus lactis subsp. *cremoris*,
Propionibacterium freudenreichii, *Enterococcus*
faecium, *Streptococcus salivarius* subsp.
thermophilus

| | | | |
|----------------------|--|--------------------------------------|------|
| Bread | <i>Bacteroides ovatus</i> , <i>Bifidobacterium adolescentis</i> | Arabinoxylan ^a | [21] |
| Bread | <i>Lactobacillus acidophilus</i> , <i>Bifidobacterium animalis</i> | Apple pomace ^a | [22] |
| Bread | <i>Bifidobacterium animalis</i> spp. <i>lactis</i> | Hydroxypropyl cellulose ^b | [23] |
| Steamed bread | <i>Bifidobacterium longum</i> | Gellan gum ^b | [24] |
| Bread | <i>Bacillus coagulans</i> | Inulin ^a | [5] |

a: direct usage, b: coating.

2.3. The Influence of Potential Synbiotic Combinations in Breads on Human Health

There are limited clinical trials based on the potential synbiotic bread consumption on human health which is mainly focused on the effect on patients with type 2 diabetes mellitus (T2DM) regarding insulin metabolism and serum high-sensitivity C-reactive protein [25], blood lipids [26,27], apolipoproteins [26], nitric oxide, biomarkers of oxidative stress, and serum liver enzymes [28]. According to results of an randomized, double-blind, controlled clinical trial, the consumption of synbiotic bread consisting of *L. sporogenes* and inulin through 3 times/day in a 40 g per serve for 8 weeks by T2DM patients ($n = 27$), led to a significant decrease in the levels of serum insulin, and thus beneficial for insulin metabolism [25]. Ghafouri et al. [26] revealed that a decrease in total cholesterol levels and Apo A1 was observed in the patients with T2DM ($n = 25$) who consumed synbiotic bread, which is composed of *B. coagulans*, β -glucan, and inulin, for 3 times in a day for 8 weeks. However, while triacylglycerol(s), and very low-density lipoprotein cholesterol were decreased, the high-density lipoprotein-cholesterol was increased, but the levels of total cholesterol and low-density lipoprotein-cholesterol were not significantly influenced ($p > 0.05$) with consumption of synbiotic bread including *Lactobacillus sporogenes* and inulin by diabetic patients ($n = 26$) [27]. The consumption of a total of 120 g per day of the same content of synbiotic bread for 8 weeks gave rise to a significant increase in nitric oxide, and a decrease in malondialdehyde was observed in patients with diabetes ($n = 27$). On the other side, no significant difference was defined in terms of blood pressure, liver enzymes, plasma glutathione, plasma total antioxidant capacity, and the levels of iron, calcium, and magnesium [28].

3. Conclusions

To sum up, the following studies should focus on the survival of more probiotic microorganisms, especially *Bacillus coagulans* and *Saccharomyces boulardii*, optimization of different encapsulation techniques, wall materials, film/coatings together with different types and concentrations of prebiotic sources used in other cereal-based food products, and also in gluten-free bread. Moreover, the viability of probiotics with prebiotics which are used directly, or as a wall material for encapsulation or edible film/coating should be assessed from a holistic perspective regarding the nutritional, technological, and sensorial properties of bread.

Author Contributions: Conceptualization, G.N.Y., O.O., T.T., I.Y.; writing—original draft preparation, G.N.Y.; writing—review and editing, G.N.Y., O.O., T.T. and M.S.O.; project administration, M.S.O.; funding acquisition, M.S.O. All authors have read and agreed to the published version of the manuscript.

