



Proceeding Paper Plant-Derived Triterpenoid Functionalization: Synthesis of α-Acyloxycarboxamides ⁺

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Abstract: The application of isocyanide-based multicomponent reactions (IMCRs) for triterpenoid functionalization is few reported. Triterpenoids and their derivatives are an important class of natural products of interest in medicinal chemistry due to their potential applications as antibacterial, antifungal, and cytotoxic agents. Herein, we describe the use of ethanol as solvent in the Passerini reaction for the functionalization of masticadienonic acid isolated from fruits and peduncles of *P. mexicana*. A small series of α -acyloxycarboxamides was synthesized with moderate to good overall yields of 33 to 57%, evaluating and extending the scope of the aldehyde component.

Keywords: triterpenoid; IMCR; Passerini-3CR; *α*-acyloxycarboxamide

1. Introduction

Triterpenoids are a large class of plant-derived natural products, with an inherent structural diversity [1]. They are widely distributed in the plant kingdom, especially in higher plants [2]. Approximately 55,000 compounds belonging to this group have been identified, however, very few have been investigated for its therapeutic applications [3]. The prevention and treatment of cancer is the most studied property of triterpenoids. Other important applications include antimicrobial, antiviral, antifungal, antiparasitic, anti-inflammatory, etc. [4–9].

Multicomponent reactions (MCRs) are synthetic procedures where three or more substances react to form a product which contains all or most of the atoms from the starting materials [10]. In these procedures, a complex molecule is assembled in a one pot chemical step, providing a huge chemical diversity [11].

Among the diverse class of MCRs, those involving isocyanide reagents were some of the first to be discovered. Mario Passerini reported in 1921 the first isocyanide-based multicomponent reaction (IMCR), employing aryl isocyanides, ketones, and carboxylic acids to afford α -acyloxycarboxamides [12]. Almost 40 years later, Ivar Ugi presented his four-component reactions in which an isocyanide, a carbonyl compound, an amine, and an isocyanide react to afford *bis*-amides [13].

In the field of combinatorial chemistry, the IMCRs are an important and versatile tool which have several advantages such as atom economy, convergent design, easiness of performance, and the generation of molecular diversity [14]. On the other hand, IMCR products can be versatile synthetic platforms for further structural diversification [15].

In recent years, IMCRs have been applied to the rapid and efficient functionalization of natural products, especially steroids, peptides, and glycosides [16]. However, there are only a few reports on the use of triterpenoids as components in IMCRs. For the case of the

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Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/). Passerini reaction, there are two reports where either naturally functionalized triterpenoids or their derivatives are used as components [17,18]. Triterpenoids and their amide derivatives have been extensively studied over the last decades as an alternative for cancer treatment, due to their important cytotoxic properties [19].

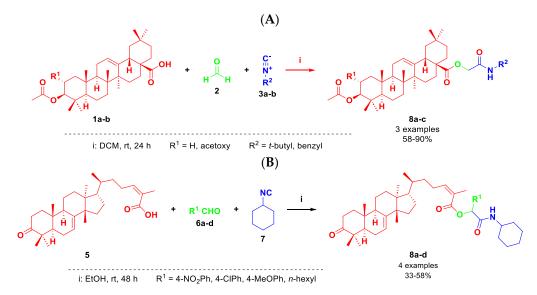


Figure 1. (A). Previous work by Wiemann, J. et al. (2018) [18], (B). This work.

2. Results and Discussion

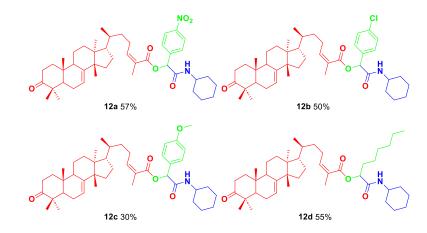
In the present work, we report the synthesis of a series of α -acyloxycarboxamides employing a plant-derived triterpenoid as component in the Passerini three-component reactions. Masticadienonic acid (5) isolated from hexane extracts of fruits and peduncles of dried *P. mexicana*, along with 4-nitrobenzaldehyde (6a) and cyclohexyl isocyanide (7) were used as components for reaction optimization to synthesize target molecule 8a. The performed experiments are depicted in Table 1.