Funding: This study was funded by Cukurova University, Turkey, as part of the FDK-2021-14015 project.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Tadesse, S. Probiotics, prebiotics and synbiotics as functional food ingredients: Production, health benefits and safety. *J. Biol. Act. Prod. Nat.* **2012**, *2*, 124–134. <https://doi.org/10.1080/22311866.2012.10719119>.
2. González-Herrera, S.M.; Bermúdez-Quiñones, G.; Ochoa-Martínez, L.A.; Rutiaga-Quiñones, O.M.; Gallegos-Infante, J.A. Synbiotics: A technological approach in food applications. *J. Food Sci. Technol.* **2021**, *58*, 811–824. <https://doi.org/10.1007/s13197-020-04532-0>.
3. Kumar, A.; Tomer, V.; Kaur, A.; Joshi, V.K. Synbiotics: A culinary art to creative health foods. *Int. J. Food Ferment. Technol.* **2015**, *5*, 1–14. <https://doi.org/10.5958/2277-9396.2015.00001.x>.
4. Chaturvedi, S.; Chakraborty, S. Review on potential non-dairy synbiotic beverages: A preliminary approach using legumes. *Int. J. Food Sci. Technol.* **2021**, *56*, 2068–2077. <https://doi.org/10.1111/ijfs.14779>.
5. Majzoobi, M.; Aghdam, M.B.K.; Eskandari, M.H.; Farahnaky, A. Quality and microbial properties of symbiotic bread produced by straight dough and frozen part-baking methods. *J. Texture Stud.* **2019**, *50*, 165–171. <https://doi.org/10.1111/jtxs.12386>.
6. Rashidinejad, A.; Bahrami, A.; Rehman, A.; Rezaei, A.; Babazadeh, A.; Singh, H.; Jafari, S.M. Co-encapsulation of probiotics with prebiotics and their application in functional/synbiotic dairy products. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 2470–2494. <https://doi.org/10.1080/10408398.2020.1854169>.
7. Zhao, N.; Yu, T.; Yan, F. Probiotic role and application of thermophilic *Bacillus* as novel food materials. *Trends Food Sci. Technol.* **2023**, *138*, 1–15. <https://doi.org/10.1016/j.tifs.2023.05.020>.
8. Adibpour, N.; Hosseini-zhad, M.; Pahlevanlo, A.; Hussain, M.A. A review on *Bacillus coagulans* as a spore-forming probiotic. *Appl. Food Biotechnol.* **2019**, *61*, 91–100. <https://doi.org/10.22037/afb.v%vi%i.23958>.
9. Konuray, G.; Erginkaya, Z. Potential use of *Bacillus coagulans* in the food industry. *Foods* **2018**, *7*, 92. <https://doi.org/10.3390/foods7060092>.
10. Chan, M.Z.A.; Liu, S.Q. Fortifying foods with synbiotic and postbiotic preparations of the probiotic yeast, *Saccharomyces boulardii*. *Curr. Opin. Food Sci.* **2022**, *43*, 216–224. <https://doi.org/10.1016/j.cofs.2021.12.009>.
11. Adedeji, O.E.; Okechie, I.D.; Ezekiel, O.O. Prebiotic influence of baobab pulp on the stability of *Lactobacillus rhamnosus* GG in white-pan bread. *J. Food Meas. Charact.* **2022**, *16*, 2221–2228.
12. Ezekiel, O.O.; Okechie, I.D.; Adedeji, O.E. Viability of *Lactobacillus rhamnosus* GG in simulated gastrointestinal conditions and after baking white pan bread at different temperature and time regimes. *Curr. Microbiol.* **2020**, *77*, 3869–3877. <https://doi.org/10.1007/s00284-020-02203-z>.
13. Mirzamani, S.S.; Bassiri, A.R.; Tavakolipour, H.; Azizi, M.H.; Kargozari, M. Survival of fluidized bed encapsulated *Lactobacillus acidophilus* under simulated gastro-intestinal conditions and heat treatment during bread baking. *J. Food Meas. Charact.* **2021**, *15*, 5477–5484. <https://doi.org/10.1007/s11694-021-01108-0>.
14. Ghasemi, L.; Nouri, L.; Mohammadi Nafchi, A.; Al-Hassan, A.A. The effects of encapsulated probiotic bacteria on the physicochemical properties, staling, and viability of probiotic bacteria in gluten-free bread. *J. Food Process. Preserv.* **2022**, *46*, 1–11. <https://doi.org/10.1111/jfpp.16359>.
15. Pruksarojanakul, P.; Prakitchaiwattana, C.; Settachaimongkon, S.; Borompichaichartkul, C. Synbiotic edible film from konjac glucomannan composed of *Lactobacillus casei*-01[®] and Orafti[®]GR, and its application as coating on bread buns. *J. Sci. Food Agric.* **2020**, *100*, 2610–2617. <https://doi.org/10.1002/jsfa.10287>.
16. Gregirchak, N.; Stabnikova, O.; Stabnikov, V. Application of lactic acid bacteria for coating of wheat bread to protect it from microbial spoilage. *Plant Foods Hum. Nutr.* **2020**, *75*, 223–229. <https://doi.org/10.1007/s11130-020-00803-5>.
17. Altamirano-Fortoul, R.; Moreno-Terrazas, R.; Quezada-Gallo, A.; Rosell, C.M. Viability of some probiotic coatings in bread and its effect on the crust mechanical properties. *Food Hydrocoll.* **2012**, *29*, 166–174. <https://doi.org/10.1016/j.foodhyd.2012.02.015>.
18. Duc Thang, T.; Hanh Quyen, L.T.; Thuy Hang, H.T.; Thien Luan, N.; KimThuy, D.T.; Lieu, D.M. Survival survey of *Lactobacillus acidophilus* in additional probiotic bread. *Turkish J. Agric. -Food Sci. Technol.* **2019**, *7*, 588–592. <https://doi.org/10.24925/turjaf.v7i4.588-592.2247>.
19. Zhang, L.; Chen, X.D.; Boom, R.M.; Schutyser, M.A.I. Survival of encapsulated *Lactobacillus plantarum* during isothermal heating and bread baking. *Lwt* **2018**, *93*, 396–404. <https://doi.org/10.1016/j.lwt.2018.03.067>.
20. Seyedain-Ardabili, M.; Sharifan, A.; Tarzi, B.G. The production of synbiotic bread by microencapsulation. *Food Technol. Biotechnol.* **2016**, *54*, 52–59. <https://doi.org/10.17113/ftb.54.01.16.4234>.
21. Zhang, D.; Rudjito, R.C.; Pietiäinen, S.; Chang, S.C.; Idström, A.; Evenäs, L.; Vilaplana, F.; Jiménez-Quero, A. Arabinoxylan supplemented bread: From extraction of fibers to effect of baking, digestion, and fermentation. *Food Chem.* **2023**, *413*. <https://doi.org/10.1016/j.foodchem.2023.135660>.
22. Jagelaviciute, J.; Staniulyte, G.; Cizeikiene, D.; Basinskiene, L. Influence of enzymatic hydrolysis on composition and technological properties of apple pomace and its application for wheat bread making. *Plant Foods Hum. Nutr.* **2023**. <https://doi.org/10.1007/s11130-023-01054-w>.