	$ \begin{array}{c} 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \begin{array}{c} 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 $		NO ₂ H NO ₂ H N N N N N N N N N N N N N N N N N N
Entry	Solvent	Time	Yield
1	MeOH	48 h	30%
2	EtOH	48 h	57%
3	H ₂ O	72 h	NR

Table 1. Screening conditions for the synthesis of target molecule 16a.

In previous reports from our research group, dichloromethane has been used as solvent for Passerini reaction, however, to improve the greenness of our procedures, some alternative solvents have been evaluated. There are some reports of the use of polar protic solvents for this methodology, for example methanol, however this experiment led to a longer reaction time and a low yield of 30%. The starting material was not fully consumed, which might be a consequence of the poor solubility of compound **5**. Another experiment using ethanol afforded better yields (57%) since **5** is fairly soluble in this solvent. Finally, an experiment using water did not afford the desired product under the evaluated conditions.

Using the best conditions found in these experiments, a small library of α -acyloxycarboxamides was synthesized, employing aromatic aldehydes with electron-donor and electron-withdrawing groups, as well as an aliphatic aldehyde.



3. Experimental Section

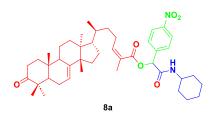
3.1. General Information, Chemicals and Instrumentation

Bruker Avance III spectrometers (500 and 125 MHz, respectively) were used for ¹H and ¹³C NMR spectra acquisition. Deuterated chloroform (CDCl3) was used as the solvent for NMR experiments. Chemical shifts (δ) are given in ppm relative to tetramethylsilane (TMS). Coupling constants are reported in Hertz (Hz). Multiplicities of the signals are described using standard abbreviations: singlet (s), doublet (d), triplet (t), quartet (q), and multiplet (m). NMR spectra were analyzed using MestReNova software version 12.0.0-20080. Reaction progress was monitored by thin-layer chromatography (TLC) on precoated silica gel F254 aluminum sheets. The spots were visualized under UV light at 254 nm. Column chromatography was performed using silica gel (230–400 mesh) as stationary phase. Mixtures of hexanes and ethyl acetate were used as mobile phase for column chromatography and in TLC for reaction progress monitoring and measuring retention factors (R_i). All reagents were purchased from Sigma Aldrich and were used without further purification. Chemical names and drawings were obtained using the ChemDraw 22.2.0.3300 software package.

3.2. General Procedure

Masticadienonic acid (5, 1.0 equiv.), aldehyde **6a–d** (1.0 equiv.) and cyclohexyl isocyanide (7, 1.0 equiv.) were dissolved in ethanol (0.5 M) and placed in a sealed vial with a magnetic stir bar. The mixture was stirred at room temperature for 48 h. Then, solvent was removed under reduced pressure, and the crude reaction mixture was purified by column chromatography, using silica gel as stationary phase and a mixtures of ethyl acetate in hexanes, to afford the corresponding α -acyloxycarboxamides **8a–d**.

3.3. Spectral Data

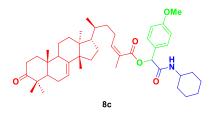


2-(cyclohexylamino)-1-(4-nitrophenyl)-2-oxoethyl-3-oxotirucalla-7,24Z-dien-26oate (8a). White solid; Rf = 0.3 (20% ethyl acetate in hexanes): ¹H NMR (500 MHz, CDCl₃,