23. Penhasi, A.; Reuveni, A.; Baluashvili, I. Microencapsulation may preserve the viability of probiotic bacteria during a baking process and digestion: A case study with *Bifidobacterium animalis* subsp. *lactis* in bread. *Curr. Microbiol.* **2021**, *78*, 576–589. <https://doi.org/10.1007/s00284-020-02292-w>.
24. Yang, Z.; Li, M.; Li, Y.; Wang, X.; Li, Z.; Shi, J.; Huang, X.; Zhai, X.; Zou, X.; Gong, Y.; et al. Entrapment of probiotic (*Bifidobacterium longum*) in bilayer emulsion film with enhanced barrier property for improving viability. *Food Chem.* **2023**, *423*, 136300. <https://doi.org/10.1016/j.foodchem.2023.136300>.
25. Tajadadi-Ebrahimi, M.; Bahmani, F.; Shakeri, H.; Hadaegh, H.; Hijjafari, M.; Abedi, F.; Asemi, Z. Effects of daily consumption of synbiotic bread on insulin metabolism and serum high-sensitivity C-reactive protein among diabetic patients: A double-blind, randomized, controlled clinical trial. *Ann. Nutr. Metab.* **2014**, *65*, 34–41. <https://doi.org/10.1159/000365153>.
26. Ghafouri, A.; Heshmati, J.; Heydari, I.; Shokouhi Shoormasti, R.; Estêvão, M.D.; Hoseini, A.S.; Morvaridzadeh, M.; Akbari-Fakhrabadi, M.; Farsi, F.; Zarrati, M.; et al. Effect of synbiotic bread containing lactic acid on blood lipids and apolipoproteins in patients with Type 2 diabetes: A randomized controlled trial. *Food Sci. Nutr.* **2022**, *10*, 4419–4430. <https://doi.org/10.1002/fsn3.3039>.
27. Shakeri, H.; Hadaegh, H.; Abedi, F.; Tajadadi-Ebrahimi, M.; Mazrooi, N.; Ghandi, Y.; Asemi, Z. Consumption of synbiotic bread decreases triacylglycerol and VLDL levels while increasing HDL levels in serum from patients with Type-2 diabetes. *Lipids* **2014**, *49*, 695–701. <https://doi.org/10.1007/s11745-014-3901-z>.
28. Bahmani, F.; Tajadadi-Ebrahimi, M.; Kolahdooz, F.; Mazouchi, M.; Hadaegh, H.; Jamal, A.S.; Mazrooi, N.; Asemi, S.; Asemi, Z. The Consumption of synbiotic bread containing *Lactobacillus sporogenes* and inulin affects nitric oxide and malondialdehyde in patients with Type 2 diabetes mellitus: Randomized, double-blind, placebo-controlled trial. *J. Am. Coll. Nutr.* **2016**, *35*, 506–513. <https://doi.org/10.1080/07315724.2015.1032443>.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.