25 °C, TMS): δ 8.23 (m, 2H), 7.63 (m, 2H), 6.14 (s, 1H), 6.10 (td, *J* = 7.6, 1.6 Hz, 1H), 6.00 (s, 1H), 5.30 (dd, *J* = 6.1, 3.1 Hz, 1H), 3.84 (dtt, *J* = 10.4, 7.8, 4.0 Hz, 1H), 2.76 (td, *J* = 14.5, 5.5 Hz, 1H), 2.59 (m, 1H), 2.45 (m, 1H), 2.28 (dt, *J* = 14.1, 3.8, 1H), 2.25 (m, 1H), 2.10 (m, 2H), 2.07 (d, *J* = 1.5 Hz, 3H), 1.99 (m, 2H), 1.98 (m, 1H), 198 (m, 4H), 1.81 (m, 2H), 1.73 (t, *J* = 8.7 Hz, 1H), 1.69 (m, 2H), 1.64 (m, 1H), 1.56 (m, 2H), 1.53 (m, 2H), 1.49 (m, 1H), 1.48 (m, 2H), 1.48 (m, 1H), 1.40 (m, 1H), 1.28 (m, 1H), 1.18 (m, 4H) 1.14 (m, 1H), 1.12 (s, 3H), 1.05 (s, 3H), 1.01 (s, 3H), 0.89 (d, *J* = 6.2 Hz, 3H), 0.81 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 216.8, 166.3, 165.0, 148.9, 146.6, 145.88, 143.2, 127.9, 125.2, 123.7, 117.8, 74.2, 52.9, 52.3, 51.1, 48.4, 47.8, 47.7, 43.5, 38.5, 36.0, 35.6, 35.0, 34.9, 34.0, 33.63, 32.9, 28.2, 27.4, 26.9, 25.4, 24.6, 24.5, 24.3, 21.9, 21.6, 20.6, 18.2, 12.7.



2-(cyclohexylamino)-1-(4-chlorophenyl)-2-oxoethyl-3-oxotirucalla-7,24Z-dien-26-oate (<u>8b</u>). Yellow oil; Rf = 0.31 (20% ethyl acetate in hexanes): ¹H NMR (500 MHz, CDCl₃, 25 °C, TMS): δ 7.40 (m, 2H), 7.35 (m, 2H), 6.04 (d, *J* = 8.2 Hz, 1H), 6.00 (dt, *J* = 7.6, 1.6 Hz, 1H), 5.98 (d, *J* = 2.6 Hz, 1H), 5.31 (dd, *J* = 6.0, 3.2 Hz, 1H), 3.85 (dtt, *J* = 10.5, 7.8, 4.1 Hz, 1H), 2.77 (td, *J* = 14.5, 5.4 Hz, 1H), 2.58 (m, 1H), 2.45 (m, 1H), 2.29 (dt, *J* = 14.1, 3.7, 1H), 2.24 (m, 1H), 2.10 (m, 2H), 1.99 (m, 2H), 1.98 (m, 1H), 1.98 (d, *J* = 1.6 Hz, 3H), 1.93 (m, 4H), 1.82 (m, 1H), 1.73 (t, *J* = 8.7 Hz, 1H), 1.69 (m, 2H) 1.65 (m, 1H), 1.56 (m, 2H), 1.53 (m, 2H), 1.51 (m, 1H), 1.48 (m, 2H), 1.47 (m, 1H), 1.40 (m, 1H), 1.36 (s, 9H), 1.29 (m, 1H), 1.18 (m, 4H), 1.14 (m, 1H), 1.12 (s, 3H), 1.06 (s, 3H), 1.01 (s, 3H), 1.00 (s, 3H), 0.89 (d, *J* = 6.2 Hz, 3H), 0.80 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 216.8, 167.3, 165.7, 146.6, 145.9, 135.0, 134.5, 129.1, 128.8, 125.2, 117.9, 73.9, 52.9, 52.3, 51.2, 48.5, 47.9, 47.7, 43.5, 38.5, 36.1, 35.7, 35.0, 34.9, 34.1, 33.6, 33.0, 28.3, 27.4, 27.0, 25.4, 24.7, 24.5, 24.4, 22.0, 21.6, 20.7, 18.2, 12.8.



2-(cyclohexylamino)-1-(4-methoxyphenyl)-2-oxoethyl-3-oxotirucalla-7,24Z-dien-

26-oate (8c). Yellow oil; Rf = 0.38 (20% ethyl acetate in hexanes): ¹H NMR (400 MHz, CDCl₃, 25 °C, TMS): δ 7.34 (m, 2H), 6.87 (m, 2H), 6.09 (dt, *J* = 7.5, 1.2 Hz, 1H), 6.02 (bs, 1H), 5.98 (d, *J* = 2.9 Hz, 1H), 5.31 (dd, *J* = 6.1, 3.4 Hz, 1H), 3.86 (dtt, *J* = 10.5, 7.7, 4.0 Hz, 1H), 3.80 (s, 3H), 2.74 (td, *J* = 14.5, 5.4 Hz, 1H), 2.59 (m, 1H), 2.44 (m, 1H), 2.28 (dt, *J* = 14.1, 3.8, 1H), 2.25 (m, 1H), 2.11 (m, 2H), 1.98 (m, 1H), 1.99 (m, 1H), 1.97 (d, *J* = 1.5 Hz, 3H), 193 (m, 4H), 1.81 (m, 1H), 1.73 (t, *J* = 8.7 Hz, 1H), 169 (m, 2H), 1.65 (m, 1H), 1.56 (m, 2H), 1.55 (m, 2H), 1.49 (m, 1H), 1.48 (m, 2H), 1.48 (m, 1H), 1.41 (m, 1H), 1.36 (s, 9H), 1.28 (m, 1H), 1.18 (M, 4H), 1.15 (m, 1H), 1.12 (s, 3H), 1.05 (s, 3H), 1.01 (s, 3H), 0.99 (s, 3H), 0.89 (d, *J* = 6.2 Hz, 3H), 0.81 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 216.8, 166.3, 165.7, 159.8, 146.8, 145.8, 128.8, 128.4, 125.3, 117.7, 75.0, 52.9, 52.3, 51.2, 48.6, 47.8, 47.7, 43.5, 38.5, 36.0, 35.6, 35.0, 34.9, 34.0, 33.6, 32.9, 28.2, 27.4, 26.8, 25.4, 24.4, 24.2, 21.9, 21.6, 20.6, 18.3, 18.2, 12.8.



1-(cyclohexylamino)-1-oxooctan-2-yl-3-oxotirucalla-7,24Z-dien-26-oate (8d). Yellow oil; Rf = 0.55 (20% ethyl acetate in hexanes): ¹H NMR (400 MHz, CDCl₃, 25 °C, TMS): δ 7.39 (m, 2H), 7.35 (m, 2H), 6.06 (bs, 1H), 6.03 (dt, *J* = 7.6, 1.6 Hz, 1H), 5.97 (d, *J* = 2.6 Hz, 1H), 5.30 (dd, *J* = 6.0, 3.2 Hz, 1H), 3.83 (dtt, *J* = 10.4, 7.9, 4.1 Hz, 1H), 2.77 (td, *J* = 14.5, 5.4 Hz, 1H), 2.59 (m, 1H), 2.45 (m, 1H), 2.28 (dt, *J* = 14.1, 3.8, 1H), 2.25 (m, 1H), 2.10 (m, 2H), 1.99 (m, 2H), 1.97 (m, 1H), 1.98 (d, J = 1.6 Hz, 3H), 193 (m, 4H), 1.82 (m, 1H), 1.76 (m, 2H), 1.73 (t, *J* = 8.7 Hz, 1H), 169 (m, 2H), 1.64 (m, 1H), 1.56 (m, 2H), 1.53 (m, 2H), 1.49 (m, 1H), 1.48 (m, 2H), 1.48 (m, 1H), 1.47 (m, 2H), 1.40 (m, 1H), 1.36 (s, 9H), 1.31 (m, 2H), 1.29 (m, 2H), 1.28 (m, 1H), 1.26 (m, 2H), 1.18 (m, 4H), 1.16 (m, 1H), 1.11 (s, 3H), 1.05 (s, 3H), 1.01 (s, 3H), 1.00 (s, 3H), 0.89 (d, *J* = 6.2 Hz, 3H), 0.88 (t, *J* = 6.7 Hz, 3H) 0.81 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 216.9, 166.3, 165.8, 146.7, 145.9, 125.3, 117.8, 74.0, 52.9, 52.3, 51.2, 48.5, 47.8, 47.8, 43.4, 38.5, 36.1, 35.5, 35.0, 34.9, 34.0, 33.6, 32.9, 31.7, 31.6, 28.9, 28.2, 27.4, 26.9, 25.4, 24.6, 24.4, 24.5, 24.3, 22.5, 21.9, 21.6, 20.6, 18.4, 18.3, 14.0, 12.8.

4. Conclusions

Herein we developed an efficient and versatile IMCR of Passerini for the functionalization of natural products like triterpenoids. It is important to highlight that alternative solvents can be used for improving the greenness of Passerini reaction. Aliphatic and deactivated aromatic aldehydes lead to the best results in our study, as it was expected. Finally, it is highlighted that the source of the used triterpenoid is sustainable and was obtained without causing harm to the plant and its environment.

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